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PATENT APPLICATION			First Inventor or	First Inventor or Application Identifier Crompton			
TRANSMITTAL			Title M	Title Methods and Compositions for Regulating Lymphocyte Activity			
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	APPLICATION ELEME See MPEP chapter 600 concerning utility patent		nts.	ADDRESS TO:			0 =
1. Specification [Total pages 102]  2. Specification [Total pages 102]  (referred arrangement set forth below)  - Descriptive title of the Invention  - Cross References to Related Applications  - Statement Regarding Fed sponsored R&D  - Reference to Microfiche Appendix  - Background of the Invention  - Brief Summary of the Invention  - Brief Description of the Drawings (if filed)  - Detailed Description  - Claim(s)  - Abstract of the Disclosure  3. Drawing(s) (35 U.S.C. 113) (Total Sheets 4)  Total Sheets 1)  a. Coath or Declaration (Total Sheets 1)  a. Description of Inventor (S)  Signed statement attached deleting inventor(s) named in the prior application see 37 C.F.R. § 1 63 (d)(2) and 1.33(b)				5. Microfiche Computer Program (Appendix)  6. Nucleotide and/or Amino Acid Sequence Submission (if applicable, all necessary)  a. Computer Readable Copy  b. Paper Copy (identical to computer copy)  c. Statement verifying identity of above copies  ACCOMPANYING APPLICATION PARTS  7. Assignment Papers (cover sheet & document(s)  8. (37 C.F.R. § 3.73) Statement Power of Attorney (when there is an assignee)  9. English Translation Document (if applicable)  10. Information Disclosure Copies of IDS Citations Statement (IDS)PTO-1449  11. Preliminary Amendment  12. Return Receipt Postcard (MPEP 503) (Should be specifically itemized)  13. *Small Entity Statement(s) proper and desired (PTO/SB/09-12)\  14. Certified Copy of Priority Documents(s) (if foreign priority is claimed)  15. Other			
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# Methods and Compositions for Regulating Lymphocyte Activity

### **Priority Information**

This application claims priority to U.S. Provisional Application 60/168,112, filed November 30, 1999.

#### **Background of the Invention**

The lymphoid system performs critical functions in animals that include preventing and combating infection, and surveillance and immune elimination of tumor cells. Loss of immune function leads to an immunocompromised status that can predispose the host to serious and life-threatening disease. Functional abnormalities may be present in any of the elements that participate in mediating an immune response, e.g., cellular or humoral elements such as granulocytes, lymphocytes, complement, antibody, or cytokines.

The immune system is a network of cells adapted to protect the organism against pathogens and cells that are not recognized as "self." Once the immune system is activated, it enlists the participation of a variety of cells and molecules to mount an effector function designed to eliminate the "non-self" entity within the body. Lymphocytes are cells of the immune system that are capable of specifically recognizing and selectively eliminating foreign entities. By contrast to other cells of the immune system, such as neutrophils which are considered non-specific in their reactions to invaders, the characteristics of lymphocytes confer specificity, diversity, memory and self/nonself recognition to the immune response.

There are two major populations of lymphocytes: B lymphocytes and T lymphocytes. B lymphocytes originate and mature within the bone marrow and are responsible for formation of antibody molecules. T lymphocytes also arise from the bone marrow but mature in the thymus. There are two major subpopulations of T-cells: T helper cells and T cytotoxic cells. The two types of T cells can be distinguished by the presence of one of two membrane glycoproteins, either CD4 or CD8. The T-helper cells (which express CD4) when activated by antigencomplexes (foreign molecules coupled to special proteins) respond by secreting various growth factors known collectively as cytokines. These cytokines are signals that activate other cells of the immune system, including the T-cytotoxic cells. The T-cytotoxic cells (which express CD8) when activated, proliferate and differentiate into cytotoxic T lymphocytes (CTL) which are able to monitor for and eliminate from the body pathogenic cells, foreign cells, virusinfected cells, and tumor cells.

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The normal development, maturation and differentiation of T lymphocytes are regulated by various peptide factors

# 5 Summary of the Invention

The present invention provides methods and compositions for modulating the immune function of an animal by administering an agent which mimics, or alternatively inhibits, hedgehog-mediated signal transduction. Thus, the subject method can be used as an immunosuppressive, e.g., by administering a hedgehog agonist, or as an immunostimulatant, e.g., by administering a hedgehog antagonist

Wherein the subject method is carried out using a *hedgehog* therapeutic, the *hedgehog* therapeutic preferably a polypeptide including a *hedgehog* portion comprising at least a bioactive extracellular portion of a *hedgehog* protein, e.g., the *hedgehog* portion includes at least 50, 100 or 150 (contiguous) amino acid residues of an N-terminal half of a *hedgehog* protein. In preferred embodiments, the *hedgehog* portion includes at least a portion of the *hedgehog* protein corresponding to a 19kd fragment of the extracellular domain of a *hedgehog* protein.

In preferred embodiments, the *hedgehog* portion has an amino acid sequence at least 60, 75, 85, or 95 percent identical with a hedgehog protein of any of SEQ ID Nos. 10-18 or 20, though sequences identical to those sequence listing entries are also contemplated as useful in the present method. The *hedgehog* portion can be encoded by a nucleic acid which hybridizes under stringent conditions to a nucleic acid sequence of any of SEQ ID Nos. 1-9 or 19, e.g., the *hedgehog* portion can be encoded by a vertebrate *hedgehog* gene, especially a human *hedgehog* gene.

In other embodiments, the subject method can be carried out by administering a gene activation construct, wherein the gene activation construct is deigned to recombine with a genomic *hedgehog* gene of the patient to provide a heterologous transcriptional regulatory sequence operatively linked to a coding sequence of the *hedgehog* gene.

In still other embodiments, the subject method can be practiced with the administration of a gene therapy construct encoding a *hedgehog* polypeptide. For instance, the gene therapy construct can be provided in a composition selected from a group consisting of a recombinant viral particle, a liposome, and a poly-cationic nucleic acid binding agent,

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In yet other embodiments, the subject method can be carried out using a ptc therapeutic. An exemplary ptc therapeutic is a small organic molecule which binds to a patched protein and derepresses patched-mediated inhibition of mitosis, e.g., a molecule which binds to patched and mimics hedgehog-mediated patched signal transduction, which binds to patched and regulates patched-dependent gene expression. For instance, the binding of the ptc therapeutic to patched may result in upregulation of patched and/or gli expression.

In a more generic sense, the *ptc* therapeutic can be a small organic molecule which interacts with MK cells to induce *hedgehog*-mediated *patched* signal transduction, such as by altering the localization, protein-protein binding and/or enzymatic activity of an intracellular protein involved in a *patched* signal pathway. For instance, the *ptc* therapeutic may alter the level of expression of a *hedgehog* protein, a patched protein or a protein involved in the intracellular signal transduction pathway of *patched*.

In certain embodiments, the *ptc* therapeutic is an antisense construct which inhibits the expression of a protein which is involved in the signal transduction pathway of *patched* and the expression of which antagonizes *hedgehog*-mediated signals. The antisense construct is perferably an oligonucleotide of about 20-30 nucleotides in length and having a GC content of at least 50 percent.

In other embodiments, the *ptc* therapeutic is an inhibitor of protein kinase A (PKA), such as a 5-isoquinolinesulfonamide. The PKA inhibitor can be a cyclic AMP analog. Exemplary PKA inhibitors include N-[2-((p-bromocinnamyl)amino)ethyl]-5-isoquinolinesulfonamide, 1-(5-isoquinoline-sulfonyl)-2-methylpiperazine, KT5720, 8-bromo-cAMP, dibutyryl-cAMP and PKA Heat Stable Inhibitor isoform  $\alpha$ . Another exemplary PKA inhibitor is represented in the general formula:

25 wherein,

R<sub>1</sub> and R<sub>2</sub> each can independently represent hydrogen, and as valence and stability permit a lower alkyl, a lower alkenyl, a lower alkynyl, a carbonyl (such as a carboxyl, an ester, a formate, or a ketone), a thiocarbonyl (such as a thiocster, a thioacetate, or a thioformate), an

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amino, an acylamino, an amido, a cyano, a nitro, an azido, a sulfate, a sulfonate, a sulfonamido, -(CH<sub>2</sub>)<sub>m</sub>-R<sub>8</sub>, -(CH<sub>2</sub>)<sub>m</sub>-O-lower alkyl, -(CH<sub>2</sub>)<sub>m</sub>-O-lower alkenyl, -(CH<sub>2</sub>)<sub>m</sub>-O-lower alkenyl, -(CH<sub>2</sub>)<sub>m</sub>-S-lower alkenyl, -(CH<sub>2</sub>)<sub>m</sub>-S-(CH<sub>2</sub>)<sub>m</sub>-R<sub>8</sub>, or

R<sub>1</sub> and R<sub>2</sub> taken together with N form a heterocycle (substituted or unsubstituted);

 $R_3$  is absent or represents one or more substitutions to the isoquinoline ring such as a lower alkyl, a lower alkynyl, a carbonyl (such as a carboxyl, an ester, a formate, or a ketone), a thiocarbonyl (such as a thioester, a thioacetate, or a thioformate), an amino, an acylamino, an amido, a cyano, a nitro, an azido, a sulfate, a sulfonate, a sulfonamido,  $-(CH_2)_m$ - $R_8$ ,  $-(CH_2)_m$ -O-lower alkyl,  $-(CH_2)_m$ -O-lower alkenyl,  $-(CH_2)_m$ -O-( $CH_2)_m$ - $R_8$ ,  $-(CH_2)_m$ -S-lower alkyl,  $-(CH_2)_m$ -S-lower alkenyl,  $-(CH_2)_n$ -S-( $CH_2)_m$ - $R_8$ ;

 $R_8$  represents a substituted or unsubstituted aryl, aralkyl, cycloalkyl, cycloalkenyl, or heterocycle; and

n and m are independently for each occurrence zero or an integer in the range of 1 to 6.

### **Brief Description of the Figures**

Figures 1-4 illustrate the profile for staining with various markers for T-cell maturity.

# **Detailed Description of the Invention**

I. Overview

The present application is directed to the discovery that hedgehog gene products are able to regulate maturation of T lymphocytes. Certain aspects of the invention are directed to preparations of hedgehog polypeptides, or other molecules which regulate *patched* or *smoothened* signalling, and their uses as immunomodulatory agents against both acquired and hereditary immunological disorders.

For instance, such compositions can be used to increase the population of T-helper cells to optimum levels in the host, e.g, to stimulate the immune system of the animal. Such uses of the subject compositions can be used in the treatment of bacterial or viral infections, as well as to help the body fight against cancer cells. Alternatively, these substances also enable the host to adjust to diseases arising from disarrangement of self-recognition processes in which there is excessive attack by host T-cells against endogenous tissues. In such instances, the subject

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cells in a whole animal (in vivo).

compositions can be used to reduce T-cell population so that the signs and symptoms of self-directed inflammatory (autoimmune) diseases such rheumatoid arthritis and multiple sclerosis are ameliorated.

As described herein, *hedgehog* proteins inhibit maturation of T lymphocytes. Based upon its inhibitory effect, the administration of hedgehog or ptc therapeutics (defined <u>infra</u>) is suggested herein as a treatment for several types of immunological disorders involving unwanted activation of cellular immunity, e.g., graft rejection, autoimmune disorders, and the like. In other embodiments, inhibitors of hedgehog signalling pathways can be used as immunostimulatory agents, e.g., to counteract the effects of any endogenous activation of a hedgehog/ptc pathway.

In general, the method of the present invention comprises administering to animal, or to cultured lymphocytes in vitro, an amount of a hedgehog or ptc therapeutic which produces a non-toxic response by the cell of inhibition of maturation (in the case of a hedgehog agonist) or promotion of T cell maturation (in the case of a hedgehog antagonist). The subject method can be carried out on cells which may be either dispersed in culture or a part of an intact tissue or organ. Moreover, the method can be performed on cells which are provided in culture (*in vitro*), or on

In one aspect, the present invention provides pharmaceutical preparations and methods for treating or preventing hyperimmune or hypoimmune disorders utilizing, as an active ingredient, a *hedgehog* polypeptide or a mimetic thereof. The invention also relates to methods of controlling the functional performance of T cells by use of the pharmaceutical preparations of the invention.

The subject *hedgehog* treatments are effective on both human and animal subjects afflicted with these conditions. Animal subjects to which the invention is applicable extend to both domestic animals and livestock, raised either as pets or for commercial purposes. Examples are dogs, cats, cattle, horses, sheep, hogs and goats.

Without wishing to be bound by any particular theory, the inhibitory effect of *hedgehog* on T cell maturation may be due at least in part to the ability of hedgehog proteins to antagonize (directly or indirectly) *patched*-mediated regulation of gene expression and other physiological effects mediated by that protein. The *patched* gene product, a cell surface protein, is understood to signal through a pathway which causes transcriptional repression of members of the Wnt and Dpp/BMP families of morphogens, proteins which impart positional information. In other tissue, the introduction of *hedgehog* relieves (derepresses) this inhibition conferred by *patched*, allowing expression of particular gene programs.

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Recently, it has been reported that mutations in the human version of patched, a gene first identified in a fruit fly developmental pathway, cause a hereditary skin cancer and may contribute to sporadic skin cancers. See, for example, Hahn et al. (1996) Cell 86:841-851; and Johnson et al. (1996) Science 272:1668-1671. The demonstraction that nevoid basal-cell carcinoma (NBCC) results from mutations in the human patched gene provided an example of the roles patched plays in post-embryonic deveolpment. These observations have led the art to understand one activity of patched to be a tumor suppressor gene, which may act by inhibiting proliferative signals from hedgehog. Our observations set forth below reveal potential new roles for the hedgehog/patched pathway in maintenance of mature T-cells and other lyphocytic cell lines. Accordingly, the present invention contemplates the use of other agents which are capable of mimicking or antagonizing, depending on the desired consequence, the effect of the hedgehog protein on patched signalling, e.g., as may be identified from the drug screening assays described below.

#### II. Definitions

For convience, certain terms employed in the specification, examples, and appended claims are collected here.

The term "hedgehog therapeutic" refers to various forms of hedgehog polypeptides, as well as peptidomimetics, which can modulate the proliferation/differentiation state of lymphocytes, by, as will be clear from the context of individual examples, mimicing or potentiating (agonizing) or inhibiting (antagonizing) the effects of a naturally-occurring hedgehog protein. A hedgehog therapeutic which mimics or potentiates the activity of a wild-type hedgehog protein is a "hedgehog agonist". Conversely, a hedgehog therapeutic which inhibits the activity of a wild-type hedgehog protein is a "hedgehog antagonist".

In particular, the term "hedgehog polypeptide" encompasses preparations of *hedgehog* proteins and peptidyl fragments thereof, both agonist and antagonist forms as the specific context will make clear.

As used herein the term "bioactive fragment of a *hedgehog* protein" refers to a fragment of a full-length *hedgehog* polypeptide, wherein the fragment specifically agonizes or antagonizes inductive events mediated by wild-type *hedgehog* proteins. The *hedgehog* biactive fragment preferably is a soluble extracellular portion of a *hedgehog* protein, where solubility is with reference to physiologically compatible solutions. Exemplary bioactive fragments are described in PCT publications WO 95/18856 and WO 96/17924.

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The term "ptc therapeutic" refers to agents which either (i) mimic the effect of *hedgehog* proteins on *patched* signalling, e.g., which antagonize the cell-cycle inhibitory and/or T-cell maturation promoting activity of *patched*, or (ii) activate or potentiate patched signalling. In other embodiments, the ptc therapeutic can be a *hedgehog* antagonist. The ptc therapeutic can be, e.g., a peptide, a nucleic acid, a carbohydrate, a small organic molecule, or natural product extract (or fraction thereof).

An "effective amount" of, e.g., a hedgehog or ptc therapeutic, with respect to the subject method of treatment, refers to an amount of a composition, e.g., a hedgehog polypeptide, in a preparation which, when applied as part of a desired dosage regimen brings enhances or inhibits (as the case may be) T-cell maturation, relative to the absence of the hedgehog or ptc therapeutic, according to clinically acceptable standards for the disorder to be treated.

A "patient" or "subject" to be treated by the subject method can mean either a human or non-human animal.

The "growth state" of a cell refers to the rate of proliferation of the cell and the state of differentiation of the cell.

"Homology" and "identity" each refer to sequence similarity between two polypeptide sequences, with identity being a more strict comparison. Homology and identity can each be determined by comparing a position in each sequence which may be aligned for purposes of comparison. When a position in the compared sequence is occupied by the same amino acid residue, then the polypeptides can be referred to as identical at that position; when the equivalent site is occupied by the same amino acid (e.g., identical) or a similar amino acid (e.g., similar in steric and/or electronic nature), then the molecules can be referred to as homologous at that position. A percentage of homology or identity between sequences is a function of the number of matching or homologous positions shared by the sequences. An "unrelated" or "non-homologous" sequence shares less than 40 percent identity, though preferably less than 25 percent identity, with an hedgeog sequence of the present invention.

The term "corresponds to", when referring to a particular polypeptide or nucleic acid sequence is meant to indicate that the sequence of interest is identical or homologous to the reference sequence to which it is said to correspond.

The terms "recombinant protein", "heterologous protein" and "exogenous protein" are used interchangeably throughout the specification and refer to a polypeptide which is produced by recombinant DNA techniques, wherein generally, DNA encoding the polypeptide is inserted

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into a suitable expression construct which is in turn used to transform a host cell to produce the heterologous protein. That is, the polypeptide is expressed from a heterologous nucleic acid.

A "chimeric protein" or "fusion protein" is a fusion of a first amino acid sequence encoding a *hedgehog* polypeptide with a second amino acid sequence defining a domain foreign to and not substantially homologous with any domain of hh protein. A chimeric protein may present a foreign domain which is found (albeit in a different protein) in an organism which also expresses the first protein, or it may be an "interspecies", "intergenic", etc. fusion of protein structures expressed by different kinds of organisms. In general, a fusion protein can be represented by the general formula  $(X)_n$ - $(hh)_m$ - $(Y)_n$ , wherein hh represents all or a portion of the *hedgehog* protein, X and Y each independently represent an amino acid sequences which are not naturally found as a polypeptide chain contiguous with the *hedgehog* sequence, m is an integer greater than or equal to 1, and each occurrence of n is, independently, 0 or an integer greater than or equal to 1 (n and m are preferably no greater than 5 or 10).

The term "CD4<sup>+</sup> lymphocyte" is intended to mean a lymphocyte having a plurality of cell surface CD4 molecules as evidenced by binding of an antibody specific for CD4 to the cell surface, e.g., monoclonal antibody OKT4 (Ortho Diagnostics, Piscataway, N.J.).

The term "CD8<sup>+</sup> lymphocyte" is intended to mean a lymphocyte having a plurality of cell surface CD8 molecules as evidenced by binding of an antibody specific for CD8 to the cell surface, e.g., monoclonal antibody OKT8 (Ortho Diagnostics, Piscataway, N.J.).

"Peripheral blood leukocytes", abbreviated PBL, are intended to mean the cellular components of the immune system in blood, e.g., lymphocytes, monocytes, eosinophils, basophils, neutrophils, plasma cells, mast cell precursors, and the like.

"Lymphocytes" are intended to encompass T-lymphocytes, (also referred to as T-cells), B-lymphocytes (also referred to as B-cells), natural killer-lymphocytes (NK-cells), cytotoxic T lymphocytes (CTL), T-helper lymphocytes, delta gamma -T-cell receptor bearing cells as well as precursors and activated derivatives thereof

"Activated lymphocyte" is intended to mean that subset of lymphocytes which has been i) exposed to a stimulus, and ii) has been triggered to change from a metabolically-quiescent cell into a cell with increased protein synthesis and/or DNA and RNA synthesis and possibly cell division. Illustrative stimuli and triggering pathways leading to activated lymphocytes include interaction of T cell receptors with antigens, interaction of interleukin receptors with interleukins, interaction of growth factor receptors with growth factors, interaction of lymphocyte cell surface determinants with antibody (e.g., anti-CD2) or mitogens (e.g., PHA), and the like.

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"Subject in need thereof" is intended to mean a mammal having one or more clinical or laboratory indicia of a disorder or disease. The subject may exhibit clinical disease activity or may have a subclinical or latent infection. Subjects in need thereof include human and nonhuman primates, mammals, domestic animals, livestock, and the like, e.g., dogs, cats, rodents, birds, horses, cows, pigs and fish.

### III. Exemplary Applications of Method and Compositions

According to the invention, preparations with ptc and hedgehog therapeutics are used in immunoregulatory disorders and diseases in animals and humans, for the prevention or prophylaxis, control, diagnosis or treatment thereof.

The subject method has wide applicability to the treatment or prophylaxis of disorders affecting the regulation of lymphocytes, particularly maturation and/or activation of T lymphocytes. In general, the method can be characterized as including a step of administering to an animal an amount of a ptc or hedgehog therapeutic effective to alter the proliferative and/or differentiation state of treated lymphocytes. Such therapeutic compositions may be useful in treatments designed to modulate, e.g., increase or decrease, an immunological response. Such diseases and conditions include, but are not limited to, infection (such as bacterial or viral infection), metabolic disease such as diabetes, nutritional deficiency, toxic agents, graft rejection or other hyperacute response, or autoimmune disorders. The goals of treatment in each case can be twofold: (1) to eliminate the cause of the disease or unwanted immunological response, and (2) to relieve its symptoms.

In view of their immunosuppressant activity, the hedgehog proteins (and agonists thereof) are suitable for preventing and treating diseases and conditions which require a temporary or permanent reduction or suppression of an immune response. In particular, their use extends to suppressing the activation of the proliferation of lymphocytes or cytotoxic T-cells and/or immunocytes, e.g. for preventing or treating autoimmune diseases such as diseases of the rheumatic type, multiple sclerosis, psoriasis, atopic dermatitis, or for preventing the rejection of transplanted tissues or organs such as kidneys, heart, lungs, bone marrow, spleen, skin or cornea, in undesirable reactions during or after transfusions, allergic diseases, particularly those which affect the gastrointestinal tract and which may take the form of an inflammation, or inflammatory, proliferative and hyperproliferative diseases and cutaneous manifestations of immunological disorders such as eczematous dermatitis, urticaria, vasculitis and scleroderma.

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Thus, it is particularly advantageous to use immunosuppressive forms of the subject hedgehog nd ptc therapeutics clinically for the disorders, diseases and conditions described above, i.e. when it is desirable to achieve immunosuppression in an animal or human body.

Depending on the nature and cause of the disease or disorder to be treated or the condition which is to be influenced in an animal or human body, it may be desirable to administer the hedgehog or ptc therapeutic preparation systemically, locally or topically to the tissue or organ in question. Systemic action is desirable, for example, when various organs or organ systems are in need of treatment, as is the case for example in systemic autoimmune diseases or allergies or in transplants of large, foreign organs or tissues. By contrast, a local effect would be considered if only local manifestations of an immunological occurrence had to be treated, e.g. in small transplants of skin or cornea or in cases of local dermatitis.

Depending on the duration and intensity of the immunosuppressant activity required, the hedgehog or ptc therapeutic preparations may be given one or more times a day, as well as intermittently, over a period of several days, weeks or months and in various dosages.

In still other embodiments, antagonist (lymphocyte maturation promoting) forms of the subject ptc and hedgehog therapeutics can be used to treat disorders involving hypoimmunity, e.g., immunosuppressed or immunocompromised patients. For instance, the effect of hedgehog-induced immunosuppression, e.g., resulting from endogenous or heterologous activation of the hedgehog signalling pathway, can be counteracted using antagonist forms of the subject ptc and hedgehog therapeutics.

Thus, the subject method contemplates the treatment of immunocompromised subjects to increase one or more indicia of cell mediated immunity (CMI), humoral immunity, or innate resistance to infection, by administering pharmaceutical preparations of an activating ptc or hedgehog therapeutic. In certain embodiments, such immunity-promoting activities of antagonists forms of the subject ptc and hedgehog therapeutics can be identified, e.g., by i) increased E-rosette forming cells (E-RFC) in thymocyte cultures after incubation with the subject ptc or hedgehog therapeutic agents; ii) increased E-RFC in cultures of thymocytes from aged animals after incubation with the subject ptc or hedgehog therapeutic agents; and, iii) increased expression of OKT 4< + > in cultures of human peripheral blood T-lymphocytes from patients with secondary immunodeficiency syndromes following treatment with the subject ptc or hedgehog therapeutic agents. Increased expression of CD2 and CD4 accessory molecules on T-lymphocytes is compatible with a heighten the state of innate or induced immunity to infection, e.g., by upregulating T-helper and cytotoxic T-lymphocytes to respond to lower levels of antigen.

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Immunodeficiency states fall into three general etiologic categories. First, there is immunosuppression that occurs as a consequence of disease processes. Second, there are immunodeficiencies that arise because of therapy for other diseases, so-called iatrogenic immunodeficiencies. Third, immunodeficiencies may result from direct attack of T-lymphocytes by the human immunodeficiency virus (HIV) that causes the acquired immunodeficiency syndrome (AIDS).

Common disease processes that lead to immunodeficiency are malnutrition, neoplasias, aging, and infections. Malnourished people, patients with advanced widespread cancers and people with debilitating illnesses become sick and die more often because impaired cell-mediated and humoral immune responses increase susceptibility to infections by a variety of organisms. A state of generalized deficiency in immune responses is called anergy. Various types of infections, especially viral infections, lead to immunosuppression. A drug, such as a maturation-pormoting form of a Ptc or hedgehog therapeutic, e.g., capable of making the T-helper lymphocyte components of the immune system more robust, will be an important therapeutic agent for increasing the resistance of the patient to infections. For example, Ptc or hedgehog therapeutic or its analogs, may be:

administered to patients, especially older patients, before or just after admissions to hospitals in order to reduce the risks of nosocomial (hospital-induced) infections, a common and severe clinical problem

administered to burn victims, because such individuals are especially prone to infections

administered to patients in anticipation of epidemic infections, for example, in conjunction with influenza vaccinations or hepatitis vaccinations, to invigorate the immune response to pathogens

administered to patients with asymptomatic viral infections, in order to enhance immune surveillance of pathogenic organisms and reduce the likelihood of recurrence of disease, for example, for individuals who are carriers of herpes viruses, varicella viruses, hepatitis viruses and HIV.

Iatrogenic immunosuppression is most often due to drug therapies which either kill or functionally inactivate lymphocytes. Various chemotherapeutic drugs are administered to cancer patients, and these drugs are usually cytotoxic to both mature and developing lymphocytes as well as to granulocyte and monocyte precursors. Thus, cancer chemotherapy is almost always accompanied by a period of immunosuppression and increased risk of infections. Radiation treatment of cancer carries the same risks. Medications (granulocyte-colony stimulating factor)

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exist for increasing neutrophils in blood to combat infections that occur after cancer chemotherapy, but no medications are currently used for restoring lymphocytic functions. Major surgery, for example repair of aneurysms or by-pass operations, also decrease immune function in humans. The reasons for the decline in blood lymphocytes that occur because of major surgery are not clear, but an agent that elevates lymphocyte functions in such patients have therapeutic value in decreasing the likelihood of infections.

One final form of acquired immunosuppression that should be mentioned results from the absence of a spleen, caused by surgical removal of the organ after trauma or for the treatment of certain hematologic diseases or as a result of infarction in sickle cell disease. Patients without spleens are more susceptible to infections by some organisms, particularly encapsulated bacteria such as Streptococcus pneumoniae. The spleen is apparently required for the induction of protective humoral immune responses to such organisms. The subject Ptc or hedgehog therapeutics can help individuals without a spleen or without a thymus in resistance against infection by micro-organisms.

## IV. Exemplary hedgehog therapeutic compounds.

The *hedgehog* therapeutic compositions of the subject method can be generated by any of a variety of techniques, including purification of naturally occurring proteins, recombinantly produced proteins and synthetic chemistry. Polypeptide forms of the hedgehog therapeutics are preferably derived from vertebrate hedgehog proteins, e.g., have sequences corresponding to naturally occurring hedgehog proteins, or fragments thereof, from vertebrate organisms. However, it will be appreciated that the hedgehog polypeptide can correspond to a hedgehog protein (or fragment thereof) which occurs in any metazoan organism.

The various naturally-occurring *hedgehog* proteins from which the subject therapeutics can be derived are characterized by a signal peptide, a highly conserved N-terminal region, and a more divergent C-terminal domain. In addition to signal sequence cleavage in the secretory pathway (Lee, J.J. *et al.* (1992) *Cell* 71:33-50; Tabata, T. *et al.* (1992) *Genes Dev.* 2635-2645; Chang, D.E. *et al.* (1994) *Development* 120:3339-3353), *hedgehog* precursor proteins naturally undergo an internal autoproteolytic cleavage which depends on conserved sequences in the C-terminal portion (Lee *et al.* (1994) *Science* 266:1528-1537; Porter *et al.* (1995) *Nature* 374:363-366). This autocleavage leads to a 19 kD N-terminal peptide and a C-terminal peptide of 26-28 kD (Lee *et al.* (1992) *supra*; Tabata *et al.* (1992) *supra*; Chang *et al.* (1994) *supra*; Lee *et al.* (1994) *supra*; Bumcrot, D.A., *et al.* (1995) *Mol. Cell. Biol.* 15:2294-2303; Porter *et al.* (1995) *supra*; Ekker, S.C. *et al.* (1995) *Curr. Biol.* 5:944-955; Lai, C.J. *et al.* (1995) *Development* 

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121:2349-2360). The N-terminal peptide stays tightly associated with the surface of cells in which it was synthesized, while the C-terminal peptide is freely diffusible both *in vitro* and *in vivo* (Lee *et al.* (1994) *supra*; Bumcrot *et al.* (1995) *supra*; Mart', E. *et al.* (1995) *Development* 121:2537-2547; Roelink, H. *et al.* (1995) *Cell* 81:445-455). Cell surface retention of the N-terminal peptide is dependent on autocleavage, as a truncated form of *hedgehog* encoded by an RNA which terminates precisely at the normal position of internal cleavage is diffusible *in vitro* (Porter *et al.* (1995) *supra*) and *in vivo* (Porter, J.A. *et al.* (1996) *Cell* 86, 21-34). Biochemical studies have shown that the autoproteolytic cleavage of the *hedgehog* precursor protein proceeds through an internal thioester intermediate which subsequently is cleaved in a nucleophilic substitution. It is suggested that the nucleophile is a small lipophilic molecule, more particularly cholesterol, which becomes covalently bound to the C-terminal end of the N-peptide (Porter *et al.* (1996) *supra*), tethering it to the cell surface.

The vertebrate family of *hedgehog* genes includes at least four members, e.g., paralogs of the single drosophila *hedgehog* gene (SEQ ID No. 19). Three of these members, herein referred to as Desert *hedgehog* (*Dhh*), Sonic *hedgehog* (*Shh*) and Indian *hedgehog* (*Ihh*), apparently exist in all vertebrates, including fish, birds, and mammals. A fourth member, herein referred to as tiggie-winkle *hedgehog* (*Thh*), appears specific to fish. According to the appended sequence listing, (see also Table 1) a chicken *Shh* polypeptide is encoded by SEQ ID No:1; a mouse *Dhh* polypeptide is encoded by SEQ ID No:2; a mouse *Ihh* polypeptide is encoded by SEQ ID No:3; a mouse *Shh* polypeptide is encoded by SEQ ID No:6; a human *Ihh* polypeptide is encoded by SEQ ID No:6; a human *Ihh* polypeptide is encoded by SEQ ID No:7; a human *Dhh* polypeptide is encoded by SEQ ID No. 8; and a zebrafish *Thh* is encoded by SEQ ID No. 9.

Table 1
Guide to hedgehog sequences in Sequence Listing

	Nucleotide	Amino Acid
Chicken Shh	SEQ ID No. 1	SEQ ID No. 10
Mouse Dhh	SEQ ID No. 2	SEQ ID No. 11
Mouse Ihh	SEQ ID No. 3	SEQ ID No. 12
Mouse Shh	SEQ ID No. 4	SEQ ID No. 13
Zebrafish Shh	SEQ ID No. 5	SEQ ID No. 14
Human Shh	SEQ ID No. 6	SEQ ID No. 15
Human Ihh	SEQ ID No. 7	SEQ ID No. 16
Human Dhh	SEQ ID No. 8	SEQ ID No. 17
Zebrafish Thh	SEQ ID No. 9	SEQ ID No. 18
Drosophila HH	SEQ ID No. 19	SEQ ID No. 20

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In addition to the sequence variation between the various *hedgehog* homologs, the *hedgehog* proteins are apparently present naturally in a number of different forms, including a pro-form, a full-length mature form, and several processed fragments thereof. The pro-form includes an N-terminal signal peptide for directed secretion of the extracellular domain, while the full-length mature form lacks this signal sequence.

As described above, further processing of the mature form occurs in some instances to yield biologically active fragments of the protein. For instance, *sonic hedgehog* undergoes additional proteolytic processing to yield two peptides of approximately 19 kDa and 27 kDa, the 19kDa fragment corresponding to an proteolytic N-terminal portion of the mature protein.

In addition to proteolytic fragmentation, the vertebrate *hedgehog* proteins can also be modified post-translationally, such as by glycosylation and/or addition of lipophilic moieties, such as stents, fatty acids, etc., though bacterially produced (e.g. unmodified) forms of the proteins still maintain certain of the bioactivities of the native protein. Bioactive fragments of *hedgehog* polypeptides of the present invention have been generated and are described in great detail in, e.g., PCT publications WO 95/18856 and WO 96/17924.

There are a wide range of lipophilic moieties with which hedgehog polypeptides can be derivatived. The term "lipophilic group", in the context of being attached to a hedgehog polypeptide, refers to a group having high hydrocarbon content thereby giving the group high affinity to lipid phases. A lipophilic group can be, for example, a relatively long chain alkyl or cycloalkyl (preferably n-alkyl) group having approximately 7 to 30 carbons. The alkyl group may terminate with a hydroxy or primary amine "tail". To further illustrate, lipophilic molecules include naturally-occurring and synthetic aromatic and non-aromatic moieties such as fatty acids, sterols, esters and alcohols, other lipid molecules, cage structures such as adamantane and buckminsterfullerenes, and aromatic hydrocarbons such as benzene, perylene, phenanthrene, anthracene, naphthalene, pyrene, chrysene, and naphthacene.

In one embodiment, the hedgehog polypeptide is modified with one or more sterol moieties, such as cholesterol. See, for example, PCT publication WO 96/17924. In certain embodiments, the cholesterol is preferably added to the C-terminal glycine were the hedgehog polypeptide corresponds to the naturally-occurring N-terminal proteolytic fragment.

In another embodiment, the hedgehog polypeptide can be modified with a fatty acid moiety, such as a myrostoyl, palmitoyl, stearoyl, or arachidoyl moiety. See, e.g., Pepinsky et al. (1998) J Biol. Chem 273: 14037.

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In addition to those effects seen by cholesterol-addition to the C-terminus or fatty acid addition to the N-terminus of extracellular fragments of the protein, at least certain of the biological activities of the hedgehog gene products are unexpectedly potentiated by derivativation of the protein with lipophilic moieties at other sites on the protein and/or by moieties other than cholesterol or fatty acids. Certain aspects of the invention are directed to the use of preparations of hedgehog polypeptides which are modified at sites other than N-terminal or C-terminal residues of the natural processed form of the protein, and/or which are modified at such terminal residues with lipophilic moieties other than a sterol at the C-terminus or fatty acid at the N-terminus.

Particularly useful as lipophilic molecules are alicyclic hydrocarbons, saturated and unsaturated fatty acids and other lipid and phospholipid moieties, waxes, cholesterol, isoprenoids, terpenes and polyalicyclic hydrocarbons including adamantane buckminsterfullerenes, vitamins, polyethylene glycol or oligoethylene glycol, (C1-C18)-alkyl phosphate diesters, -O-CH2-CH(OH)-O-(C12-C18)-alkyl, and in particular conjugates with pyrene derivatives. The lipophilic moiety can be a lipophilic dye suitable for use in the invention include, but are not limited to, diphenylhexatriene, Nile Red, N-phenyl-1-naphthylamine, Prodan, Laurodan, Pyrene, Perylene, rhodamine, rhodamine B, tetramethylrhodamine, Texas Red, sulforhodamine. 1.1'-didodecyl-3.3.3',3'tetramethylindocarbocyanine perchlorate, rhodamine B and the BODIPY dyes available from Molecular Probes Inc.

Other exemplary lipophilic moietites include aliphatic carbonyl radical groups include 1-or 2-adamantylacetyl, 3-methyladamant-1-ylacetyl, 3-methyl-3-bromo-1-adamantylacetyl, 1-decalinacetyl, camphoracetyl, camphaneacetyl, noradamantylacetyl, norbornaneacetyl, bicyclo[2.2.2.]-oct-5-eneacetyl, 1-methoxybicyclo[2.2.2.]-oct-5-ene-2-carbonyl, cis-5-norbornene-endo-2,3-dicarbonyl, 5-norbornen-2-ylacetyl, (1R)-( - )-myrtentaneacetyl, 2-norbornaneacetyl, anti-3-oxo-tricyclo[2.2.1.0<2,6> ]-heptane-7-carbonyl, decanoyl, dodecanoyl, dodecanoyl, tetradecadienoyl, decynoyl or dodecynoyl.

The hedgehog polypeptide can be linked to the hydrophobic moiety in a number of ways including by chemical coupling means, or by genetic engineering.

There are a large number of chemical cross-linking agents that are known to those skilled in the art. For the present invention, the preferred cross-linking agents are heterobifunctional cross-linkers, which can be used to link the hedgehog polypeptide and hydrophobic moiety in a stepwise manner. Heterobifunctional cross-linkers provide the ability to design more specific coupling methods for conjugating to proteins, thereby reducing the occurrences of unwanted side reactions such as homo-protein polymers. A wide variety of heterobifunctional cross-linkers are

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known in the art. These include: succinimidyl 4-(N-maleimidomethyl) cyclohexane- 1-carboxylate (SMCC), m-Maleimidobenzoyl-N- hydroxysuccinimide ester (MBS); N-succinimidyl (4-iodoacetyl) aminobenzoate (SIAB), succinimidyl 4-(p-maleimidophenyl) butyrate (SMPB), 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride (EDC); 4-succinimidyloxycarbonyl- a-methyl-a-(2-pyridyldithio)-tolune (SMPT), N-succinimidyl 3-(2-pyridyldithio) propionate (SPDP), succinimidyl 6-[3-(2-pyridyldithio) propionate] hexanoate (LC-SPDP). Those cross-linking agents having N-hydroxysuccinimide moieties can be obtained as the N-hydroxysulfosuccinimide analogs, which generally have greater water solubility. In addition, those cross-linking agents having disulfide bridges within the linking chain can be synthesized instead as the alkyl derivatives so as to reduce the amount of linker cleavage *in vivo*.

In addition to the heterobifunctional cross-linkers, there exists a number of other crosslinking agents including homobifunctional and photoreactive cross-linkers. Disuccinimidyl suberate (DSS), bismaleimidohexane (BMH) and dimethylpimelimidate-2 HCl (DMP) are homobifunctional cross-linking and bis-[ß-(4examples of useful agents, and N-succinimidyl-6(4'-azido-2'-nitrophenylazidosalicvlamido)ethylldisulfide (BASED) amino)hexanoate (SANPAH) are examples of useful photoreactive cross-linkers for use in this For a recent review of protein coupling techniques, see Means et al. (1990) Bioconjugate Chemistry 1:2-12, incorporated by reference herein.

One particularly useful class of heterobifunctional cross-linkers, included above, contain the primary amine reactive group, N-hydroxysuccinimide (NHS), or its water soluble analog N-hydroxysulfosuccinimide (sulfo-NHS). Primary amines (lysine epsilon groups) at alkaline pH's are unprotonated and react by nucleophilic attack on NHS or sulfo-NHS esters. This reaction results in the formation of an amide bond, and release of NHS or sulfo-NHS as a by-product.

Another reactive group useful as part of a heterobifunctional cross-linker is a thiol reactive group. Common thiol reactive groups include maleimides, halogens, and pyridyl disulfides. Maleimides react specifically with free sulfhydryls (cysteine residues) in minutes, under slightly acidic to neutral (pH 6.5-7.5) conditions. Halogens (iodoacetyl functions) react with -SH groups at physiological pH's. Both of these reactive groups result in the formation of stable thioether bonds.

The third component of the heterobifunctional cross-linker is the spacer arm or bridge. The bridge is the structure that connects the two reactive ends. The most apparent attribute of the bridge is its effect on steric hindrance. In some instances, a longer bridge can more easily span the distance necessary to link two complex biomolecules. For instance, SMPB has a span of 14.5 angstroms.

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Preparing protein-protein conjugates using heterobifunctional reagents is a two-step process involving the amine reaction and the sulfhydryl reaction. For the first step, the amine reaction, the protein chosen should contain a primary amine. This can be lysine epsilon amines or a primary alpha amine found at the N-terminus of most proteins. The protein should not contain free sulfhydryl groups. In cases where both proteins to be conjugated contain free sulfhydryl groups, one protein can be modified so that all sulfhydryls are blocked using for instance, N-ethylmaleimide (see Partis et al. (1983) J. Pro. Chem. 2:263, incorporated by reference herein). Ellman's Reagent can be used to calculate the quantity of sulfhydryls in a particular protein (see for example Ellman et al. (1958) Arch. Biochem. Biophys. 74:443 and Riddles et al. (1979) Anal. Biochem. 94:75, incorporated by reference herein).

The reaction buffer should be free of extraneous amines and sulfhydryls. The pH of the reaction buffer should be 7.0-7.5. This pH range prevents maleimide groups from reacting with amines, preserving the maleimide group for the second reaction with sulfhydryls.

The NHS-ester containing cross-linkers have limited water solubility. They should be dissolved in a minimal amount of organic solvent (DMF or DMSO) before introducing the cross-linker into the reaction mixture. The cross-linker/solvent forms an emulsion which will allow the reaction to occur.

The sulfo-NHS ester analogs are more water soluble, and can be added directly to the reaction buffer. Buffers of high ionic strength should be avoided, as they have a tendency to "salt out" the sulfo-NHS esters. To avoid loss of reactivity due to hydrolysis, the cross-linker is added to the reaction mixture immediately after dissolving the protein solution.

The reactions can be more efficient in concentrated protein solutions. The more alkaline the pH of the reaction mixture, the faster the rate of reaction. The rate of hydrolysis of the NHS and sulfo-NHS esters will also increase with increasing pH. Higher temperatures will increase the reaction rates for both hydrolysis and acylation.

Once the reaction is completed, the first protein is now activated, with a sulfhydryl reactive moiety. The activated protein may be isolated from the reaction mixture by simple gel filtration or dialysis. To carry out the second step of the cross-linking, the sulfhydryl reaction, the lipophilic group chosen for reaction with maleimides, activated halogens, or pyridyl disulfides must contain a free sulfhydryl. Alternatively, a primary amine may be modified with to add a sulfhydryl

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In all cases, the buffer should be degassed to prevent oxidation of sulfhydryl groups. EDTA may be added to chelate any oxidizing metals that may be present in the buffer. Buffers should be free of any sulfhydryl containing compounds.

Maleimides react specifically with -SH groups at slightly acidic to neutral pH ranges (6.5-7.5). A neutral pH is sufficient for reactions involving halogens and pyridyl disulfides. Under these conditions, maleimides generally react with -SH groups within a matter of minutes. Longer reaction times are required for halogens and pyridyl disulfides.

The first sulfhydryl reactive-protein prepared in the amine reaction step is mixed with the sulfhydryl-containing lipophilic group under the appropriate buffer conditions. The conjugates can be isolated from the reaction mixture by methods such as gel filtration or by dialysis.

Exemplary activated lipophilic moieties for conjugation include: N-(1-pyrene)maleimide; fluorescein-5-maleimide; N-(4-(6-2,5-dimethoxystilbene-4'-maleimide, eosin-5-maleimide; dimethylamino-2-benzofuranyl)phenyl)maleimide; benzophenone-4-maleimide; 4dimethylaminophenylazophenyl- 4'-maleimide (DABMI), tetramethylrhodamine-5-maleimide, N-(5-Rhodamine RedTM C2 maleimide, tetramethylrhodamine-6-maleimide, aminopentyl)maleimide, trifluoroacetic acid salt, N-(2-aminoethyl)maleimide, trifluoroacetic acid salt, Oregon GreenTM 488 maleimide, N-(2-((2-(((4-azido- 2,3,5,6-tetrafluoro)benzoyl) amino)ethyl)dithio)ethyl)maleimide (TFPAM-SS1), 2-(1-(3-dimethylaminopropyl) -indol-3-yl)-3-(indol-3-yl) maleimide (bisindolylmaleimide; GF 109203X), BODIPY® FL N-(2aminoethyl)maleimide, N-(7-dimethylamino- 4-methylcoumarin-3-yl)maleimide (DACM), AlexaTM 488 C5 maleimide, AlexaTM 594 C5 maleimide, sodium saltN-(1-pyrene)maleimide, 2,5-dimethoxystilbene-4'-maleimide, eosin-5-maleimide, fluorescein-5-maleimide, N-(4-(6benzophenone-4-maleimide, 4dimethylamino-2-benzofuranyl)phenyl)maleimide, (4dimethylaminophenylazophenyl-4'-maleimide, 1-(2-maleimidylethyl)-4-(5methoxyphenyl)oxazol-2- yl)pyridinium methanesulfonate, tetramethylrhodamine-5-maleimide, maleimide, N-(5tetramethylrhodamine-6-maleimide, Rhodamine RedTM C2 N-(2-((2-(((4-azido-N-(2-aminoethyl)maleimide, 2,3,5,6aminopentyl)maleimide, tetrafluoro)benzoyl) amino)ethyl)dithio)ethyl)maleimide, 2-(1-(3-dimethylaminopropyl) -indol-3vl)-3-(indol-3-vl) maleimide, N-(7-dimethylamino- 4-methylcoumarin-3-yl)maleimide (DACM), 11H-Benzo[a]fluorene, Benzo[a]pyrene.

In one embodiment, the hedgehog polypeptide can be derivatived using pyrene maleimide, which can be purchased from Molecular Probes (Eugene, Oreg.), e.g., N-(1-pyrene)maleimide or 1-pyrenemethyl iodoacetate (PMIA ester).

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For those embodiments wherein the hydophobic moiety is a polypeptide, the modified hedgehog polypeptide of this invention can be constructed as a fusion protein, containing the hedgehog polypeptide and the hydrophobic moiety as one contiguous polypeptide chain.

In certain embodiments, the lipophilic moiety is an amphipathic polypeptide, such as magainin, cecropin, attacin, melittin, gramicidin S, alpha-toxin of Staph. aureus, alamethicin or a synthetic amphipathic polypeptide. Fusogenic coat proteins from viral particles can also be a convenient source of amphipathic sequences for the subject hedgehog proteins

Moreover, mutagenesis can be used to create modified *hh* polypeptides, e.g., for such purposes as enhancing therapeutic or prophylactic efficacy, or stability (e.g., *ex vivo* shelf life and resistance to proteolytic degradation *in vivo*). Such modified peptides can be produced, for instance, by amino acid substitution, deletion, or addition. Modified *hedgehog* polypeptides can also include those with altered post-translational processing relative to a naturally occurring *hedgehog* protein, e.g., altered glycosylation, cholesterolization, prenylation and the like.

In one embodiment, the hedgehog therapeutic is a polypeptide encodable by a nucleotide sequence that hybridizes under stringent conditions to a hedgehog coding sequence represented in one or more of SEQ ID Nos:1-7. Appropriate stringency conditions which promote DNA hybridization, for example, 6.0 x sodium chloride/sodium citrate (SSC) at about 45°C, followed by a wash of 2.0 x SSC at 50°C, are known to those skilled in the art or can be found in *Current Protocols in Molecular Biology*, John Wiley & Sons, N.Y. (1989), 6.3.1-6.3.6. For example, the salt concentration in the wash step can be selected from a low stringency of about 2.0 x SSC at 50°C to a high stringency of about 0.2 x SSC at 50°C. In addition, the temperature in the wash step can be increased from low stringency conditions at room temperature, about 22°C, to high stringency conditions at about 65°C.

As described in the literature, genes for other hedgehog proteins, e.g., from other animals, can be obtained from mRNA or genomic DNA samples using techniques well known in the art. For example, a cDNA encoding a *hedgehog* protein can be obtained by isolating total mRNA from a cell, e.g. a mammalian cell, e.g. a human cell, including embryonic cells. Double stranded cDNAs can then be prepared from the total mRNA, and subsequently inserted into a suitable plasmid or bacteriophage vector using any one of a number of known techniques. The gene encoding a *hedgehog* protein can also be cloned using established polymerase chain reaction techniques.

Preferred nucleic acids encode a *hedgehog* polypeptide comprising an amino acid sequence at least 60% homologous or identical, more preferably 70% homologous or identical, and most preferably 80% homologous or identical with an amino acid sequence selected from the

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group consisting of SEQ ID Nos:8-14. Nucleic acids which encode polypeptides at least about 90%, more preferably at least about 95%, and most preferably at least about 98-99% homology or identity with an amino acid sequence represented in one of SEQ ID Nos:8-14 are also within the scope of the invention.

In addition to native *hedgehog* proteins, h*edgehog* polypeptides preferred by the present invention are at least 60% homologous or identical, more preferably 70% homologous or identical and most preferably 80% homologous or identical with an amino acid sequence represented by any of SEQ ID Nos:8-14. Polypeptides which are at least 90%, more preferably at least 95%, and most preferably at least about 98-99% homologous or identical with a sequence selected from the group consisting of SEQ ID Nos:8-14 are also within the scope of the invention. The only prerequisite is that the *hedgehog* polypeptide is capable of modulating the growth state of T lymphocytes.

The term "recombinant protein" refers to a polypeptide of the present invention which is produced by recombinant DNA techniques, wherein generally, DNA encoding a *hedgehog* polypeptide is inserted into a suitable expression vector which is in turn used to transform a host cell to produce the heterologous protein. Moreover, the phrase "derived from", with respect to a recombinant *hedgehog* gene, is meant to include within the meaning of "recombinant protein" those proteins having an amino acid sequence of a native *hedgehog* protein, or an amino acid sequence similar thereto which is generated by mutations including substitutions and deletions (including truncation) of a naturally occurring form of the protein.

The method of the present invention can also be carried out using variant forms of the naturally occurring *hedgehog* polypeptides, e.g., mutational variants.

As is known in the art, hedgehog polypeptides can be produced by standard biological techniques or by chemical synthesis. For example, a host cell transfected with a nucleic acid vector directing expression of a nucleotide sequence encoding the subject polypeptides can be cultured under appropriate conditions to allow expression of the peptide to occur. The polypeptide hedgehog may be secreted and isolated from a mixture of cells and medium containing the recombinant hedgehog polypeptide. Alternatively, the peptide may be retained cytoplasmically by removing the signal peptide sequence from the recombinant hedgehog gene and the cells harvested, lysed and the protein isolated. A cell culture includes host cells, media and other byproducts. Suitable media for cell culture are well known in the art. The recombinant hedgehog polypeptide can be isolated from cell culture medium, host cells, or both using techniques known in the art for purifying proteins including ion-exchange chromatography, gel filtration chromatography, ultrafiltration, electrophoresis, and immunoaffinity purification with

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antibodies specific for such peptide. In a preferred embodiment, the recombinant *hedgehog* polypeptide is a fusion protein containing a domain which facilitates its purification, such as an *hedgehog*/GST fusion protein. The host cell may be any prokaryotic or eukaryotic cell.

Recombinant *hedgehog* genes can be produced by ligating nucleic acid encoding an *hedgehog* protein, or a portion thereof, into a vector suitable for expression in either prokaryotic cells, eukaryotic cells, or both. Expression vectors for production of recombinant forms of the subject *hedgehog* polypeptides include plasmids and other vectors. For instance, suitable vectors for the expression of a *hedgehog* polypeptide include plasmids of the types: pBR322-derived plasmids, pEMBL-derived plasmids, pEX-derived plasmids, pBTac-derived plasmids and pUC-derived plasmids for expression in prokaryotic cells, such as *E. coli*.

A number of vectors exist for the expression of recombinant proteins in yeast. For instance, YEP24, YIP5, YEP51, YEP52, pYES2, and YRP17 are cloning and expression vehicles useful in the introduction of genetic constructs into *S. cerevisiae* (see, for example, Broach *et al.* (1983) in *Experimental Manipulation of Gene Expression*, ed. M. Inouye Academic Press, p. 83, incorporated by reference herein). These vectors can replicate in *E. coli* due to the presence of the pBR322 ori, and in *S. cerevisiae* due to the replication determinant of the yeast 2 micron plasmid. In addition, drug resistance markers such as ampicillin can be used. In an illustrative embodiment, an *hedgehog* polypeptide is produced recombinantly utilizing an expression vector generated by sub-cloning the coding sequence of one of the *hedgehog* genes represented in SEQ ID Nos:1-7.

The preferred mammalian expression vectors contain both prokaryotic sequences, to facilitate the propagation of the vector in bacteria, and one or more eukaryotic transcription units that are expressed in eukaryotic cells. The pcDNAI/amp, pcDNAI/neo, pRc/CMV, pSV2gpt, pSV2neo, pSV2-dhfr, pTk2, pRSVneo, pMSG, pSVT7, pko-neo and pHyg derived vectors are examples of mammalian expression vectors suitable for transfection of eukaryotic cells. Some of these vectors are modified with sequences from bacterial plasmids, such as pBR322, to facilitate replication and drug resistance selection in both prokaryotic and eukaryotic cells. Alternatively, derivatives of viruses such as the bovine papillomavirus (BPV-1), or Epstein-Barr virus (pHEBo, pREP-derived and p205) can be used for transient expression of proteins in eukaryotic cells. The various methods employed in the preparation of the plasmids and transformation of host organisms are well known in the art. For other suitable expression systems for both prokaryotic and eukaryotic cells, as well as general recombinant procedures, see *Molecular Cloning A Laboratory Manual*, 2nd Ed., ed. by Sambrook, Fritsch and Maniatis (Cold Spring Harbor Laboratory Press: 1989) Chapters 16 and 17.

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In some instances, it may be desirable to express the recombinant *hedgehog* polypeptide by the use of a baculovirus expression system. Examples of such baculovirus expression systems include pVL-derived vectors (such as pVL1392, pVL1393 and pVL941), pAcUW-derived vectors (such as pAcUW1), and pBlueBac-derived vectors (such as the β-gal containing pBlueBac III).

When it is desirable to express only a portion of an *hedgehog* protein, such as a form lacking a portion of the N-terminus, i.e. a truncation mutant which lacks the signal peptide, it may be necessary to add a start codon (ATG) to the oligonucleotide fragment containing the desired sequence to be expressed. It is well known in the art that a methionine at the N-terminal position can be enzymatically cleaved by the use of the enzyme methionine aminopeptidase (MAP). MAP has been cloned from *E. coli* (Ben-Bassat et al. (1987) *J. Bacteriol.* 169:751-757) and *Salmonella typhimurium* and its *in vitro* activity has been demonstrated on recombinant proteins (Miller et al. (1987) *PNAS* 84:2718-1722). Therefore, removal of an N-terminal methionine, if desired, can be achieved either *in vivo* by expressing *hedgehog*-derived polypeptides in a host which produces MAP (e.g., *E. coli* or CM89 or *S. cerevisiae*), or *in vitro* by use of purified MAP (e.g., procedure of Miller et al., *supra*).

Alternatively, the coding sequences for the polypeptide can be incorporated as a part of a fusion gene including a nucleotide sequence encoding a different polypeptide. It is widely appreciated that fusion proteins can also facilitate the expression of proteins, and accordingly, can be used in the expression of the *hedgehog* polypeptides of the present invention. For example, *hedgehog* polypeptides can be generated as glutathione-S-transferase (GST-fusion) proteins. Such GST-fusion proteins can enable easy purification of the *hedgehog* polypeptide, as for example by the use of glutathione-derivatized matrices (see, for example, *Current Protocols in Molecular Biology*, eds. Ausubel et al. (N.Y.: John Wiley & Sons, 1991)). In another embodiment, a fusion gene coding for a purification leader sequence, such as a poly-(His)/enterokinase cleavage site sequence, can be used to replace the signal sequence which naturally occurs at the N-terminus of the *hedgehog* protein (e.g. of the pro-form, in order to permit purification of the poly(His)-*hedgehog* protein by affinity chromatography using a Ni<sup>2+</sup> metal resin. The purification leader sequence can then be subsequently removed by treatment with enterokinase (e.g., see Hochuli et al. (1987) *J. Chromatography* 411:177; and Janknecht et al. *PNAS* 88:8972).

Techniques for making fusion genes are known to those skilled in the art. Essentially, the joining of various DNA fragments coding for different polypeptide sequences is performed in accordance with conventional techniques, employing blunt-ended or stagger-ended termini for

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ligation, restriction enzyme digestion to provide for appropriate termini, filling-in of cohesive ends as appropriate, alkaline phosphatase treatment to avoid undesirable joining, and enzymatic ligation. In another embodiment, the fusion gene can be synthesized by conventional techniques including automated DNA synthesizers. Alternatively, PCR amplification of gene fragments can be carried out using anchor primers which give rise to complementary overhangs between two consecutive gene fragments which can subsequently be annealed to generate a chimeric gene sequence (see, for example, *Current Protocols in Molecular Biology*, eds. Ausubel et al. John Wiley & Sons: 1992).

Hedgehog polypeptides may also be chemically modified to create hedgehog derivatives by forming covalent or aggregate conjugates with other chemical moieties, such as glycosyl groups, cholesterol, isoprenoids, lipids, phosphate, acetyl groups and the like. Covalent derivatives of hedgehog proteins can be prepared by linking the chemical moieties to functional groups on amino acid sidechains of the protein or at the N-terminus or at the C-terminus of the polypeptide.

For instance, *hedgehog* proteins can be generated to include a moiety, other than sequence naturally associated with the protein, that binds a component of the extracellular matrix and enhances localization of the analog to cell surfaces. For example, sequences derived from the fibronectin "type-III repeat", such as a tetrapeptide sequence R-G-D-S (Pierschbacher et al. (1984) *Nature* 309:30-3; and Kornblihtt et al. (1985) *EMBO* 4:1755-9) can be added to the *hedgehog* polypeptide to support attachment of the chimeric molecule to a cell through binding ECM components (Ruoslahti et al. (1987) *Science* 238:491-497; Pierschbacheret al. (1987) *J. Biol. Chem.* 262:17294-8.; Hynes (1987) *Cell* 48:549-54; and Hynes (1992) *Cell* 69:11-25).

In a preferred embodiment, the *hedgehog* polypeptide is isolated from, or is otherwise substantially free of, other cellular proteins, especially other extracellular or cell surface associated proteins which may normally be associated with the *hedgehog* polypeptide, unless provided in the form of fusion protein with the *hedgehog* polypeptide. The term "substantially free of other cellular or extracellular proteins" (also referred to herein as "contaminating proteins") or "substantially pure preparations" or "purified preparations" are defined as encompassing preparations of *hedgehog* polypeptides having less than 20% (by dry weight) contaminating protein, and preferably having less than 5% contaminating protein. By "purified", it is meant that the indicated molecule is present in the substantial absence of other biological macromolecules, such as other proteins. The term "purified" as used herein preferably means at least 80% by dry weight, more preferably in the range of 95-99% by weight, and most preferably at least 99.8% by weight, of biological macromolecules of the same type present (but water,

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buffers, and other small molecules, especially molecules having a molecular weight of less than 5000, can be present). The term "pure" as used herein preferably has the same numerical limits as "purified" immediately above.

As described above for recombinant polypeptides, isolated *hedgehog* polypeptides can include all or a portion of the amino acid sequences represented in any of SEQ ID Nos:10-18 or 20, or a homologous sequence thereto. Preferred fragments of the subject *hedgehog* proteins correspond to the N-terminal and C-terminal proteolytic fragments of the mature protein. Bioactive fragments of *hedgehog* polypeptides are described in great detail in PCT publications WO 95/18856 and WO 96/17924.

With respect to bioctive fragments of *hedgehog* polypeptide, preferred *hedgehog* therapeutics include at least 50 (contiguous) amino acid residues of a *hedgehog* polypeptide, more preferably at least 100 (contiguous), and even more preferably at least 150 (contiguous) residues.

Another preferred *hedgehog* polypeptide which can be included in the *hedgehog* therapeutic is an N-terminal fragment of the mature protein having a molecular weight of approximately 19 kDa.

Preferred human *hedgehog* proteins include N-terminal fragments corresponding approximately to residues 24-197 of SEQ ID No. 15, 28-202 of SEQ ID No. 16, and 23-198 of SEQ ID No. 17. By "corresponding approximately" it is meant that the sequence of interest is at most 20 amino acid residues different in length to the reference sequence, though more preferably at most 5, 10 or 15 amino acid different in length.

As described above for recombinant polypeptides, isolated *hedgehog* polypeptides can include all or a portion of the amino acid sequences represented in SEQ ID No:8, SEQ ID No:9, SEQ ID No:10, SEQ ID No:11, SEQ ID No:12, SEQ ID No:13 or SEQ ID No:14, or a homologous sequence thereto. Preferred fragments of the subject *hedgehog* proteins correspond to the N-terminal and C-terminal proteolytic fragments of the mature protein. Bioactive fragments of hedgehog polypeptides are described in great detail in PCT publications WO 95/18856 and WO 96/17924.

Still other preferred *hedgehog* polypeptides includes an amino acid sequence represented by the formula A-B wherein: (i) A represents all or the portion of the amino acid sequence designated by residues 1-168 of SEQ ID No:21; and B represents at least one amino acid residue of the amino acid sequence designated by residues 169-221 of SEQ ID No:21; (ii) A represents all or the portion of the amino acid sequence designated by residues 24-193 of SEQ ID No:15;

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and B represents at least one amino acid residue of the amino acid sequence designated by residues 194-250 of SEQ ID No:15; (iii) A represents all or the portion of the amino acid sequence designated by residues 25-193 of SEQ ID No:13; and B represents at least one amino acid residue of the amino acid sequence designated by residues 194-250 of SEQ ID No:13; (iv) A represents all or the portion of the amino acid sequence designated by residues 23-193 of SEQ ID No:11; and B represents at least one amino acid residue of the amino acid sequence designated by residues 194-250 of SEQ ID No:11; (v) A represents all or the portion of the amino acid sequence designated by residues 28-197 of SEQ ID No:12; and B represents at least one amino acid residue of the amino acid sequence designated by residues 198-250 of SEQ ID No:12; (vi) A represents all or the portion of the amino acid sequence designated by residues 29-197 of SEQ ID No:16; and B represents at least one amino acid residue of the amino acid sequence designated by residues 198-250 of SEQ ID No:16; or (vii) A represents all or the portion of the amino acid sequence designated by residues 23-193 of SEQ ID No. 17, and B represents at least one amino acid residue of the amino acid sequence designated by residues 194-250 of SEQ ID No. 17. In certain preferred embodiments, A and B together represent a contiguous polypeptide sequence designated sequence, A represents at least 25, 50, 75, 100, 125 or 150 (contiguous) amino acids of the designated sequence, and B represents at least 5, 10, or 20 (contiguous) amino acid residues of the amino acid sequence designated by corresponding entry in the sequence listing, and A and B together preferably represent a contiguous sequence corresponding to the sequence listing entry. Similar fragments from other hedgehog also contemplated, e.g., fragments which correspond to the preferred fragments from the sequence listing entries which are enumerated above. In preferred embodiments, the hedgehog polypeptide includes a C-terminal glycine (or other appropriate residue) which is derivatized with a cholesterol.

Isolated peptidyl portions of *hedgehog* proteins can be obtained by screening peptides recombinantly produced from the corresponding fragment of the nucleic acid encoding such peptides. In addition, fragments can be chemically synthesized using techniques known in the art such as conventional Merrifield solid phase f-Moc or t-Boc chemistry. For example, a *hedgehog* polypeptide of the present invention may be arbitrarily divided into fragments of desired length with no overlap of the fragments, or preferably divided into overlapping fragments of a desired length. The fragments can be produced (recombinantly or by chemical synthesis) and tested to identify those peptidyl fragments which can function as either agonists or antagonists of a wild-type (e.g., "authentic") *hedgehog* protein. For example, Román et al. (1994) *Eur J Biochem* 222:65-73 describe the use of competitive-binding assays using short, overlapping synthetic peptides from larger proteins to identify binding domains.

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The recombinant *hedgehog* polypeptides of the present invention also include homologs of the authentic *hedgehog* proteins, such as versions of those protein which are resistant to proteolytic cleavage, as for example, due to mutations which alter potential cleavage sequences or which inactivate an enzymatic activity associated with the protein. *Hedgehog* homologs of the present invention also include proteins which have been post-translationally modified in a manner different than the authentic protein. Exemplary derivatives of *hedgehog* proteins include polypeptides which lack N-glycosylation sites (e.g. to produce an unglycosylated protein), which lack sites for cholesterolization, and/or which lack N-terminal and/or C-terminal sequences.

Modification of the structure of the subject *hedgehog* polypeptides can also be for such purposes as enhancing therapeutic or prophylactic efficacy, or stability (e.g., *ex vivo* shelf life and resistance to proteolytic degradation *in vivo*). Such modified peptides, when designed to retain at least one activity of the naturally-occurring form of the protein, are considered functional equivalents of the *hedgehog* polypeptides described in more detail herein. Such modified peptides can be produced, for instance, by amino acid substitution, deletion, or addition.

It is well known in the art that one could reasonably expect that certain isolated replacements of amino acids, e.g., replacement of an amino acid residue with another related amino acid (i.e. isosteric and/or isoelectric mutations), can be carried out without major effect on the biological activity of the resulting molecule. Conservative replacements are those that take place within a family of amino acids that are related in their side chains. Genetically encoded amino acids are can be divided into four families: (1) acidic = aspartate, glutamate; (2) basic = lysine, arginine, histidine; (3) nonpolar = alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan; and (4) uncharged polar = glycine, asparagine, glutamine, cysteine, serine, threonine, tyrosine. Phenylalanine, tryptophan, and tyrosine are sometimes classified jointly as aromatic amino acids. In similar fashion, the amino acid repertoire can be grouped as (1) acidic = aspartate, glutamate; (2) basic = lysine, arginine histidine, (3) aliphatic = glycine, alanine, valine, leucine, isoleucine, serine, threonine, with serine and threonine optionally be grouped separately as aliphatic-hydroxyl; (4) aromatic = phenylalanine, tyrosine, tryptophan; (5) amide = asparagine, glutamine; and (6) sulfur -containing = cysteine and methionine. (see, for example, Biochemistry, 2nd ed., Ed. by L. Stryer, WH Freeman and Co.: 1981). Whether a change in the amino acid sequence of a peptide results in a functional hedgehog homolog (e.g. functional in the sense that it acts to mimic or antagonize the wild-type form) can be readily determined by assessing the ability of the variant peptide to produce a response in cells in a fashion similar to the wild-type protein, or competitively inhibit such a response. Polypeptides in which more than one replacement has taken place can readily be tested in the same manner.

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It is specifically contemplated that the methods of the present invention can be carried using homologs of naturally occurring hedgehog proteins. In one embodiment, the invention contemplates using hedgehog polypeptides generated by combinatorial mutagenesis. methods, as are known in the art, are convenient for generating both point and truncation mutants, and can be especially useful for identifying potential variant sequences (e.g. homologs) that are functional in binding to a receptor for hedgehog proteins. The purpose of screening such combinatorial libraries is to generate, for example, novel hedgehog homologs which can act as either agonists or antagonist. To illustrate, hedgehog homologs can be engineered by the present method to provide more efficient binding to a cognate receptor, such as patched, yet still retain at least a portion of an activity associated with hedgehog. Thus, combinatorially-derived homologs can be generated to have an increased potency relative to a naturally occurring form of the protein. Likewise, hedgehog homologs can be generated by the present combinatorial approach to act as antagonists, in that they are able to mimic, for example, binding to other extracellular matrix components (such as receptors), yet not induce any biological response, thereby inhibiting the action of authentic hedgehog or hedgehog agonists. Moreover, manipulation of certain domains of hedgehog by the present method can provide domains more suitable for use in fusion proteins, such as one that incorporates portions of other proteins which are derived from the extracellular matrix and/or which bind extracellular matrix components.

To further illustrate the state of the art of combinatorial mutagenesis, it is noted that the review article of Gallop et al. (1994) *J Med Chem* 37:1233 describes the general state of the art of combinatorial libraries as of the earlier 1990's. In particular, Gallop et al state at page 1239 "[s]creening the analog libraries aids in determining the minimum size of the active sequence and in identifying those residues critical for binding and intolerant of substitution". In addition, the Ladner et al. PCT publication WO90/02809, the Goeddel et al. U.S. Patent 5,223,408, and the Markland et al. PCT publication WO92/15679 illustrate specific techniques which one skilled in the art could utilize to generate libraries of *hedgehog* variants which can be rapidly screened to identify variants/fragments which retained a particular activity of the *hedgehog* polypeptides. These techniques are exemplary of the art and demonstrate that large libraries of related variants/truncants can be generated and assayed to isolate particular variants without undue experimentation. Gustin et al. (1993) *Virology* 193:653, and Bass et al. (1990) *Proteins: Structure, Function and Genetics* 8:309-314 also describe other exemplary techniques from the art which can be adapted as means for generating mutagenic variants of *hedgehog* polypeptides.

Indeed, it is plain from the combinatorial mutagenesis art that large scale mutagenesis of hedgehog proteins, without any preconceived ideas of which residues were critical to the biological function, and generate wide arrays of variants having equivalent biological activity.

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Indeed, it is the ability of combinatorial techniques to screen billions of different variants by high throughout analysis that removes any requirement of *a priori* understanding or knowledge of critical residues.

To illsutrate, the amino acid sequences for a population of *hedgehog* homologs or other related proteins are aligned, preferably to promote the highest homology possible. Such a population of variants can include, for example, *hedgehog* homologs from one or more species. Amino acids which appear at each position of the aligned sequences are selected to create a degenerate set of combinatorial sequences. In a preferred embodiment, the variegated library of *hedgehog* variants is generated by combinatorial mutagenesis at the nucleic acid level, and is encoded by a variegated gene library. For instance, a mixture of synthetic oligonucleotides can be enzymatically ligated into gene sequences such that the degenerate set of potential *hedgehog* sequences are expressible as individual polypeptides, or alternatively, as a set of larger fusion proteins (e.g. for phage display) containing the set of *hedgehog* sequences therein.

As illustrated in PCT publication WO 95/18856, to analyze the sequences of a population of variants, the amino acid sequences of interest can be aligned relative to sequence homology. The presence or absence of amino acids from an aligned sequence of a particular variant is relative to a chosen consensus length of a reference sequence, which can be real or artificial.

In an illustrative embodiment, alignment of exons 1, 2 and a portion of exon 3 encoded sequences (e.g. the N-terminal approximately 221 residues of the mature protein) of each of the *Shh* clones produces a degenerate set of *Shh* polypeptides represented by the general formula:

C-G-P-G-R-G-X(1)-G-X(2)-R-R-H-P-K-K-L-T-P-L-A-Y-K-Q-F-I-P-N-V-A-E-K-T-L-G-A-S-G-R-Y-E-G-K-I-X(3)-R-N-S-E-R-F-K-E-L-T-P-N-Y-N-P-D-I-I-F-K-D-E-E-N-T-G-A-D-R-L-M-T-Q-R-C-K-D-K-L-N-X(4)-L-A-I-S-V-M-N-X(5)-W-P-G-V-X(6)-L-R-V-T-E-G-W-D-E-D-G-H-H-X(7)-E-E-S-L-H-Y-E-G-R-A-V-D-I-T-T-S-D-R-D-X(8)-S-K-Y-G-X(9)-L-X(10)-R-L-A-V-E-A-G-F-D-W-V-Y-Y-E-S-K-A-H-I-H-C-S-V-K-A-E-N-S-V-A-A-K-S-G-G-C-F-P-G-S-A-X(11)-V-X(12)-L-X(13)-X(14)-G-G-X(15)-K-X-(16)-V-K-D-L-X(17)-P-G-D-X(18)-V-L-A-A-D-X(19)-X(20)-G-X(21)-L-X(22)-X(23)-S-D-F-X(24)-X(25)-F-X(26)-D-R (SEQ ID No: 21

wherein each of the degenerate positions "X" can be an amino acid which occurs in that position in one of the human, mouse, chicken or zebrafish *Shh* clones, or, to expand the library, each X can also be selected from amongst amino acid residue which would be conservative substitutions for the amino acids which appear naturally in each of those positions. For instance, Xaa(1) represents Gly, Ala, Val, Leu, Ile, Phe, Tyr or Trp; Xaa(2) represents Arg, His or Lys; Xaa(3) represents Gly, Ala, Val, Leu, Ile, Ser or Thr; Xaa(4) represents Gly, Ala, Val, Leu, Ile, Ser or

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Thr; Xaa(5) represents Lys, Arg, His, Asn or Gln; Xaa(6) represents Lys, Arg or His; Xaa(7) represents Ser, Thr, Tyr, Trp or Phe; Xaa(8) represents Lys, Arg or His; Xaa(9) represents Met, Cys, Ser or Thr; Xaa(10) represents Gly, Ala, Val, Leu, Ile, Ser or Thr; Xaa(11) represents Leu, Val, Met, Thr or Ser; Xaa(12) represents His, Phe, Tyr, Ser, Thr, Met or Cys; Xaa(13) represents Gln, Asn, Glu, or Asp; Xaa(14) represents His, Phe, Tyr, Thr, Gln, Asn, Glu or Asp; Xaa(15) represents Gln, Asn, Glu, Asp, Thr, Ser, Met or Cys; Xaa(16) represents Ala, Gly, Cys, Leu, Val or Met; Xaa(17) represents Arg, Lys, Met, Ile, Asn, Asp, Glu, Gln, Ser, Thr or Cys; Xaa(18) represents Arg, Lys, Met or Ile; Xaa(19) represents Ala, Gly, Cys, Asp, Glu, Gln, Asn, Ser, Thr or Met; Xaa(20) represents Ala, Gly, Cys, Asp, Asn, Glu or Gln; Xaa(21) represents Arg, Lys, Met, Ile, Asn, Asp, Glu or Gln; Xaa(22) represent Leu, Val, Met or Ile; Xaa(23) represents Phe, Tyr, Thr, His or Trp; Xaa(24) represents Ile, Val, Leu or Met; .Xaa(25) represents Met, Cys, Ile, Leu, Val, Thr or Ser; Xaa(26) represents Leu, Val, Met, Thr or Ser. In an even more expansive library, each X can be selected from any amino acid.

In similar fashion, alignment of each of the human, mouse, chicken and zebrafish *hedgehog* clones, can provide a degenerate polypeptide sequence represented by the general formula:

C-G-P-G-R-G-X(1)-X(2)-X(3)-R-R-X(4)-X(5)-X(6)-P-K-X(7)-L-X(8)-P-L-X(9)-Y-K-Q-F-X(10)-P-X(11)-X(12)-X(13)-E-X(14)-T-L-G-A-S-G-X(15)-X(16)-E-G-X(17)-X(18)-X(19)-R-X(20)-S-E-R-F-X(21)-X(22)-L-T-P-N-Y-N-P-D-I-I-F-K-D-E-E-N-X(23)-G-A-D-R-L-M-T-X(24)-R-C-K-X(25)-X(26)-X(27)-N-X(28)-L-A-I-S-V-M-N-X(29)-W-P-G-V-X(30)-L-R-V-T-E-G-X(31)-D-E-D-G-H-H-X(32)-X(33)-X(34)-S-L-H-Y-E-G-R-A-X(35)-D-I-T-T-S-D-R-D-X(36)-X(37)-K-Y-G-X(38)-L-X(39)-R-L-A-V-E-A-G-F-D-W-V-Y-Y-E-S-X(40)-X(41)-H-X(42)-H-X(43)-S-V-K-X(44)-X(45) (SEQ IDNo:22

wherein, as above, each of the degenerate positions "X" can be an amino acid which occurs in a corresponding position in one of the wild-type clones, and may also include amino acid residue which would be conservative substitutions, or each X can be any amino acid residue. In an exemplary embodiment, Xaa(1) represents Gly, Ala, Val, Leu, Ile, Pro, Phe or Tyr; Xaa(2) represents Gly, Ala, Val, Leu or Ile; Xaa(3) represents Gly, Ala, Val, Leu, Ile, Lys, His or Arg; Xaa(4) represents Lys, Arg or His; Xaa(5) represents Phe, Trp, Tyr or an amino acid gap; Xaa(6) represents Gly, Ala, Val, Leu, Ile or an amino acid gap; Xaa(7) represents Asn, Gln, His, Arg or Lys; Xaa(8) represents Gly, Ala, Val, Leu, Ile, Ser or Thr; Xaa(10) represents Gly, Ala, Val, Leu, Ile, Ser or Thr; Xaa(11) represents Ser, Thr, Gln or Asn; Xaa(12) represents Met, Cys, Gly, Ala, Val, Leu, Ile, Ser or Thr; Xaa(15) represents Gly, Ala, Val, Leu, Ile, Pro, Arg, His or Lys; Xaa(16) represents Gly, Ala, Val, Leu, Ile, Pro, Arg, His or Lys; Xaa(16) represents Gly, Ala, Val, Leu, Ile, Phe or

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Tyr; Xaa(17) represents Arg, His or Lys; Xaa(28) represents Gly, Ala, Val, Leu, Ile, Ser or Thr; Xaa(19) represents Thr or Ser; Xaa(20) represents Gly, Ala, Val, Leu, Ile, Asn or Gln; Xaa(21) represents Arg, His or Lys; Xaa(22) represents Asp or Glu; Xaa(23) represents Ser or Thr; Xaa(24) represents Glu, Asp, Gln or Asn; Xaa(25) represents Glu or Asp; Xaa(26) represents Arg, His or Lys; Xaa(27) represents Gly, Ala, Val, Leu or Ile; Xaa(28) represents Gly, Ala, Val, Leu, Ile, Thr or Ser; Xaa(29) represents Met, Cys, Gln, Asn, Arg, Lys or His; Xaa(30) represents Arg, His or Lys; Xaa(31) represents Trp, Phe, Tyr, Arg, His or Lys; Xaa(32) represents Gly, Ala, Val, Leu, Ile, Ser, Thr, Tyr or Phe; Xaa(33) represents Gln, Asn, Asp or Glu; Xaa(34) represents Asp or Glu; Xaa(35) represents Gly, Ala, Val, Leu, or Ile; Xaa(36) represents Arg, His or Lys; Xaa(37) represents Asn, Gln, Thr or Ser; Xaa(38) represents Gly, Ala, Val, Leu, Ile, Ser, Thr, Met or Cys; Xaa(39) represents Gly, Ala, Val, Leu, Ile, Thr or Ser; Xaa(40) represents Arg, His or Lys; Xaa(41) represents Asn, Gln, Gly, Ala, Val, Leu or Ile; Xaa(42) represents Gly, Ala, Val, Leu or Ile; Xaa(44) represents Gly, Ala, Val, Leu, Ile, Ser, Thr or Cys; Xaa(44) represents Gly, Ala, Val, Leu, Ile, Ser, Thr or Cys; Xaa(44) represents Gly, Ala, Val, Leu, Ile, Ser, Thr or Cys; Xaa(45) represents Asp or Glu.

There are many ways by which the library of potential *hedgehog* homologs can be generated from a degenerate oligonucleotide sequence. Chemical synthesis of a degenerate gene sequence can be carried out in an automatic DNA synthesizer, and the synthetic genes then ligated into an appropriate expression vector. The purpose of a degenerate set of genes is to provide, in one mixture, all of the sequences encoding the desired set of potential *hedgehog* sequences. The synthesis of degenerate oligonucleotides is well known in the art (see for example, Narang, SA (1983) *Tetrahedron* 39:3; Itakura et al. (1981) *Recombinant DNA, Proc* 3rd Cleveland Sympos. Macromolecules, ed. AG Walton, Amsterdam: Elsevier pp273-289; Itakura et al. (1984) Annu. Rev. Biochem. 53:323; Itakura et al. (1984) Science 198:1056; Ike et al. (1983) Nucleic Acid Res. 11:477. Such techniques have been employed in the directed evolution of other proteins (see, for example, Scott et al. (1990) Science 249:386-390; Roberts et al. (1992) PNAS 89:2429-2433; Devlin et al. (1990) Science 249: 404-406; Cwirla et al. (1990) PNAS 87: 6378-6382; as well as U.S. Patents Nos. 5,223,409, 5,198,346, and 5,096,815).

A wide range of techniques are known in the art for screening gene products of combinatorial libraries made by point mutations, and for screening cDNA libraries for gene products having a certain property. Such techniques will be generally adaptable for rapid screening of the gene libraries generated by the combinatorial mutagenesis of *hedgehog* homologs. The most widely used techniques for screening large gene libraries typically comprises cloning the gene library into replicable expression vectors, transforming appropriate cells with the resulting library of vectors, and expressing the combinatorial genes under conditions in which detection of a desired activity facilitates relatively easy isolation of the

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vector encoding the gene whose product was detected. Each of the illustrative assays described below are amenable to high through-put analysis as necessary to screen large numbers of degenerate *hedgehog* sequences created by combinatorial mutagenesis techniques.

In one embodiment, the combinatorial library is designed to be secreted (e.g. the polypeptides of the library all include a signal sequence but no transmembrane or cytoplasmic domains), and is used to transfect a eukaryotic cell that can be co-cultured with peripehral nerve cells. A functional *hedgehog* protein secreted by the cells expressing the combinatorial library will diffuse to neighboring T lymphocytes and induce a particular biological response, such as proliferation or differentiation. The pattern of detection of such a change in phenotype will resemble a gradient function, and will allow the isolation (generally after several repetitive rounds of selection) of cells producing *hedgehog* homologs active as neurotrophic agents. Likewise, *hedgehog* antagonists can be selected in similar fashion by the ability of the cell producing a functional antagonist to protect neighboring cells (e.g., to inhibit proliferation) from the effect of wild-type *hedgehog* added to the culture media.

To illustrate, target T lymphocytes are cultured in 24-well microtitre plates. Other eukaryotic cells are transfected with the combinatorial *hedgehog* gene library and cultured in cell culture inserts (e.g. Collaborative Biomedical Products, Catalog #40446) that are able to fit into the wells of the microtitre plate. The cell culture inserts are placed in the wells such that recombinant *hedgehog* homologs secreted by the cells in the insert can diffuse through the porous bottom of the insert and contact the target cells in the microtitre plate wells. After a period of time sufficient for functional forms of a *hedgehog* protein to produce a measurable response in the target cells, such as growth state, the inserts are removed and the effect of the variant *hedgehog* proteins on the target cells determined. Cells from the inserts corresponding to wells which score positive for activity can be split and re-cultured on several inserts, the process being repeated until the active clones are identified.

In yet another screening assay, the candidate *hedgehog* gene products are displayed on the surface of a cell or viral particle, and the ability of particular cells or viral particles to associate with a *hedgehog*-binding moiety (such as the *patched* protein or other *hedgehog* receptor) via this gene product is detected in a "panning assay". Such panning steps can be carried out on cells cultured from embryos. For instance, the gene library can be cloned into the gene for a surface membrane protein of a bacterial cell, and the resulting fusion protein detected by panning (Ladner et al., WO 88/06630; Fuchs et al. (1991) *Bio/Technology* 9:1370-1371; and Goward et al. (1992) *TIBS* 18:136-140). In a similar fashion, fluorescently labeled molecules which bind *hedgehog* can be used to score for potentially functional *hedgehog* homologs. Cells can be

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visually inspected and separated under a fluorescence microscope, or, where the morphology of the cell permits, separated by a fluorescence-activated cell sorter.

In an alternate embodiment, the gene library is expressed as a fusion protein on the surface of a viral particle. For instance, in the filamentous phage system, foreign peptide sequences can be expressed on the surface of infectious phage, thereby conferring two significant benefits. First, since these phage can be applied to affinity matrices at very high concentrations, large number of phage can be screened at one time. Second, since each infectious phage displays the combinatorial gene product on its surface, if a particular phage is recovered from an affinity matrix in low yield, the phage can be amplified by another round of infection. The group of almost identical *E.coli* filamentous phages M13, fd, and f1 are most often used in phage display libraries, as either of the phage gIII or gVIII coat proteins can be used to generate fusion proteins without disrupting the ultimate packaging of the viral particle (Ladner et al. PCT publication WO 90/02909; Garrard et al., PCT publication WO 92/09690; Marks et al. (1992) J. Biol. Chem. 267:16007-16010; Griffths et al. (1993) EMBO J 12:725-734; Clackson et al. (1991) Nature 352:624-628; and Barbas et al. (1992) PNAS 89:4457-4461).

In an illustrative embodiment, the recombinant phage antibody system (RPAS, Pharamacia Catalog number 27-9400-01) can be easily modified for use in expressing and screening hedgehog combinatorial libraries. For instance, the pCANTAB 5 phagemid of the RPAS kit contains the gene which encodes the phage gIII coat protein. The hedgehog combinatorial gene library can be cloned into the phagemid adjacent to the gIII signal sequence such that it will be expressed as a gIII fusion protein. After ligation, the phagemid is used to transform competent E. coli TG1 cells. Transformed cells are subsequently infected with M13KO7 helper phage to rescue the phagemid and its candidate hedgehog gene insert. The resulting recombinant phage contain phagemid DNA encoding a specific candidate hedgehog, and display one or more copies of the corresponding fusion coat protein. The phage-displayed candidate hedgehog proteins which are capable of binding an hedgehog receptor are selected or enriched by panning. For instance, the phage library can be applied to cells which express the patched protein and unbound phage washed away from the cells. The bound phage is then isolated, and if the recombinant phage express at least one copy of the wild type gIII coat protein, they will retain their ability to infect E. coli. Thus, successive rounds of reinfection of E. coli, and panning will greatly enrich for hedgehog homologs, which can then be screened for further biological activities in order to differentiate agonists and antagonists.

Combinatorial mutagenesis has a potential to generate very large libraries of mutant proteins, e.g., in the order of  $10^{26}$  molecules. Combinatorial libraries of this size may be

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technically challenging to screen even with high throughput screening assays such as phage display. To overcome this problem, a new technique has been developed recently, recursive ensemble mutagenesis (REM), which allows one to avoid the very high proportion of non-functional proteins in a random library and simply enhances the frequency of functional proteins, thus decreasing the complexity required to achieve a useful sampling of sequence space. REM is an algorithm which enhances the frequency of functional mutants in a library when an appropriate selection or screening method is employed (Arkin and Yourvan, 1992, *PNAS USA* 89:7811-7815; Yourvan et al., 1992, *Parallel Problem Solving from Nature*, 2., In Maenner and Manderick, eds., Elsevir Publishing Co., Amsterdam, pp. 401-410; Delgrave et al., 1993, *Protein Engineering* 6(3):327-331).

The invention also provides for reduction of the hedgehog protein to generate mimetics, e.g. peptide or non-peptide agents, which are able to disrupt binding of a hedgehog polypeptide of the present invention with an hedgehog receptor. Thus, such mutagenic techniques as described above are also useful to map the determinants of the hedgehog proteins which participate in protein-protein interactions involved in, for example, binding of the subject hedgehog polypeptide to other extracellular matrix components. To illustrate, the critical residues of a subject hedgehog polypeptide which are involved in molecular recognition of an hedgehog receptor such as patched can be determined and used to generate hedgehog-derived peptidomimetics which competitively inhibit binding of the authentic hedgehog protein with that moiety. By employing, for example, scanning mutagenesis to map the amino acid residues of each of the subject hedgehog proteins which are involved in binding other extracellular proteins, peptidomimetic compounds can be generated which mimic those residues of the hedgehog protein which facilitate the interaction. Such mimetics may then be used to interfere with the normal function of a hedgehog protein. For instance, non-hydrolyzable peptide analogs of such residues can be generated using benzodiazepine (e.g., see Freidinger et al. in Peptides: Chemistry and Biology, G.R. Marshall ed., ESCOM Publisher: Leiden, Netherlands, 1988), azepine (e.g., see Huffman et al. in Peptides: Chemistry and Biology, G.R. Marshall ed., ESCOM Publisher: Leiden, Netherlands, 1988), substituted gama lactam rings (Garvey et al. in Peptides: Chemistry and Biology, G.R. Marshall ed., ESCOM Publisher: Leiden, Netherlands, 1988), keto-methylene pseudopeptides (Ewenson et al. (1986) J Med Chem 29:295; and Ewenson et al. in Peptides: Structure and Function (Proceedings of the 9th American Peptide Symposium) Pierce Chemical Co. Rockland, IL, 1985), \(\beta\)-turn dipeptide cores (Nagai et al. (1985) Tetrahedron Lett 26:647; and Sato et al. (1986) J Chem Soc Perkin Trans 1:1231), and β-aminoalcohols (Gordon et al. (1985) Biochem Biophys Res Commun126:419; and Dann et al. (1986) Biochem Biophys Res Commun 134:71).

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Recombinantly produced forms of the hedgehog proteins can be produced using, e.g, expression vectors containing a nucleic acid encoding a hedgehog polypeptide, operably linked to at least one transcriptional regulatory sequence. Operably linked is intended to mean that the nucleotide sequence is linked to a regulatory sequence in a manner which allows expression of the nucleotide sequence. Regulatory sequences are art-recognized and are selected to direct expression of a hedgehog polypeptide. Accordingly, the term transcriptional regulatory sequence includes promoters, enhancers and other expression control elements. Such regulatory sequences are described in Goeddel; Gene Expression Technology: Methods in Enzymology 185, Academic Press, San Diego, CA (1990). For instance, any of a wide variety of expression control sequences, sequences that control the expression of a DNA sequence when operatively linked to it, may be used in these vectors to express DNA sequences encoding hedgehog polypeptide. Such useful expression control sequences, include, for example, a viral LTR, such as the LTR of the Moloney murine leukemia virus, the early and late promoters of SV40, adenovirus or cytomegalovirus immediate early promoter, the lac system, the trp system, the TAC or TRC system, T7 promoter whose expression is directed by T7 RNA polymerase, the major operator and promoter regions of phage  $\lambda$ , the control regions for fd coat protein, the promoter for 3phosphoglycerate kinase or other glycolytic enzymes, the promoters of acid phosphatase, e.g., Pho5, the promoters of the yeast  $\alpha$ -mating factors, the polyhedron promoter of the baculovirus system and other sequences known to control the expression of genes of prokaryotic or eukaryotic cells or their viruses, and various combinations thereof. It should be understood that the design of the expression vector may depend on such factors as the choice of the host cell to be transformed and/or the type of protein desired to be expressed. Moreover, the vector's copy number, the ability to control that copy number and the expression of any other proteins encoded by the vector, such as antibiotic markers, should also be considered.

In addition to providing a ready source of hedgehog polypeptides for purification, the gene constructs of the present invention can also be used as a part of a gene therapy protocol to deliver nucleic acids encoding either an agonistic or antagonistic form of a *hedgehog* polypeptide. Thus, another aspect of the invention features expression vectors for *in vivo* transfection of a *hedgehog* polypeptide in particular cell types so as cause ectopic expression of a *hedgehog* polypeptide in an T lymphocytes or other cells associated therewith.

Formulations of such expression constructs may be administered in any biologically effective carrier, e.g. any formulation or composition capable of effectively delivering the recombinant gene to cells *in vivo*. Approaches include insertion of the hedgehog coding sequence in viral vectors including recombinant retroviruses, adenovirus, adeno-associated virus, and herpes simplex virus-1, or recombinant bacterial or eukaryotic plasmids. Viral vectors transfect

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cells directly; plasmid DNA can be delivered with the help of, for example, cationic liposomes (lipofectin) or derivatized (e.g. antibody conjugated), polylysine conjugates, gramacidin S, artificial viral envelopes or other such intracellular carriers, as well as direct injection of the gene construct or CaPO<sub>4</sub> precipitation carried out *in vivo*. It will be appreciated that because transduction of appropriate target cells represents the critical first step in gene therapy, choice of the particular gene delivery system will depend on such factors as the phenotype of the intended target and the route of administration, e.g. locally or systemically. Furthermore, it will be recognized that the particular gene construct provided for *in vivo* transduction of *hedgehog* expression are also useful for *in vitro* transduction of cells, such as for use in the *ex vivo* tissue culture systems described below.

A preferred approach for *in vivo* introduction of nucleic acid into a cell is by use of a viral vector containing nucleic acid, e.g. a cDNA, encoding the particular form of the *hedgehog* polypeptide desired. Infection of cells with a viral vector has the advantage that a large proportion of the targeted cells can receive the nucleic acid. Additionally, molecules encoded within the viral vector, e.g., by a cDNA contained in the viral vector, are expressed efficiently in cells which have taken up viral vector nucleic acid.

Retrovirus vectors and adeno-associated virus vectors are generally understood to be the recombinant gene delivery system of choice for the transfer of exogenous genes in vivo, particularly into humans. These vectors provide efficient delivery of genes into cells, and the transferred nucleic acids are stably integrated into the chromosomal DNA of the host. A major prerequisite for the use of retroviruses is to ensure the safety of their use, particularly with regard to the possibility of the spread of wild-type virus in the cell population. The development of specialized cell lines (termed "packaging cells") which produce only replication-defective retroviruses has increased the utility of retroviruses for gene therapy, and defective retroviruses are well characterized for use in gene transfer for gene therapy purposes (for a review see Miller, A.D. (1990) Blood 76:271). Thus, recombinant retrovirus can be constructed in which part of the retroviral coding sequence (gag, pol, env) has been replaced by nucleic acid encoding a hedgehog polypeptide and renders the retrovirus replication defective. The replication defective retrovirus is then packaged into virions which can be used to infect a target cell through the use of a helper virus by standard techniques. Protocols for producing recombinant retroviruses and for infecting cells in vitro or in vivo with such viruses can be found in Current Protocols in Molecular Biology, Ausubel, F.M. et al. (eds.) Greene Publishing Associates, (1989), Sections 9.10-9.14 and other standard laboratory manuals. Examples of suitable retroviruses include pLJ, pZIP, pWE and pEM which are well known to those skilled in the art. Examples of suitable packaging virus lines for preparing both ecotropic and amphotropic retroviral systems include

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Crip, Cre, 2 and Am. Retroviruses have been used to introduce a variety of genes into many different cell types, including T lymphocytes, in vitro and/or in vivo (see for example Eglitis, et al. (1985) Science 230:1395-1398; Danos and Mulligan (1988) Proc. Natl. Acad. Sci. USA 85:6460-6464; Wilson et al. (1988) Proc. Natl. Acad. Sci. USA 85:3014-3018; Armentano et al. (1990) Proc. Natl. Acad. Sci. USA 87:6141-6145; Huber et al. (1991) Proc. Natl. Acad. Sci. USA 88:8039-8043; Ferry et al. (1991) Proc. Natl. Acad. Sci. USA 88:8377-8381; Chowdhury et al. (1991) Science 254:1802-1805; van Beusechem et al. (1992) Proc. Natl. Acad. Sci. USA 89:7640-7644; Kay et al. (1992) Human Gene Therapy 3:641-647; Dai et al. (1992) Proc. Natl. Acad. Sci. USA 89:10892-10895; Hwu et al. (1993) J. Immunol. 150:4104-4115; U.S. Patent No. 4,868,116; U.S. Patent No. 4,980,286; PCT Application WO 89/07136; PCT Application WO 89/02468; PCT Application WO 89/05345; and PCT Application WO 92/07573).

Furthermore, it has been shown that it is possible to limit the infection spectrum of retroviruses and consequently of retroviral-based vectors, by modifying the viral packaging proteins on the surface of the viral particle (see, for example PCT publications WO93/25234 and WO94/06920). For instance, strategies for the modification of the infection spectrum of retroviral vectors include: coupling antibodies specific for cell surface antigens to the viral *env* protein (Roux et al. (1989) *PNAS* 86:9079-9083; Julan et al. (1992) *J. Gen Virol* 73:3251-3255; and Goud et al. (1983) *Virology* 163:251-254); or coupling cell surface receptor ligands to the viral *env* proteins (Neda et al. (1991) *J Biol Chem* 266:14143-14146). Coupling can be in the form of the chemical cross-linking with a protein or other variety (e.g. lactose to convert the *env* protein to an asialoglycoprotein), as well as by generating fusion proteins (e.g. single-chain antibody/*env* fusion proteins). This technique, while useful to limit or otherwise direct the infection to certain tissue types, can also be used to convert an ecotropic vector in to an amphotropic vector.

Moreover, use of retroviral gene delivery can be further enhanced by the use of tissue- or cell-specific transcriptional regulatory sequences which control expression of the *hedgehog* gene of the retroviral vector.

Another viral gene delivery system useful in the present method utilizes adenovirus-derived vectors. The genome of an adenovirus can be manipulated such that it encodes and expresses a gene product of interest but is inactivated in terms of its ability to replicate in a normal lytic viral life cycle. See for example Berkner et al. (1988) *BioTechniques* 6:616; Rosenfeld et al. (1991) *Science* 252:431-434; and Rosenfeld et al. (1992) *Cell* 68:143-155. Suitable adenoviral vectors derived from the adenovirus strain Ad type 5 dl324 or other strains of adenovirus (e.g., Ad2, Ad3, Ad7 etc.) are well known to those skilled in the art. Recombinant

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adenoviruses can be advantageous in certain circumstances in that they can be used to infect a wide variety of cell types, including T lymphocytes. Furthermore, the virus particle is relatively stable and amenable to purification and concentration, and as above, can be modified so as to affect the spectrum of infectivity. Additionally, introduced adenoviral DNA (and foreign DNA contained therein) is not integrated into the genome of a host cell but remains episomal, thereby avoiding potential problems that can occur as a result of insertional mutagenesis in situations where introduced DNA becomes integrated into the host genome (e.g., retroviral DNA). Moreover, the carrying capacity of the adenoviral genome for foreign DNA is large (up to 8 kilobases) relative to other gene delivery vectors (Berkner et al. cited supra; Haj-Ahmand and Graham (1986) J. Virol. 57:267). Most replication-defective adenoviral vectors currently in use and therefore favored by the present invention are deleted for all or parts of the viral E1 and E3 genes but retain as much as 80% of the adenoviral genetic material (see, e.g., Jones et al. (1979) Cell 16:683; Berkner et al., supra; and Graham et al. in Methods in Molecular Biology, E.J. Murray, Ed. (Humana, Clifton, NJ, 1991) vol. 7. pp. 109-127). Expression of the inserted hedgehog gene can be under control of, for example, the E1A promoter, the major late promoter (MLP) and associated leader sequences, the E3 promoter, or exogenously added promoter sequences.

In addition to viral transfer methods, such as those illustrated above, non-viral methods can also be employed to cause expression of a *hedgehog* polypeptide in the tissue of an animal. Most nonviral methods of gene transfer rely on normal mechanisms used by mammalian cells for the uptake and intracellular transport of macromolecules. In preferred embodiments, non-viral gene delivery systems of the present invention rely on endocytic pathways for the uptake of the *hedgehog* polypeptide gene by the targeted cell. Exemplary gene delivery systems of this type include liposomal derived systems, poly-lysine conjugates, and artificial viral envelopes.

In clinical settings, the gene delivery systems for the therapeutic *hedgehog* gene can be introduced into a patient by any of a number of methods, each of which is familiar in the art. For instance, a pharmaceutical preparation of the gene delivery system can be introduced systemically, e.g. by intravenous injection, and specific transduction of the protein in the target cells occurs predominantly from specificity of transfection provided by the gene delivery vehicle, cell-type or tissue-type expression due to the transcriptional regulatory sequences controlling expression of the receptor gene, or a combination thereof. In other embodiments, initial delivery of the recombinant gene is more limited with introduction into the animal being quite localized. For example, the gene delivery vehicle can be introduced by catheter (see U.S. Patent 5,328,470) or by stereotactic injection (e.g. Chen et al. (1994) *PNAS* 91: 3054-3057). A *hedgehog* expression construct can be delivered in a gene therapy construct to dermal cells by, e.g.,

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electroporation using techniques described, for example, by Dev et al. ((1994) Cancer Treat Rev 20:105-115).

The pharmaceutical preparation of the gene therapy construct can consist essentially of the gene delivery system in an acceptable diluent, or can comprise a slow release matrix in which the gene delivery vehicle is imbedded. Alternatively, where the complete gene delivery system can be produced intact from recombinant cells, e.g. retroviral vectors, the pharmaceutical preparation can comprise one or more cells which produce the gene delivery system.

In yet another embodiment, the hedgehog or ptc therapeutic can be a "gene activation" construct which, by homologous recombination with a genomic DNA, alters the transcriptional regulatory sequences of an endogenous gene. For instance, the gene activation construct can replace the endogenous promoter of a *hedgehog* gene with a heterologous promoter, e.g., one which causes consitutive expression of the *hedgehog* gene or which causes inducible expression of the gene under conditions different from the normal expression pattern of the gene. Other genes in the *patched* signaling pathway can be similarly targeted. A vareity of different formats for the gene activation constructs are available. See, for example, the Transkaryotic Therapies, Inc PCT publications WO93/09222, WO95/31560, WO96/29411, WO95/31560 and WO94/12650.

In preferred embodiments, the nucleotide sequence used as the gene activation construct can be comprised of (1) DNA from some portion of the endogenous *hedgehog* gene (exon sequence, intron sequence, promoter sequences, etc.) which direct recombination and (2) heterologous transcriptional regulatory sequence(s) which is to be operably linked to the coding sequence for the genomic *hedgehog* gene upon recombination of the gene activation construct. For use in generating cultures of *hedgehog* producing cells, the construct may further include a reporter gene to detect the presence of the knockout construct in the cell.

The gene activation construct is inserted into a cell, and integrates with the genomic DNA of the cell in such a position so as to provide the heterologous regulatory sequences in operative association with the native *hedgehog* gene. Such insertion occurs by homologous recombination, i.e., recombination regions of the activation construct that are homologous to the endogenous *hedgehog* gene sequence hybridize to the genomic DNA and recombine with the genomic sequences so that the construct is incorporated into the corresponding position of the genomic DNA.

The terms "recombination region" or "targeting sequence" refer to a segment (i.e., a portion) of a gene activation construct having a sequence that is substantially identical to or substantially complementary to a genomic gene sequence, e.g., including 5' flanking sequences

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of the genomic gene, and can facilitate homologous recombination between the genomic sequence and the targeting transgene construct.

As used herein, the term "replacement region" refers to a portion of a activation construct which becomes integrated into an endogenous chromosomal location following homologous recombination between a recombination region and a genomic sequence.

The heterologous regulatory sequences, e.g., which are provided in the replacement region, can include one or more of a variety elements, including: promoters (such as constitutive or inducible promoters), enhancers, negative regualtory elements, locus control regions, transcription factor binding sites, or combinations thereof. Promoters/enhancers which may be used to control the expression of the targeted gene in vivo include, but are not limited to, the cytomegalovirus (CMV) promoter/enhancer (Karasuyama et al., 1989, J. Exp. Med., 169:13), the human β-actin promoter (Gunning et al. (1987) PNAS 84:4831-4835), the glucocorticoidinducible promoter present in the mouse mammary tumor virus long terminal repeat (MMTV LTR) (Klessig et al. (1984) Mol. Cell Biol. 4:1354-1362), the long terminal repeat sequences of Moloney murine leukemia virus (MuLV LTR) (Weiss et al. (1985) RNA Tumor Viruses, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York), the SV40 early or late region promoter (Bernoist et al. (1981) Nature 290:304-310; Templeton et al. (1984) Mol. Cell Biol., 4:817; and Sprague et al. (1983) J. Virol., 45:773), the promoter contained in the 3' long terminal repeat of Rous sarcoma virus (RSV) (Yamamoto et al., 1980, Cell, 22:787-797), the herpes simplex virus (HSV) thymidine kinase promoter/enhancer (Wagner et al. (1981) PNAS 82:3567-71), and the herpes simplex virus LAT promoter (Wolfe et al. (1992) Nature Genetics, 1:379-384).

In an exemplary embodiment, portions of the 5' flanking region of the human Shh gene are amplified using primers which add restriction sites, to generate the following fragments

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As illustrated, primer 1 includes a 5' non-coding region of the human Shh gene and is flanked by an AsuII and ClaI restriction sites. Primer 2 includes a portion of the 5' non-coding region

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immediately 3' to that present in primer 1. The hedgehog gene sequence is flanked by XhoII and BamHI restriction sites. The purified amplimers are cut with each of the enzymes as appropriate.

The vector pCDNA1.1 (Invitrogen) includes a CMV promoter. The plasmid is cut with with AsuII, which cleaves just 3' to the CMV promoter sequence. The AsuII/ClaI fragment of primer 1 is ligated to the AsuII cleavage site of the pcDNA vector. The ClaI/AsuII ligation destroys the AsuII site at the 3' end of a properly inserted primer 1.

The vector is then cut with BamHI, and an XhoII/BamHI fragment of primer 2 is ligated to the BamHI cleavage site. As above, the BamHI/XhoII ligation destroys the BamHI site at the 5' end of a properly inserted primer 2.

Individual colonies are selected, cut with AsuII and BamHI, and the size of the AsuII/BamHI fragment determined. Colonies in which both the primer 1 and primer 2 sequences are correctly inserted are further amplified, an cut with AsuII and BamHI to produce the gene activation construct

cqaagcgaggcagccagcgagggagagagggggggggagccggagcgaggaaATCGAAGGTTC GAATCCTTCCCCCACCACCATCACTTTCAAAAGTCCGAAAGAATCTGCTCCCTGCTTGTGTTTG GAGGTCGCTGAGTAGTGCGCGAGTAAAATTTAAGCTACAACAAGGCAAGGCTTGACCGACAATTG CATGAAGAATCTGCTTAGGGTTAGGCGTTTTGCGCTGCTTCGCGATGTACGGGCCAGATATACGC GTTGACATTGATTATTGACTAGTTATTAATAGTAATCAATTACGGGGTCATTAGTTCATAGCCCA TATATGGAGTTCCGCGTTACATAACTTACGGTAAATGGCCCGCCTGGCTGACCGCCCAACGACCC  $\tt CCGCCCATTGACGTCAATAATGACGTATGTTCCCATAGTAACGCCAATAGGGACTTTCCATTGAC$ GTCAATGGGTGGACTATTTACGGTAAACTGCCCACTTGGCAGTACATCAAGTGTATCATATGCCA AGTACGCCCCTATTGACGTCAATGACGGTAAATGGCCCGCCTGGCATTATGCCCAGTACATGAC  $\tt CTTATGGGACTTTCCTACTTGGCAGTACATCTACGTATTAGTCATCGCTATTACCATGGTGATGC$  ${\tt GGTTTTGGCAGTACATCAATGGGCGTGGATAGCGGTTTGACTCACGGGGATTTCCAAGTCTCCACCGGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCCACCGGGGATTTCCAAGTCTCACCGGGGATTTCCAAGTCTCACCGGGGATTTCCAAGTCTCACCGGGGATTTCCAAGTCTCACCGGGGATTTCCAAGTCTCACCGGGGATTTCCAAGTCTCACCGGGGATTTCCAAGTCTCACCGGGGATTTCCAAGTCTCACCGGGGATTTCCAAGTCTCACCGGGGATTTCCAAGTCTCACCGGGGATTTCCAAGTCTCACCGGGGATTTCCAAGTCTCACCGGGGATTTCCAAGTCTCACCGGGGATTTCCAAGTCTCACCGGGGATTTCCAAGTCTCACCGGGGATTTCCAAGTCACCGGGGATTTCCAAGTCACCGGGGATTTCCAAGTCTCACCGGGGATTTCCAAGTCTCACCGGGGATTTCCAAGTCTCACAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTCAAGTC$ CCCATTGACGTCAATGGGAGTTTGTTTTGGCACCAAAATCAACGGGACTTTCCAAAATGTCGTAA CAACTCCGCCCCATTGACGCAAATGGGCGGTAGGCGTGTACGGTGGGAGGTCTATATAAGCAGAG CTCTCTGGCTAACTAGAGAACCCACTGCTTACTGGCTTATCGAAATTAATACGACTCACTATAGG GAGACCCAAGCTTGGTACCGAGCTCGGATCqatctqqqaaaqcqcaagagagagcgcacacgcac acacccgccgcgcgcactcgg

In this construct, the flanking primer 1 and primer 2 sequences provide the recombination region which permits the insertion of the CMV promoter in front of the coding sequence for the human *Shh* gene. Other heterologous promoters (or other transcriptional regulatory sequences) can be inserted in a genomic *hedgehog* gene by a similar method.

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In still other embodiments, the replacement region merely deletes a negative transcriptional control element of the native gene, e.g., to activate expression, or ablates a positive control element, e.g., to inhibit expression of the targeted gene.

#### V. Exemplary ptc therapeutic compounds.

In another embodiment, the subject method is carried out using a ptc therapeutic composition. Such compositions can be generated with, for example, compounds which bind to patched and alter its signal transduction activity, compounds which alter the binding and/or enzymatic activity of a protein (e.g., intracellular) involved in patched signal pathway, and compounds which alter the level of expression of a hedgehog protein, a patched protein or a protein involved in the intracellular signal transduction pathway of patched.

The availability of purified and recombinant *hedgehog* polypeptides facilitates the generation of assay systems which can be used to screen for drugs, such as small organic molecules, which are either agonists or antagonists of the normal cellular function of a *hedgehog* and/or patched protein, particularly their role in the pathogenesis of peripheral nerve proliferation and/or differentiation. In one embodiment, the assay evaluates the ability of a compound to modulate binding between a *hedgehog* polypeptide and a *hedgehog* receptor such as *patched*. In other embodiments, the assay merely scores for the ability of a test compound to alter the signal transduction actitity of the *patched* protein. In this manner, a variety of *hedgehog* and/or *ptc* therapeutics, both agonists (inhibit T-cell maturation) and antagonist (promote T-cell maturation) can be identified. A variety of assay formats will suffice and, in light of the present disclosure, will be comprehended by skilled artisan.

In many drug screening programs which test libraries of compounds and natural extracts, high throughput assays are desirable in order to maximize the number of compounds surveyed in a given period of time. Assays which are performed in cell-free systems, such as may be derived with purified or semi-purified proteins, are often preferred as "primary" screens in that they can be generated to permit rapid development and relatively easy detection of an alteration in a molecular target which is mediated by a test compound. Moreover, the effects of cellular toxicity and/or bioavailability of the test compound can be generally ignored in the *in vitro* system, the assay instead being focused primarily on the effect of the drug on the molecular target as may be manifest in an alteration of binding affinity with receptor proteins.

Acordingly, in an exemplary screening assay for ptc therapeutics, the compound of interest is contacted with a mixture including a hedgehog receptor protein (e.g., a cell expressing

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the *patched* receptor) and a hedgehog protein under conditions in which it is ordinarily capable of binding the *hedgehog* protein. To the mixture is then added a composition containing a test compound. Detection and quantification of receptor/*hedgehog* complexes provides a means for determining the test compound's efficacy at inhibiting (or potentiating) complex formation between the receptor protein and the *hedgehog* polypeptide. The efficacy of the compound can be assessed by generating dose response curves from data obtained using various concentrations of the test compound. Moreover, a control assay can also be performed to provide a baseline for comparison. In the control assay, isolated and purified *hedgehog* polypeptide is added to the receptor protein, and the formation of receptor/*hedgehog* complex is quantitated in the absence of the test compound.

In other embodiments, a ptc therapeutic of the present invention is one which disrupts the association of *patched* with *smoothened*.

Agonist and antagonists of peripheral nerve maintanence can be distinguished, and the efficacy of the compound can be assessed, by subsequent testing with T lymphocytes, e.g., in culture.

In an illustrative embodiment, the polypeptide utilized as a hedgehog receptor can be generated from the patched protein. Accordingly, an exemplary screening assay includes all or a suitable portion of the patched protein which can be obtained from, for example, the human patched gene (GenBank U43148) or other vertebrate sources (see GenBank Accession numbers U40074 for chicken patched and U46155 for mouse patched), as well as from drosophila (GenBank Accession number M28999) or other invertebrate sources. The patched protein can be provided in the screening assay as a whole protein (preferably expressed on the surface of a cell), or alternatively as a fragment of the full length protein which binds to hedgehog polypeptides, e.g., as one or both of the substantial extracellular domains (e.g. corresponding to residues Asn120-Ser438 and/or Arg770-Trp1027 of the human patched protein - which are also potential antagonists of hedgehog-dependent signal transduction). For instance, the patched protein can be provided in soluble form, as for example a preparation of one of the extracellular domains, or a preparation of both of the extracellular domains which are covalently connected by an unstructured linker (see, for example, Huston et al. (1988) PNAS 85:4879; and U.S. Patent No. 5,091,513). In other embodiments, the protein can be provided as part of a liposomal preparation or expressed on the surface of a cell. The patched protein can derived from a recombinant gene, e.g., being ectopically expressed in a heterologous cell. For instance, the protein can be expressed on oocytes, mammalian cells (e.g., COS, CHO, 3T3 or the like), or yeast cell by standard recombinant DNA techniques. These recombinant cells can be used for

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receptor binding, signal transduction or gene expression assays. Marigo et al. (1996) Development 122:1225-1233 illustrates a binding assay of human hedgehog to chick patched protein ectopically expressed in Xenopus laevis oocytes. The assay system of Marigo et al. can be adapted to the present drug screening assays. As illustrated in that reference, Shh binds to the patched protein in a selective, saturable, dose-dependent manner, thus demonstrating that patched is a receptor for Shh.

Complex formation between the *hedgehog* polypeptide and a *hedgehog* receptor may be detected by a variety of techniques. For instance, modulation of the formation of complexes can be quantitated using, for example, detectably labelled proteins such as radiolabelled, fluorescently labelled, or enzymatically labelled *hedgehog* polypeptides, by immunoassay, or by chromatographic detection.

Typically, for cell-free assays, it will be desirable to immobilize either the *hedgehog* receptor or the *hedgehog* polypeptide to facilitate separation of receptor/*hedgehog* complexes from uncomplexed forms of one of the proteins, as well as to accommodate automation of the assay. In one embodiment, a fusion protein can be provided which adds a domain that allows the protein to be bound to a matrix. For example, glutathione-S-transferase/receptor (GST/receptor) fusion proteins can be adsorbed onto glutathione sepharose beads (Sigma Chemical, St. Louis, MO) or glutathione derivatized microtitre plates, which are then combined with the *hedgehog* polypeptide, e.g. an <sup>35</sup>S-labeled *hedgehog* polypeptide, and the test compound and incubated under conditions conducive to complex formation, e.g. at physiological conditions for salt and pH, though slightly more stringent conditions may be desired. Following incubation, the beads are washed to remove any unbound *hedgehog* polypeptide, and the matrix bead-bound radiolabel determined directly (e.g. beads placed in scintillant), or in the supernatant after the receptor/*hedgehog* complexes are dissociated. Alternatively, the complexes can be dissociated from the bead, separated by SDS-PAGE gel, and the level of *hedgehog* polypeptide found in the bead fraction quantitated from the gel using standard electrophoretic techniques.

Other techniques for immobilizing proteins on matrices are also available for use in the subject assay. For instance, soluble portions of the *hedgehog* receptor protein can be immobilized utilizing conjugation of biotin and streptavidin. For instance, biotinylated receptor molecules can be prepared from biotin-NHS (N-hydroxy-succinimide) using techniques well known in the art (e.g., biotinylation kit, Pierce Chemicals, Rockford, IL), and immobilized in the wells of streptavidin-coated 96 well plates (Pierce Chemical). Alternatively, antibodies reactive with *hedgehog* receptor but which do not interfere with *hedgehog* binding can be derivatized to the wells of the plate, and the receptor trapped in the wells by antibody conjugation. As

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above, preparations of a *hedgehog* polypeptide and a test compound are incubated in the receptor-presenting wells of the plate, and the amount of receptor/*hedgehog* complex trapped in the well can be quantitated. Exemplary methods for detecting such complexes, in addition to those described above for the GST-immobilized complexes, include immunodetection of complexes using antibodies reactive with the *hedgehog* polypeptide, or which are reactive with the receptor protein and compete for binding with the *hedgehog* polypeptide; as well as enzymelinked assays which rely on detecting an enzymatic activity associated with the *hedgehog* polypeptide. In the instance of the latter, the enzyme can be chemically conjugated or provided as a fusion protein with the *hedgehog* polypeptide. To illustrate, the *hedgehog* polypeptide can be chemically cross-linked or genetically fused with alkaline phosphatase, and the amount of *hedgehog* polypeptide trapped in the complex can be assessed with a chromogenic substrate of the enzyme, e.g. paranitrophenylphosphate. Likewise, a fusion protein comprising the *hedgehog* polypeptide and glutathione-S-transferase can be provided, and complex formation quantitated by detecting the GST activity using 1-chloro-2,4-dinitrobenzene (Habig et al (1974) *J Biol Chem* 249:7130).

For processes which rely on immunodetection for quantitating one of the proteins trapped in the complex, antibodies against the protein, such as the anti-hedgehog antibodies described herein, can be used. Alternatively, the protein to be detected in the complex can be "epitope tagged" in the form of a fusion protein which includes, in addition to the hedgehog polypeptide or hedgehog receptor sequence, a second polypeptide for which antibodies are readily available (e.g. from commercial sources). For instance, the GST fusion proteins described above can also be used for quantification of binding using antibodies against the GST moiety. Other useful epitope tags include myc-epitopes (e.g., see Ellison et al. (1991) J Biol Chem 266:21150-21157) which includes a 10-residue sequence from c-myc, as well as the pFLAG system (International Biotechnologies, Inc.) or the pEZZ-protein A system (Pharamacia, NJ).

Where the desired portion of the *hedgehog* receptor (or other *hedgehog* binding molecule) cannot be provided in soluble form, liposomal vesicles can be used to provide manipulatable and isolatable sources of the receptor. For example, both authentic and recombinant forms of the *patched* protein can be reconstituted in artificial lipid vesicles (e.g. phosphatidylcholine liposomes) or in cell membrane-derived vesicles (see, for example, Bear et al. (1992) *Cell* 68:809-818; Newton et al. (1983) *Biochemistry* 22:6110-6117; and Reber et al. (1987) *J Biol Chem* 262:11369-11374).

In addition to cell-free assays, such as described above, the readily available source of hedgehog proteins provided by the art also facilitates the generation of cell-based assays for

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identifying small molecule agonists/antagonists and the like. Analogous to the cell-based assays described above for screening combinatorial libraries, cells which are sensitive to *hedgehog* induction, e.g. *patched*-expressing cells or other myoblast-derived cells sensitive to *hedgehog* induction, can be contacted with a *hedgehog* protein and a test agent of interest, with the assay scoring for anything from simple binding to the cell to modulation in *hedgehog* inductive responses by the target cell in the presence and absence of the test agent. As with the cell-free assays, agents which produce a statistically significant change in *hedgehog* activities (either inhibition or potentiation) can be identified.

In other emdodiments, the cell-based assay scores for agents which disrupt association of patched and *smoothened* proteins, e.g., in the cell surface membrane or liposomal preparation.

In addition to characterizing cells that naturally express the *patched* protein, cells which have been genetically engineered to ectopically express *patched* can be utilized for drug screening assays. As an example, cells which either express low levels or lack expression of the *patched* protein, e.g. *Xenopus laevis* oocytes, COS cells or yeast cells, can be genetically modified using standard techniques to ectopically express the *patched* protein. (see Marigo et al., *supra*).

The resulting recombinant cells, e.g., which express a functional patched receptor, can be utilized in receptor binding assays to identify agonist or anatagonsts of hedgehog binding. Binding assays can be performed using whole cells. Furthermore, the recombinant cells of the present invention can be engineered to include other heterolgous genes encoding proteins involved in hedgehog-dependent siganl pathways. For example, the gene products of one or more of smoothened, costal-2 and/or fused can be co-expressed with patched in the reagent cell, with assays being sensitive to the functional reconstituion of the hedgehog signal transduction cascade.

Alternatively, liposomal preparations using reconstituted patched protein can be utilized. Patched protein purified from detergent extracts from both authentic and recombinant origins can be reconstituted in in artificial lipid vesicles (e.g. phosphatidylcholine liposomes) or in cell membrane-derived vesicles (see, for example, Bear et al. (1992) Cell 68:809-818; Newton et al. (1983) Biochemistry 22:6110-6117; and Reber et al. (1987) J Biol Chem 262:11369-11374). The lamellar structure and size of the resulting liposomes can be characterized using electron microscopy. External orientation of the patched protein in the reconstituted membranes can be demonstrated, for example, by immunoelectron microscopy. The hedgehog protein binding activity of liposomes containing patched and liposomes without the protein in the presence of

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candidate agents can be compared in order to identify potential modulators of the *hedgehog-patched* interaction.

The *hedgehog* protein used in these cell-based assays can be provided as a purified source (natural or recombinant in origin), or in the form of cells/tissue which express the protein and which are co-cultured with the target cells. As in the cell-free assays, where simple binding (rather than induction) is the hedgehog activity scored for in the assay, the protein can be labelled by any of the above-mentioned techniques, e.g., fluorescently, enzymatically or radioactively, or detected by immunoassay.

In addition to binding studies, functional assays can be used to identified modulators, i.e., agonists or antagonists, of *hedgehog* or *patched* activities. By detecting changes in intracellular signals, such as alterations in second messengers or gene expression, in *patched*-expressing cells contacted with a test agent, candidate agonists and antagonists to *patched* signaling can be identified.

A number of gene products have been implicated in *patched*-mediated signal transduction, including *patched*, the transcription factor *cubitus interruptus* (ci), the serine/threonine kinase *fused* (fu) and the gene products of *costal-2*, *smoothened* and *suppressor* of *fused*.

The interaction of a hedgehog protein with patched sets in motion a cascade involving the activation and inhibition of downstream effectors, the ultimate consequence of which is, in some instances, a detectable change in the transcription or translation of a gene. transcriptional targets of patched signaling are the patched gene itself (Hidalgo and Ingham, 1990 Development 110, 291-301; Marigo et al., 1996 ) and the vertebrate homologs of the drosophila cubitus interruptus gene, the GLI genes (Hui et al. (1994) Dev Biol 162:402-413). Patched gene expression has been shown to be induced in cells of the limb bud and the neural plate that are responsive to Shh. (Marigo et al. (1996) PNAS, in press; Marigo et al. (1996) Development 122:1225-1233). The GLI genes encode putative transcription factors having zinc finger DNA binding domains (Orenic et al. (1990) Genes & Dev 4:1053-1067; Kinzler et al. (1990) Mol Cell Biol 10:634-642). Transcription of the GLI gene has been reported to be upregulated in response to hedgehog in limb buds, while transcription of the GLI3 gene is downregulated in response to hedgehog induction (Marigo et al. (1996) Development 122:1225-1233). By selecting transcriptional regulatory sequences from such target genes, e.g. from patched or GLI genes, that are responsible for the up- or down regulation of these genes in response to patched signalling, and operatively linking such promoters to a reporter gene, one can derive a transcription based assay which is sensitive to the ability of a specific test compound

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to modify *patched* signalling pathways. Expression of the reporter gene, thus, provides a valuable screening tool for the development of compounds that act as agonists or antagonists of *ptc* induction of differentiation/quiescence.

Reporter gene based assays of this invention measure the end stage of the above described cascade of events, e.g., transcriptional modulation. Accordingly, in practicing one embodiment of the assay, a reporter gene construct is inserted into the reagent cell in order to generate a detection signal dependent on *ptc* signaling. To identify potential regulatory elements responsive to *ptc* signaling present in the transcriptional regulatory sequence of a target gene, nested deletions of genomic clones of the target gene can be constructed using standard techniques. See, for example, <u>Current Protocols in Molecular Biology</u>, Ausubel, F.M. et al. (eds.) Greene Publishing Associates, (1989); U.S. Patent 5,266,488; Sato et al. (1995) *J Biol Chem* 270:10314-10322; and Kube et al. (1995) *Cytokine* 7:1-7. A nested set of DNA fragments from the gene's 5'-flanking region are placed upstream of a reporter gene, such as the luciferase gene, and assayed for their ability to direct reporter gene expression in *patched* expressing cells. Host cells transiently transfected with reporter gene constructs can be scored for the induction of expression of the reporter gene in the presence and absence of *hedgehog* to determine regulatory sequences which are responsice to *patched*-dependent signalling.

In practicing one embodiment of the assay, a reporter gene construct is inserted into the reagent cell in order to generate a detection signal dependent on second messengers generated by induction with *hedgehog* protein. Typically, the reporter gene construct will include a reporter gene in operative linkage with one or more transcriptional regulatory elements responsive to the *hedgehog* activity, with the level of expression of the reporter gene providing the *hedgehog*-dependent detection signal. The amount of transcription from the reporter gene may be measured using any method known to those of skill in the art to be suitable. For example, mRNA expression from the reporter gene may be detected using RNAse protection or RNA-based PCR, or the protein product of the reporter gene may be identified by a characteristic stain or an intrinsic activity. The amount of expression from the reporter gene is then compared to the amount of expression in either the same cell in the absence of the test compound (or *hedgehog*) or it may be compared with the amount of transcription in a substantially identical cell that lacks the target receptor protein. Any statistically or otherwise significant difference in the amount of transcription indicates that the test compound has in some manner altered the signal transduction of the *patched* protein, e.g., the test compound is a potential ptc therapeutic.

As described in further detail below, in preferred embodiments the gene product of the reporter is detected by an intrinsic activity associated with that product. For instance, the reporter gene may encode a gene product that, by enzymatic activity, gives rise to a detection

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signal based on color, fluorescence, or luminescence. In other preferred embodiments, the reporter or marker gene provides a selective growth advantage, e.g., the reporter gene may enhance cell viability, relieve a cell nutritional requirement, and/or provide resistance to a drug.

Preferred reporter genes are those that are readily detectable. The reporter gene may also be included in the construct in the form of a fusion gene with a gene that includes desired transcriptional regulatory sequences or exhibits other desirable properties. Examples of reporter genes include, but are not limited to CAT (chloramphenicol acetyl transferase) (Alton and Vapnek (1979), Nature 282: 864-869) luciferase, and other enzyme detection systems, such as beta-galactosidase; firefly luciferase (deWet et al. (1987), Mol. Cell. Biol. 7:725-737); bacterial luciferase (Engebrecht and Silverman (1984), PNAS 1: 4154-4158; Baldwin et al. (1984), Biochemistry 23: 3663-3667); alkaline phosphatase (Toh et al. (1989) Eur. J. Biochem. 182: 231-238, Hall et al. (1983) J. Mol. Appl. Gen. 2: 101), human placental secreted alkaline phosphatase (Cullen and Malim (1992) Methods in Enzymol. 216:362-368).

Transcriptional control elements which may be included in a reporter gene construct include, but are not limited to, promoters, enhancers, and repressor and activator binding sites. Suitable transcriptional regulatory elements may be derived from the transcriptional regulatory regions of genes whose expression is induced after modulation of a *patched* signal transduction pathway. The characteristics of preferred genes from which the transcriptional control elements are derived include, but are not limited to, low or undetectable expression in quiescent cells, rapid induction at the transcriptional level within minutes of extracellular simulation, induction that is transient and independent of new protein synthesis, subsequent shut-off of transcription requires new protein synthesis, and mRNAs transcribed from these genes have a short half-life. It is not necessary for all of these properties to be present.

In yet other embodiments, second messenger generation can be measured directly in the detection step, such as mobilization of intracellular calcium, phospholipid metabolism or adenylate cyclase activity are quantitated, for instance, the products of phospholipid hydrolysis IP<sub>3</sub>, DAG or cAMP could be measured For example, recent studies have implicated protein kinase A (PKA) as a possible component of hedgehog/patched signaling (Hammerschmidt et al. (1996) Genes & Dev 10:647). High PKA activity has been shown to antagonize hedgehog signaling in these systems. Although it is unclear whether PKA acts directly downstream or in parallel with hedgehog signaling, it is possible that hedgehog signalling occurs via inhibition of PKA activity. Thus, detection of PKA activity provides a potential readout for the instant assays.

In a preferred embodiment, the ptc therapeutic is a PKA inhibitor. A variety of PKA inhibitors are known in the art, including both peptidyl and organic compounds. For instance,

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the *ptc* therapeutic can be a 5-isoquinolinesulfonamide, such as represented in the general formula:

wherein,

 $R_1$  and  $R_2$  each can independently represent hydrogen, and as valence and stability permit a lower alkyl, a lower alkenyl, a lower alkynyl, a carbonyl (such as a carboxyl, an ester, a formate, or a ketone), a thiocarbonyl (such as a thioester, a thioacetate, or a thioformate), an amino, an acylamino, an amido, a cyano, a nitro, an azido, a sulfate, a sulfonate, a sulfonamido,  $-(CH_2)_m-R_8$ ,  $-(CH_2)_m-OH$ 

R<sub>1</sub> and R<sub>2</sub> taken together with N form a heterocycle (substituted or unsubstituted);

 $R_3$  is absent or represents one or more substitutions to the isoquinoline ring such as a lower alkyl, a lower alkenyl, a lower alkynyl, a carbonyl (such as a carboxyl, an ester, a formate, or a ketone), a thiocarbonyl (such as a thioester, a thioacetate, or a thioformate), an amino, an acylamino, an amido, a cyano, a nitro, an azido, a sulfate, a sulfonate, a sulfonamido,  $-(CH_2)_m$ - $R_8$ ,  $-(CH_2)_m$ -O-lower alkyl,  $-(CH_2)_m$ -O-lower alkenyl,  $-(CH_2)_m$ -O-( $CH_2)_m$ - $R_8$ ,  $-(CH_2)_m$ -S-lower alkyl,  $-(CH_2)_m$ -S-lower alkenyl,  $-(CH_2)_n$ -S-( $CH_2)_m$ - $R_8$ ;

R<sub>8</sub> represents a substituted or unsubstituted aryl, aralkyl, cycloalkyl, cycloalkenyl, or heterocycle; and

n and m are independently for each occurrence zero or an integer in the range of 1 to 6. In a preferred embodiment, the PKA inhibitor is N-[2-((p-bromocinnamyl)amino)ethyl]-5-isoquinolinesulfonamide (H-89; Calbiochem Cat. No. 371963), e.g., having the formula:

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In another embodiment, the PKA inhibitor is 1-(5-isoquinolinesulfonyl)-2-methylpiperazine (H-7; Calbiochem Cat. No. 371955), e.g., having the formula:

In still other embodiments, the PKA inhibitor is KT5720 (Calbiochem Cat. No. 420315), having the structure

A variety of nucleoside analogs are also useful as PKA inhibitors. For example, the subject method can be carried out cyclic AMP analogs which inhibit the kinase activity of PKA, as for example, 8-bromo-cAMP or dibutyryl-cAMP

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Exemplary peptidyl inhibitors of PKA activity include the PKA Heat Stable Inhibitor (isoform  $\alpha$ ; see, for example, Calbiochem Cat. No. 539488, and Wen et al. (1995) *J Biol Chem* 270:2041).

Certain *hedehog* receptors may stimulate the activity of phospholipases. Inositol lipids can be extracted and analyzed using standard lipid extraction techniques. Water soluble derivatives of all three inositol lipids (IP<sub>1</sub>, IP<sub>2</sub>, IP<sub>3</sub>) can also be quantitated using radiolabelling techniques or HPLC.

The mobilization of intracellular calcium or the influx of calcium from outside the cell may be a response to *hedgehog* stimulation or lack there of. Calcium flux in the reagent cell can be measured using standard techniques. The choice of the appropriate calcium indicator, fluorescent, bioluminescent, metallochromic, or Ca<sup>++</sup>-sensitive microelectrodes depends on the cell type and the magnitude and time constant of the event under study (Borle (1990) *Environ Health Perspect* 84:45-56). As an exemplary method of Ca<sup>++</sup> detection, cells could be loaded with the Ca<sup>++</sup> sensitive fluorescent dye fura-2 or indo-1, using standard methods, and any change in Ca<sup>++</sup> measured using a fluorometer.

In certain embodiments of the assay, it may be desirable to screen for changes in cellular phosphorylation. As an example, the drosophila gene *fused* (fu) which encodes a serine/threonine kinase has been identified as a potential downstream target in *hedgehog* signaling. (Preat et al., 1990 *Nature* 347, 87-89; Therond et al. 1993, *Mech. Dev.* 44. 65-80). The ability of compounds to modulate serine/threonine kinase activation could be screened using colony immunoblotting (Lyons and Nelson (1984) *Proc. Natl. Acad. Sci. USA* 81:7426-7430) using antibodies against phosphorylated serine or threonine residues. Reagents for performing such assays are commercially available, for example, phosphoserine and phosphothreonine specific antibodies which measure increases in phosphorylation of those residues can be purchased from comercial sources.

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In yet another embodiment, the *ptc* therapeutic is an antisense molecule which inhibits expression of a protein involved in a *patched*-mediated signal transduction pathway. To illustrate, by inhibiting the expression of a protein which are involved in *patched* signals, such as fused, costal-2, smoothened and/or Gli genes, the ability of the patched signal pathway(s) to inhibit proliferation of a cell can be altered, e.g., potentiated or repressed.

As used herein, "antisense" therapy refers to administration or *in situ* generation of oligonucleotide probes or their derivatives which specifically hybridize (e.g. bind) under cellular conditions with cellular mRNA and/or genomic DNA encoding a *hedgehog* protein, patched, or a protein involved in patched-mediated signal transduction. The hybridization should inhibit expression of that protein, e.g. by inhibiting transcription and/or translation. The binding may be by conventional base pair complementarity, or, for example, in the case of binding to DNA duplexes, through specific interactions in the major groove of the double helix. In general, "antisense" therapy refers to the range of techniques generally employed in the art, and includes any therapy which relies on specific binding to oligonucleotide sequences.

An antisense construct of the present invention can be delivered, for example, as an expression plasmid which, when transcribed in the cell, produces RNA which is complementary to at least a unique portion of the target cellular mRNA. Alternatively, the antisense construct is an oligonucleotide probe which is generated *ex vivo* and which, when introduced into the cell causes inhibition of expression by hybridizing with the mRNA and/or genomic sequences of a target gene. Such oligonucleotide probes are preferably modified oligonucleotide which are resistant to endogenous nucleases, e.g. exonucleases and/or endonucleases, and is therefore stable *in vivo*. Exemplary nucleic acid molecules for use as antisense oligonucleotides are phosphoramidate, phosphothioate and methylphosphonate analogs of DNA (see also U.S. Patents 5,176,996; 5,264,564; and 5,256,775). Additionally, general approaches to constructing oligomers useful in antisense therapy have been reviewed, for example, by Van der Krol et al. (1988) *Biotechniques* 6:958-976; and Stein et al. (1988) *Cancer Res* 48:2659-2668.

Several considerations should be taken into account when constructing antisense oligonucleotides for the use in the methods of the invention: (1) oligos should have a GC content of 50% or more; (2) avoid sequences with stretches of 3 or more G's; and (3) oligonucleotides should not be longer than 25-26 mers. When testing an antisense oligonucleotide, a mismatched control can be constructed. The controls can be generated by reversing the sequence order of the corresponding antisense oligonucleotide in order to conserve the same ratio of bases.

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In an illustrative embodiment, the *ptc* therapeutic can be an antisense construct for inhibiting the expression of *patched*, e.g., to mimic the inhibition of *patched* by *hedgehog*. Exemplary antisense constructs include:

- 5'-GTCCTGGCGCCGCCGCCGTCGCC
- 5 5'-TTCCGATGACCGGCCTTTCGCGGTGA
  - 5'-GTGCACGGAAAGGTGCAGGCCACACT

#### VI. Exemplary pharmaceutical preparations of hedgehog and ptc therapeutics

The source of the hedgehog and ptc therapeutics to be formulated will depend on the particular form of the agent. Small organic molecules and peptidyl fragments can be chemically synthesized and provided in a pure form suitable for pharmaceutical/cosmetic usage. Products of natural extracts can be purified according to techniques known in the art. For example, the Cox et al. U.S. Patent 5,286,654 describes a method for purifying naturally occurring forms of a secreted protein and can be adapted for purification of hedgehog polypeptides. Recombinant sources of hedgehog polypeptides are also available. For example, the gene encoding *hedgehog* polypeptides, are known, *inter alia*, from PCT publications WO 95/18856 and WO 96/17924.

Those of skill in treating peripheral neuropathies can determine the effective amount of an hedgehog or ptc therapeutic to be formulated in a pharmaceutical or cosmetic preparation.

The hedgehog or ptc therapeutic formulations used in the method of the invention are most preferably applied in the form of appropriate compositions. As appropriate compositions there may be cited all compositions usually employed for systemically or topically administering drugs. The pharmaceutically acceptable carrier should be substantially inert, so as not to act with the active component. Suitable inert carriers include water, alcohol polyethylene glycol, mineral oil or petroleum gel, propylene glycol and the like.

To prepare the pharmaceutical compositions of this invention, an effective amount of the particular hedgehog or ptc therapeutic as the active ingredient is combined in intimate admixture with a pharmaceutically acceptable carrier, which carrier may take a wide variety of forms depending on the form of preparation desired for administration. These pharmaceutical compositions are desirable in unitary dosage form suitable, particularly, for administration orally, rectally, percutaneously, or by parenteral injection. For example, in preparing the compositions in oral dosage form, any of the usual pharmaceutical media may be employed such as, for example, water, glycols, oils, alcohols and the like in the case of oral liquid preparations such as

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suspensions, syrups, elixirs and solutions; or solid carriers such as starches, sugars, kaolin, lubricants, binders, disintegrating agents and the like in the case of powders, pills, capsules, and tablets. Because of their ease in administration, tablets and capsules represents the most advantageous oral dosage unit form, in which case solid pharmaceutical carriers are obviously employed. For parenteral compositions, the carrier will usually comprise sterile water, at least in large part, though other ingredients, for example, to aid solubility, may be included. Injectable solutions, for example, may be prepared in which the carrier comprises saline solution, glucose solution or a mixture of saline and glucose solution. Injectable suspensions may also be prepared in which case appropriate liquid carriers, suspending agents and the like may be employed. Also included are solid form preparations which are intended to be converted, shortly before use, to liquid form preparations. In the compositons suitable for percutaneous administration, the carrier optionally comprises a penetration enhancing agent and/or a suitable wetting agent, optionally combined with suitable additives of any nature in minor proportions, which additives do not introduce a significant deleterious effect on the skin.

In addition to the direct topical application of the preparations they can be topically administered by other methods, for example, encapsulated in a temperature and/or pressure sensitive matrix or in film or solid carrier which is soluble in body fluids and the like for subsequent release, preferably sustained-release of the active component.

As appropriate compositions for topical application there may be cited all compositions usually employed for topically administering therapeuitcs, e.g., creams, gellies, dressings, shampoos, tinctures, pastes, ointments, salves, powders, liquid or semiliquid formulation and the like. Application of said compositions may be by aerosol e.g. with a propellent such as nitrogen carbon dioxide, a freon, or without a propellent such as a pump spray, drops, lotions, or a semisolid such as a thickened composition which can be applied by a swab. In particular compositions, semisolid compositions such as salves, creams, pastes, gellies, ointments and the like will conveniently be used.

It is especially advantageous to formulate the subject compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form as used in the specification and claims herein refers to physically discreate units suitable as unitary dosages, each unit containing a predetermined quantity of active ingredient calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. Examples of such dosage unit forms are tablets (including scored or coated tablets), capsules, pills, powders packets, wafers, injectable solutions or suspensions, teaspoonfuls, tablespoonfuls and the like, and segregated multiples thereof.

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The pharmaceutical preparations of the present invention can be used, as stated above, for the many applications which can be considered cosmetic uses. Cosmetic compositions known in the art, preferably hypoallergic and pH controlled are especially preferred, and include toilet waters, packs, lotions, skin milks or milky lotions. The preparations contain, besides the hedgehog or ptc therapeutic, components usually employed in such preparations. Examples of such components are oils, fats, waxes, surfactants, humectants, thickening agents, antioxidants, viscosity stabilizers, chelating agents, buffers, preservatives, perfumes, dyestuffs, lower alkanols, and the like. If desired, further ingredients may be incorporated in the compositions, e.g. antiinflammatory agents, antibacterials, antifungals, disinfectants, vitamins, sunscreens, antibiotics, or other anti-acne agents.

Examples of oils comprise fats and oils such as olive oil and hydrogenated oils; waxes such as beeswax and lanolin; hydrocarbons such as liquid paraffin, ceresin, and squalane; fatty acids such as stearic acid and oleic acid; alcohols such as cetyl alcohol, stearyl alcohol, lanolin alcohol, and hexadecanol; and esters such as isopropyl myristate, isopropyl palmitate and butyl stearate. As examples of surfactants there may be cited anionic surfactants such as sodium stearate, sodium cetylsulfate, polyoxyethylene laurylether phosphate, sodium N-acyl glutamate; such as stearyldimethylbenzylammonium chloride and cationic surfactants stearyltrimethylammonium chloride; ampholytic surfactants such as alkylaminoethylglycine hydrocloride solutions and lecithin; and nonionic surfactants such as glycerin monostearate, sorbitan monostearate, sucrose fatty acid esters, propylene glycol monostearate, polyoxyethylene oleylether, polyethylene glycol monostearate, polyoxyethylene sorbitan monopalmitate, polyoxyethylene coconut fatty acid monoethanolamide, polyoxypropylene glycol (e.g. the materials sold under the trademark "Pluronic"), polyoxyethylene castor oil, and polyoxyethylene lanolin. Examples of humectants include glycerin, 1,3-butylene glycol, and propylene glycol; examples of lower alcohols include ethanol and isopropanol; examples of thickening agents include xanthan gum, hydroxypropyl cellulose, hydroxypropyl methyl cellulose, polyethylene glycol and sodium carboxymethyl cellulose; examples of antioxidants comprise butylated hydroxytoluene, butylated hydroxyanisole, propyl gallate, citric acid and ethoxyquin; examples of chelating agents include disodium edetate and ethanehydroxy diphosphate; examples of buffers comprise citric acid, sodium citrate, boric acid, borax, and disodium hydrogen phosphate; and examples of preservatives are methyl parahydroxybenzoate, ethyl parahydroxybenzoate, dehydroacetic acid, salicylic acid and benzoic acid.

For preparing ointments, creams, toilet waters, skin milks, and the like, typically from 0.01 to 10% in particular from 0.1 to 5% and more in particular from 0.2 to 2.5% of the active ingredient, e.g., of the hedgehog or ptc therapeutic, will be incorporated in the compositions. In

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ointments or creams, the carrier for example consists of 1 to 20%, in particular 5 to 15% of a humectant, 0.1 to 10% in particular from 0.5 to 5% of a thickener and water; or said carrier may consist of 70 to 99%, in particular 20 to 95% of a surfactant, and 0 to 20%, in particular 2.5 to 15% of a fat; or 80 to 99.9% in particular 90 to 99% of a thickener; or 5 to 15% of a surfactant, 2-15% of a humectant, 0 to 80% of an oil, very small (<2%) amounts of preservative, coloring agent and/or perfume, and water. In a toilet water, the carrier for example consists of 2 to 10% of a lower alcohol, 0.1 to 10% or in particular 0.5 to 1% of a surfactant, 1 to 20%, in particular 3 to 7% of a humectant, 0 to 5% of a buffer, water and small amounts (<2%) of preservative, dyestuff and/or perfume. In a skin milk, the carrier typically consists of 10-50% of oil, 1 to 10% of surfactant, 50-80% of water and 0 to 3% of preservative and/or perfume. In the aforementioned preparations, all % symbols refer to weight by weight percentage.

Particular compositions for use in the method of the present invention are those wherein the hedgehog or ptc therapeutic is formulated in liposome-containing compositions. Liposomes are artificial vesicles formed by amphiphatic molecules such as polar lipids, for example, phosphatidyl cholines, ethanolamines and serines, sphingomyelins, cardiolipins, plasmalogens, phosphatidic acids and cerebiosides. Liposomes are formed when suitable amphiphathic molecules are allowed to swell in water or aqueous solutions to form liquid crystals usually of multilayer structure comprised of many bilayers separated from each other by aqueous material (also referred to as coarse liposomes). Another type of liposome known to be consisting of a single bilayer encapsulating aqueous material is referred to as a unilamellar vesicle. If watersoluble materials are included in the aqueous phase during the swelling of the lipids they become entrapped in the aqueous layer between the lipid bilayers.

Water-soluble active ingredients such as, for example, various salt forms of a hedgehog polypeptide, are encapsulated in the aqueous spaces between the molecular layers. The lipid soluble active ingredient of hedgehog or ptc therapeutic, such as an organic mimetic, is predominantly incorporated into the lipid layers, although polar head groups may protude from the layer into the aqueous space. The encapsulation of these compounds can be achieved by a number of methods. The method most commonly used involves casting a thin film of phospholipid onto the walls of a flask by evaporation from an organic solvent. When this film is dispersed in a suitable aqueous medium, multilamellar liposomes are formed. Upon suitable sonication, the coarse liposomes form smaller similarly closed vesicles.

Water-soluble active ingredients are usually incorporated by dispersing the cast film with an aqueous solution of the compound. The unencapsulated compound is then removed by centrifugation, chromatography, dialysis or other art-known suitable procedures. The lipid-

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soluble active ingredient is usually incorporated by dissolving it in the organic solvent with the phospholipid prior to casting the film. If the solubility of the material in the lipid phase is not exceeded or the amount present is not in excess of that which can be bound to the lipid, liposomes prepared by the above method usually contain most of the material bound in the lipid bilayers; separation of the liposomes from unencapsulated material is not required.

A particularly convenient method for preparing liposome formulated forms of hedgehog and ptc therapeutics is the method described in EP-A-253,619, incorporated herein by reference. In this method, single bilayered liposomes containing encapsulated active ingredients are prepared by dissolving the lipid component in an organic medium, injecting the organic solution of the lipid component under pressure into an aqueous component while simultaneously mixing the organic and aqueous components with a high speed homogenizer or mixing means, whereupon the liposomes are formed spontaneously.

The single bilayered liposomes containing the encapsulated hedgehog or ptc therapeutic can be employed directly or they can be employed in a suitable pharmaceutically acceptable carrier for topical administration. The viscosity of the liposomes can be increased by the addition of one or more suitable thickening agents such as, for example xanthan gum, hydroxypropyl cellulose, hydroxypropyl methylcellulose and mixtures thereof. The aqueous component may consist of water alone or it may contain electrolytes, buffered systems and other ingredients, such as, for example, preservatives. Suitable electrolytes which can be employed include metal salts such as alkali metal and alkaline earth metal salts. The preferred metal salts are calcium chloride, sodium chloride and potassium chloride. The concentration of the electrolyte may vary from zero to 260 mM, preferably from 5 mM to 160 mM. The aqueous component is placed in a suitable vessel which can be adapted to effect homogenization by effecting great turbulence during the injection of the organic component. Homogenization of the two components can be accomplished within the vessel, or, alternatively, the aqueous and organic components may be injected separately into a mixing means which is located outside the vessel. In the latter case, the liposomes are formed in the mixing means and then transferred to another vessel for collection purpose.

The organic component consists of a suitable non-toxic, pharmaceutically acceptable solvent such as, for example ethanol, glycerol, propylene glycol and polyethylene glycol, and a suitable phospholipid which is soluble in the solvent. Suitable phospholipids which can be employed include lecithin, phosphatidylcholine, phosphatydylserine, phosphatidylethanol-amine, phosphatidylinositol, lysophosphatidylcholine and phospha-tidyl glycerol, for example. Other lipophilic additives may be employed in order to selectively modify the characteristics of the

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liposomes. Examples of such other additives include stearylamine, phosphatidic acid, tocopherol, cholesterol and lanolin extracts.

In addition, other ingredients which can prevent oxidation of the phospholipids may be added to the organic component. Examples of such other ingredients include tocopherol, butylated hydroxyanisole, butylated hydroxytoluene, ascorbyl palmitate and ascorbyl oleate. Preservatives such a benzoic acid, methyl paraben and propyl paraben may also be added.

Apart from the above-described compositions, use may be made of covers, e.g. plasters, bandages, dressings, gauze pads and the like, containing an appropriate amount of a hedgehog or ptc therapeutic. In some cases use may be made of plasters, bandages, dressings, gauze pads and the like which have been impregnated with a topical formulation containing the therapeutic formulation.

### VII. Exemplary Hedgehog Antagonists

Exemplary *hedgehog* antagonists of the invention include cyclopamine, tomatidine, jervine, AY9944, triparanol, forskolin, cAMP, dibutyryl cAMP (and other hydrophobically modified cAMP variants) and fuctionally effective derivatives thereof. Additional *hedgehog* antagonists, functionally effective derivatives, compositions and methods for making such compounds are described in detail in the following U.S. Patent applications: Baxter et al., 09/663,835; Beachy et al. entitled "Inhibitors of hedgehog signaling pathways, compositions and uses related thereto" filed October 10, 2000; Baxter et al. "Mediators of hedgehog signaling pathway, compositions and uses related thereto" filed October 13, 2000; Philip Beachy, "Regulators of the hedgehog pathway, compositions and uses related thereto" filed October 13, 2000.

#### Evaluation of the effects of hedgehog on T-cell development

The invention now being generally described, it will be more readily understood by reference to the following examples which are included merely for purposes of illustration of certain aspects and embodiments of the present invention, and are not intended to limit the invention.

Utilizing a fetal thymic organ culture, the effects of *Shh* on T-cell maturation was determined. Briefly, immature thymocytes (CD4-/CD8- [DN]) were isolated from E14 thymus.

The cells were culture for about 7 days, and the level of mature thymocytes, e.g., CD4+/CD8+ [DP], was assessed. In certain cultures, octyl-modified Shh was added to the culture; in other cultures, the anti-hedgehog 5E1 antibody was added. As illustrated in Figures 1-4, the addition of Shh to the culture resulted in an increase in the percentage of cells after 7 days with markers indicative of immature thymocytes. Conversely, addition of the 5E1 antibody resulted in an increase in the percentage of mature T lymphocytes after 7 days.

All of the above-cited references and publications are hereby incorporated by reference.

#### 10 Equivalents

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Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, numerous equivalents to the specific polypeptides, nucleic acids, methods, assays and reagents described herein. Such equivalents are considered to be within the scope of this invention.

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55	Phe			Asp					Asp					Thr		GCG Ala	864
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	Ser 305	Gly	Gln	Leu	Lys	Ser 310	Val	Ile	Val	Gln	Arg 315	Ile	Tyr	Thr	Glu	Glu 320	
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	0.0		GTG Val											152
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1 3 3 x 3	30		AGC Ser											248
a jul	, 0		GGC Gly											296
14 3 14 14 14 14 14 14 14 14 14 14 14 14 14 1	35		TAC Tyr 85											344
	40		CGC Arg										CTG Leu	392
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(	60												CAC His	632

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30	ATG GCT CTC CTG ACC AAT CTA CTG CCC TTG TGC TGC TTG GCA CTT CTG Met Ala Leu Leu Thr Asn Leu Leu Pro Leu Cys Cys Leu Ala Leu Leu  1 10 15	48
35	GCG CTG CCA GCC CAG AGC TGC GGG CCG GGC CGG GGG CCG GTT GGC CGG Ala Leu Pro Ala Gln Ser Cys Gly Pro Gly Arg Gly Pro Val Gly Arg 20 25 30	96
40	CGC CGC TAT GCG CGC AAG CAG CTC GTG CCG CTA CTC TAC AAG CAA TTT Arg Arg Tyr Ala Arg Lys Gln Leu Val Pro Leu Leu Tyr Lys Gln Phe 35 40	144
40	GTG CCC GGC GTG CCA GAG CGG ACC CTG GGC GCC AGT GGG CCA GCG GAG Val Pro Gly Val Pro Glu Arg Thr Leu Gly Ala Ser Gly Pro Ala Glu 50 55 60	192
45	GGG AGG GTG GCA AGG GGC TCC GAG CGC TTC CGG GAC CTC GTG CCC AAC Gly Arg Val Ala Arg Gly Ser Glu Arg Phe Arg Asp Leu Val Pro Asn 65 70 75 80	240
50	TAC AAC CCC GAC ATC ATC TTC AAG GAT GAG GAG AAC AGT GGA GCC GAC Tyr Asn Pro Asp Ile Ile Phe Lys Asp Glu Glu Asn Ser Gly Ala Asp 85 90 95	288
55	CGC CTG ATG ACC GAG CGT TGC AAG GAG AGG GTG AAC GCT TTG GCC ATT Arg Leu Met Thr Glu Arg Cys Lys Glu Arg Val Asn Ala Leu Ala Ile 100 105 110	336
60	GCC GTG ATG AAC ATG TGG CCC GGA GTG CGC CTA CGA GTG ACT GAG GGC Ala Val Met Asn Met Trp Pro Gly Val Arg Leu Arg Val Thr Glu Gly 115 120 125	384

			GGC Gly											432
5			ATC Ile											480
10			CTC Leu 165											528
15			CAC His											576
20			GGC Gly											624
			CGG Arg											672
₹25 11 2			GAT Asp											720
□ □ 30 =			CGG Arg 245											768
<b>35</b>			CCT Pro										CTG Leu	816
<b>1</b> 40			CGA Arg			Pro							CCG Pro	864
										Leu			GGC Gly	912
45	Asp				Ala				Val				GAA Glu 320	960
50				Ala				His					GTG Val	1008
55			Ala				Val					Gln	TGG	1056
60		Ala				. Arg					Leu		GCG Ala	1104

	CTG Leu	CTC Leu 370	CCC Pro	GGC Gly	GGG Gly	GCC Ala	GTC Val 375	CAG Gln	CCG Pro	ACT Thr	GGC Gly	ATG Met 380	CAT His	TGG Trp	TAC Tyr	TCT Ser	1152
5			CTC Leu										TG				1191
10	(2)	INFO	OR <b>MA</b> T	TION	FOR	SEQ	ID 1	10:9	:								
15		(i)	(I	A) L1 3) T' C) S'		H: 12 nucl	251 l Leic ESS:	oase acio bot	pai: d	rs							
20		(ii	) MOI	LECU!	LE T	YPE:	cDN	Ą									
다 다 날 25		(ix		A) N.	E: AME/1 OCAT:			1248									
		(xi	) SE	QUEN	CE D	ESCR	IPTI	ON:	SEQ	ID N	0:9:						
25	ATG Met 1	GAC Asp	GTA Val	AGG Arg	CTG Leu 5	CAT His	CTG Leu	AAG Lys	CAA Gln	TTT Phe 10	GCT Ala	TTA Leu	CTG Leu	TGT Cys	TTT Phe 15	ATC Ile	48
35	AGC Ser	TTG Leu	CTT Leu	CTG Leu 20	Thr	CCT Pro	TGT Cys	GGA Gly	TTA Leu 25	GCC Ala	TGT Cys	GGT Gly	CCT Pro	GGT Gly 30	AGA Arg	GGT Gly	96
1_1	TAT Tyr	GGA Gly	AAA Lys 35	Arg	AGA Arg	CAC His	CCA Pro	AAG Lys 40	Lys	TTA Leu	ACC Thr	CCG Pro	TTG Leu 45	GCT Ala	TAC Tyr	AAG Lys	144
40	CAA Gln	TTC Phe	ATC	CCC	AAC Asn	GTT Val	GCT Ala 55	Glu	AAA Lys	ACG Thr	Leu	GGA Gly	Ala	AGC Ser	GGC Gly	AAA Lys	192
45	TAC Tyr 65	Glu	GGC Gly	AAA Lys	ATC	ACA Thr	Arg	AAT Asr	TCA Ser	GAG Glu	AGA Arg	Phe	AAA Lys	GAG Glu	CTG Leu	ATT Ile 80	240
50			TAT 1 Tyr			Asp					Asp					Asn	288
55					ı Met					Lys					Ser	TTG Leu	336
60	GCC Ala	C ATA	A TCC e Ser 115	· Val	C ATO	AAC Asr	CAC His	TGC Try 120	p Pro	GGC Gly	GTC Val	B AAA Lys	CTG Leu 125	Arg	GTC Val	ACT Thr	384

	GAA Glu	GGC Gly 130	TGG Trp	GAT Asp	GAG Glu	GAT Asp	GGT Gly 135	CAC His	CAT His	TTA Leu	GAA Glu	GAA Glu 140	TCT Ser	TTG Leu	CAC His	TAT Tyr	432
5									ACC Thr								480
10	TAT Tyr	GGG Gly	ATG Met	CTA Leu	TCC Ser 165	AGG Arg	CTT Leu	GCA Ala	GTG Val	GAG Glu 170	GCA Ala	GGA Gly	TTC Phe	GAC Asp	TGG Trp 175	GTC Val	528
15	TAT Tyr	TAT Tyr	GAA Glu	TCT Ser 180	AAA Lys	GCC Ala	CAC His	ATA Ile	CAC His 185	TGC Cys	TCT Ser	GTC Val	AAA Lys	GCA Ala 190	GAA Glu	AAT Asn	576
20	TCA Ser	GTG Val	GCT Ala 195	GCT Ala	AAA Lys	TCA Ser	GGA Gly	GGA Gly 200	TGT Cys	TTT Phe	CCT Pro	GGG Gly	TCT Ser 205	GGG Gly	ACG Thr	GTG Val	624
20									CCC Pro								672
25 = 30	GAC Asp 225	CGG Arg	GTT Val	TTG Leu	GCT Ala	GCA Ala 230	GAC Asp	GAG Glu	AAG Lys	GGA Gly	AAT Asn 235	GTC Val	TTA Leu	ATA Ile	AGC Ser	GAC Asp 240	720
] ] [30									CCG Pro								768
======================================									ACC Thr 265								816
35 1 1 1 40				Phe					Ser					Ile		GCA Ala	864
40	Thr		Ala	Ser	Asn	Val	Lys	Pro		Asp	Thr	Val	Leu			GAA Glu	912
45		Thr					Lys					Lys				ACT Thr 320	960
50	GAG Glu	GAG Glu	CAC His	GAG Glu	GGC Gly 325	Ser	TTT Phe	GCG Ala	CCA Pro	GTC Val 330	Thr	GCG Ala	CAC His	GGA Gly	ACC Thr 335	ATA Ile	1008
55					Val					Tyr					Asr	CAC	1056
60	AAA Lys	TGG Trp	GCA Ala 355	His	TGG Trp	GCT Ala	TTT Phe	GCG Ala 360	Pro	GTC Val	AGG Arg	TTG Leu	TGT Cys 365	His	AAC Lys	CTG Leu	1104

- 78 -

	ATG Met	ACG Thr 370	TGG Trp	CTT Leu	TTT Phe	CCG Pro	GCT Ala 375	CGT Arg	GAA Glu	TCA Ser	AAC Asn	GTC Val 380	AAT Asn	TTT Phe	CAG Gln	GAG Glu	1152
5	GAT Asp 385	GGT Gly	ATC Ile	CAC His	TGG Trp	TAC Tyr 390	TCA Ser	AAT Asn	ATG Met	CTG Leu	TTT Phe 395	CAC His	ATC Ile	GGC Gly	TCT Ser	TGG Trp 400	1200
10	CTG Leu	CTG Leu	GAC Asp	AGA Arg	GAC Asp 405	TCT Ser	TTC Phe	CAT His	CCA Pro	CTC Leu 410	GGG Gly	ATT Ile	TTA Leu	CAC His	TTA Leu 415	AGT Ser	1248
	TGA																1251
15	(2)	INFO	ORMA'	rion	FOR	SEQ	ID 1	NO:1	0:								
20			(i) s	(A)	LEI TY		: 42 amin	5 am o ac			S						
		(:	ii) I	MOLE	CULE	TYP	E: p	rote	in								
<b>125</b>		(:	xi)	SEQU	ENCE	DES	CRIP	TION	: SE	Q ID	NO:	10:					
25	Met 1	Val	Glu	Met	Leu 5	Leu	Leu	Thr	Arg	Ile 10	Leu	Leu	Val	Gly	Phe 15	Ile	
22	Cys	Ala	Leu	Leu 20	Val	Ser	Ser	Gly	Leu 25	Thr	Cys	Gly	Pro	Gly 30	Arg	Gly	
블 글 35	Ile	Gly	Lys 35		Arg	His	Pro	Lys 40	Lys	Leu	Thr	Pro	Leu 45	Ala	Tyr	Lys	
35 M M	Gln	Phe 50	Ile	Pro	Asn	Val	Ala 55		Lys	Thr	Leu	Gly 60		Ser	Gly	Arg	
□ 40	Tyr 65	Glu	Gly	Lys	Ile	Thr 70		Asn	. Ser	Glu	Arg 75		Lys	Glu	Leu	Thr 80	
4.5	Pro	Asn	Tyr	Asn	Pro 85		Ile	: Ile	e Phe	Lys 90		Glu	Glu	Asn	Thr 95	Gly	
45	Ala	Asp	Arg	Leu 100		Thr	Gln	Arg	Cys 105		Asp	Lys	Leu	Asn 110		Leu	
50	Ala	Ile	Ser		Met	Asn	Glr	120		Gly	v Val	. Lys	Leu 125		[ Val	Thr	
	Glu	Gly 130	_	Asp	Glu	. Asp	Gl <sub>y</sub>		s His	s Ser	Glu	Glu 140		Leu	His	Tyr	
55	Glu 145	_	Arg	g Ala	ı Val	. Asp		e Thi	c Thr	Ser	Asp 155		, Asp	Arg	g Sei	Lys 160	
60	Tyr	Gly	Met	: Leu	ı Ala 165		j Lei	ı Ala	a Val	1 Glu 170		a Gly	7 Ph∈	e Asp	7rr 175	Val	

# ONV-080.01

		Tyr	Tyr	Glu	Ser 180	Lys	Ala	His	Ile	His 185	Cys	Ser	Val	Lys	Ala 190	Glu	Asn
	5	Ser	Val	Ala 195	Ala	Lys	Ser	Gly	Gly 200	Cys	Phe	Pro	Gly	Ser 205	Ala	Thr	Val
		His	Leu 210	Glu	His	Gly	Gly	Thr 215	Lys	Leu	Val	Lys	Asp 220	Leu	Ser	Pro	Gly
1	0	Asp 225	Arg	Val	Leu	Ala	Ala 230	Asp	Ala	Asp	Gly	Arg 235	Leu	Leu	Tyr	Ser	Asp 240
1	.5	Phe	Leu	Thr	Phe	Leu 245	Asp	Arg	Met	Asp	Ser 250	Ser	Arg	Lys	Leu	Phe 255	Tyr
1		Val	Ile	Glu	Thr 260	Arg	Gln	Pro	Arg	Ala 265	Arg	Leu	Leu	Leu	Thr 270	Ala	Ala
2	20	His	Leu	Leu 275	Phe	Val	Ala	Pro	Gln 280	His	Asn	Gln	Ser	Glu 285	Ala	Thr	Gly
		Ser	Thr 290	Ser	Gly	Gln	Ala	Leu 295	Phe	Ala	Ser	Asn	Val 300	Lys	Pro	Gly	Gln
*42	25	Arg 305	Val	Tyr	Val	Leu	Gly 310	Glu	Gly	Gly	Gln	Gln 315	Leu	Leu	Pro	Ala	Ser 320
	80	Val	His	Ser	Val	Ser 325	Leu	Arg	Glu	Glu	Ala 330	Ser	Gly	Ala	Tyr	Ala 335	Pro
annelly annelly annelly annelly annelly	,0	Leu	Thr	Ala	Gln 340	Gly	Thr	Ile	Leu	Ile 345	Asn	Arg	Val	Leu	Ala 350	Ser	Cys
	35	Tyr	Ala	Val 355	Ile	Glu	Glu	His	Ser 360	Trp	Ala	His	Trp	Ala 365	Phe	Ala	Pro
		Phe	Arg 370		Ala	Gln	Gly	Leu 375	Leu	Ala	Ala	Leu	Cys 380		Asp	Gly	Ala
	40	Ile 385		Thr	Ala	Ala	Thr 390	Thr	Thr	Thr	Gly	Ile 395	His	Trp	Tyr	Ser	Arg 400
4	45	Leu	Leu	Tyr	Arg	Ile 405	Gly	Ser	Trp	Val	Leu 410		Gly	Asp	Ala	Leu 415	
		Pro	Leu	Gly	Met 420		Ala	Pro	Ala	Ser 425							
;	50	(2)	INF	'ORMA	TION	FOR	SEQ	ID	NO:1	1:							
				(i)	(A	) LE	NGTH	RACT : 39 amin	6 am	ino		.s					
	55		(	(ii)	(D MOLE			GY: E: p									

60

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Gly Asp Ala Leu Gln Pro Ala Arg Val Ala Arg Val Ala Arg Glu Glu

		305					310					315					320
	5	Ala	Val	Gly	Val	Phe 325	Ala	Pro	Leu	Thr	Ala 330	His	Gly	Thr	Leu	Leu 335	Val
	3	Asn	Asp	Val	Leu 340	Ala	Ser	Cys	Tyr	Ala 345	Val	Leu	Glu	Ser	His 350	Gln	Trp
	10	Ala	His	Arg 355	Ala	Phe	Ala	Pro	Leu 360	Arg	Leu	Leu	His	Ala 365	Leu	Gly	Ala
		Leu	Leu 370	Pro	Gly	Gly	Ala	Val 375	Gln	Pro	Thr	Gly	Met 380	His	Trp	Tyr	Ser
	15	Arg 385	Leu	Leu	Tyr	Arg	Leu 390	Ala	Glu	Glu	Leu	Met 395	Gly				
	20	(2)	INFO	ORMAT	rion	FOR	SEQ	ID N	NO:12	2:							
	2.5			(i) S	SEQUE (A) (B) (D)	LEN TYI		: 411 amino	l am	ino a id	: acids	5					
The state of the s	25		(:	ii) ľ	MOLE	CULE	TYPI	E: pi	rote	in							
	30		(2	xi) s	SEQUI	ENCE	DESC	CRIP	rion	: SE	Q ID	NO:	12:				
ii ļui		Met 1	Ser	Pro	Ala	Trp 5	Leu	Arg	Pro	Arg	Leu 10	Arg	Phe	Cys	Leu	Phe 15	Leu
	35	Leu	Leu	Leu	Leu 20	Leu	Val	Pro	Ala	Ala 25	Arg	Gly	Cys	Gly	Pro 30	Gly	Arg
	40	Val	Val	Gly 35	Ser	Arg	Arg	Arg	Pro 40	Pro	Arg	Lys	Leu	Val 45	Pro	Leu	Ala
		Tyr	Lys 50	Gln	Phe	Ser	Pro	Asn 55	Val	Pro	Glu	Lys	Thr 60	Leu	Gly	Ala	Ser
	45	Gly 65	Arg	Tyr	Glu	Gly	Lys 70	Ile	Ala	Arg	Ser	Ser 75		Arg	Phe	Lys	Glu 80
		Leu	Thr	Pro	Asn	Tyr 85	Asn	Pro	Asp	Ile	Ile 90	Phe	Lys	Asp	Glu	Glu 95	Asn
	50	Thr	Gly	Ala	Asp 100	Arg	Leu	Met	Thr	Gln 105		Cys	Lys	Asp	Arg 110	Leu	Asn
	55	Ser	Leu	Ala 115	Ile	Ser	Val	Met	Asn 120		Trp	Pro	Gly	Val 125	Lys	Leu	Arg
	55	Val	Thr 130		Gly	Arg	Asp	Glu 135	Asp	Gly	His	His	Ser 140		Glu	Ser	Leu
	60	His	Tyr	Glu	Gly	Arg	Ala		Asp	Ile	Thr	Thr		Asp	Arg	Asp	Arg

	Asn	Lys	Tyr	Gly	Leu 165	Leu	Ala	Arg	Leu	Ala 170	Val	Glu	Ala	Gly	Phe 175	Asp
5	Trp	Val	Tyr	Tyr 180	Glu	Ser	Lys	Ala	His 185	Val	His	Cys	Ser	Val 190	Lys	Ser
10	Glu	His	Ser 195	Ala	Ala	Ala	Lys	Thr 200	Gly	Gly	Cys	Phe	Pro 205	Ala	Gly	Ala
10	Gln	Val 210	Arg	Leu	Glu	Asn	Gly 215	Glu	Arg	Val	Ala	Leu 220	Ser	Ala	Val	Lys
15	Pro 225	Gly	Asp	Arg	Val	Leu 230	Ala	Met	Gly	Glu	Asp 235	Gly	Thr	Pro	Thr	Phe 240
	Ser	Asp	Val	Leu	Ile 245	Phe	Leu	Asp	Arg	Glu 250	Pro	Asn	Arg	Leu	Arg 255	Ala
20	Phe	Gln	Val	Ile 260	Glu	Thr	Gln	Asp	Pro 265	Pro	Arg	Arg	Leu	Ala 270	Leu	Thr
] ]   25	Pro	Ala	His 275	Leu	Leu	Phe	Ile	Ala 280	Asp	Asn	His	Thr	Glu 285	Pro	Ala	Ala
<u>25</u>	His	Phe 290	Arg	Ala	Thr	Phe	Ala 295	Ser	His	Val	Gln	Pro 300	Gly	Gln	Tyr	Val
30	Leu 305	Val	Ser	Gly	Val	Pro 310	Gly	Leu	Gln	Pro	Ala 315	Arg	Val	Ala	Ala	Val 320
i i	Ser	Thr	His	Val	Ala 325	Leu	Gly	Ser	Tyr	Ala 330	Pro	Leu	Thr	Arg	His 335	Gly
35 5	Thr	Leu	Val	Val 340	Glu	Asp	Val	Val	Ala 345	Ser	Cys	Phe	Ala	Ala 350		Ala
40	Asp	His	His 355	Leu	Ala	Gln	Leu	Ala 360	Phe	Trp	Pro	Leu	Arg 365		Phe	Pro
40	Ser	Leu 370		Trp	Gly	Ser	Trp 375		Pro	Ser	Glu	Gly 380		His	Ser	Tyr
45	Pro 385	Gln	Met	Leu	Tyr	Arg 390		Gly	Arg	Leu	Leu 395		Glu	Glu	Ser	Thr 400
	Phe	His	Pro	Leu	Gly 405		Ser	Gly	Ala	Gly 410						
50																
	(2)	INF	ORMA	TION	FOR	SEQ	D	NO:1	.3:							
55			(i)	(A	L) LE	NGTH	: 43 amin	ERIS 7 am 10 ac 1ine	ino id		ls					
60		(	ii)	MOLE	CULE	TYF	E: p	rote	in							

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13: 5 Met Leu Leu Leu Ala Arg Cys Phe Leu Val Ile Leu Ala Ser Ser Leu Leu Val Cys Pro Gly Leu Ala Cys Gly Pro Gly Arg Gly Phe Gly 10 Lys Arg Arg His Pro Lys Lys Leu Thr Pro Leu Ala Tyr Lys Gln Phe Ile Pro Asn Val Ala Glu Lys Thr Leu Gly Ala Ser Gly Arg Tyr Glu 15 Gly Lys Ile Thr Arg Asn Ser Glu Arg Phe Lys Glu Leu Thr Pro Asn 20 Tyr Asn Pro Asp Ile Ile Phe Lys Asp Glu Glu Asn Thr Gly Ala Asp Arg Leu Met Thr Gln Arg Cys Lys Asp Lys Leu Asn Ala Leu Ala Ile T. 25 Ser Val Met Asn Gln Trp Pro Gly Val Arg Leu Arg Val Thr Glu Gly T. 115 1 Trp Asp Glu Asp Gly His His Ser Glu Glu Ser Leu His Tyr Glu Gly **30** 135 Arg Ala Val Asp Ile Thr Thr Ser Asp Arg Asp Arg Ser Lys Tyr Gly Ħ 145 Met Leu Ala Arg Leu Ala Val Glu Ala Gly Phe Asp Trp Val Tyr Tyr Glu Ser Lys Ala His Ile His Cys Ser Val Lys Ala Glu Asn Ser Val **=** 40 Ala Ala Lys Ser Gly Gly Cys Phe Pro Gly Ser Ala Thr Val His Leu Glu Gln Gly Gly Thr Lys Leu Val Lys Asp Leu Arg Pro Gly Asp Arg 45 Val Leu Ala Ala Asp Asp Gln Gly Arg Leu Leu Tyr Ser Asp Phe Leu 50 Thr Phe Leu Asp Arg Asp Glu Gly Ala Lys Lys Val Phe Tyr Val Ile Glu Thr Leu Glu Pro Arg Glu Arg Leu Leu Leu Thr Ala Ala His Leu 55 Leu Phe Val Ala Pro His Asn Asp Ser Gly Pro Thr Pro Gly Pro Ser Ala Leu Phe Ala Ser Arg Val Arg Pro Gly Gln Arg Val Tyr Val Val 60 295

		Ala 305	Glu	Arg	Gly	Gly	Asp 310	Arg	Arg	Leu	Leu	Pro 315	Ala	Ala	Val	His	Ser 320
;	5	Val	Thr	Leu	Arg	Glu 325	Glu	Glu	Ala	Gly	Ala 330	Tyr	Ala	Pro	Leu	Thr 335	Ala
1	0	His	Gly	Thr	Ile 340	Leu	Ile	Asn	Arg	Val 345	Leu	Ala	Ser	Cys	Tyr 350	Ala	Val
1	U	Ile	Glu	Glu 355	His	Ser	Trp	Ala	His 360	Arg	Ala	Phe	Ala	Pro 365	Phe	Arg	Leu
1	5	Ala	His 370	Ala	Leu	Leu	Ala	Ala 375	Leu	Ala	Pro	Ala	Arg 380	Thr	Asp	Gly	Gly
		Gly 385	Gly	Gly	Ser	Ile	Pro 390	Ala	Ala	Gln	Ser	Ala 395	Thr	Glu	Ala	Arg	Gly 400
	0	Ala	Glu	Pro	Thr	Ala 405	Gly	Ile	His	Trp	Tyr 410	Ser	Gln	Leu	Leu	Tyr 415	His
□ □ ≒2	5	Ile	Gly	Thr	Trp 420	Leu	Leu	Asp	Ser	Glu 425	Thr	Met	His	Pro	Leu 430	Gly	Met
3	J	Ala	Val	Lys 435	Ser	Ser											
	^	(2)															
<b>II</b> 3	U	(2)	TNF.	ORMA'	rion	FOR	SEQ	ID 1	NO:1	4:							
# 3 # 3		(2)			SEQUI	ENCE ) LEI ) TYI	CHA NGTH PE:	RACT: : 41 amin	ERIS' 8 am o ac line	TICS ino a		s					
		(2)	(:	(i) :	SEQUI (A) (B) (D)	ENCE ) LEI ) TY: ) TO:	CHANGTH PE: 6 POLOG	RACT: : 41 amin GY: E: p	ERIS' 8 am o ac	TICS ino id ar in	acid		14:				
4			(:	(i) : ii) ! xi) :	SEQUI (A (B (D (D	ENCE ) LEI ) TY: ) TO: CULE	CHAINGTH PE: POLO	RACT: 41 amin GY: E: p	ERIS' 8 am o ac line rote	TICS ino id ar in	acid	NO:	14: Leu	Leu	Thr	Leu 15	Ser
3 11 12 14		Met 1	(; (; Arg	(i) : ii) ! xi) : Leu	SEQUI (A) (B) (D) MOLE SEQUI	ENCE ) LEI ) TY: ) TO: CULE ENCE Thr	CHANGTH PE: OPOLOG TYP: DES	RACT: : 41 amin GY: E: p CRIP	ERIS'8 am o ac line rote TION Leu	TICS ino id ar in . : SE	Q ID Val 10	NO: Ser	Leu			15	Ser
	35 10	Met 1 Leu	(: Arg Val	(i) : ii) ! xi) : Leu Val	SEQUI (A) (B) (D) MOLE SEQUI Leu Ser 20	ENCE ) LEI ) TY: CULE  ENCE  Thr 5	CHANGTH PE: POLO TYP  DES Arg	RACT: : 41 amin GY: E: p CRIP Val	ERIS'8 am o ac line rote TION Leu Cys	rics ino id ar in : SE Leu Gly 25	Q ID Val 10 Pro	NO: Ser	Leu Arg	Gly	Tyr 30	15 Gly	
	55 10 15	Met 1 Leu Arg	(; Arg Val Arg	(i) s ii) l xi) s Leu Val His 35	SEQUI (A) (B) (D) MOLE SEQUI Leu Ser 20	ENCE ) LET ) TO: CULE  ENCE  Thr 5 Gly Lys	CHANGTH PE: POLO TYP  DES Arg  Leu  Lys	RACT: : 41 amin GY: E: p  CRIP  Val  Ala  Leu	ERIS'8 am o ac line rote TION Leu Cys Thr 40 Leu	rics ino id ar in : SE Leu Gly 25 Pro	Q ID Val 10 Pro	NO: Ser Gly	Leu Arg Tyr	Gly Lys 45	Tyr 30 Gln	15 Gly Phe	Arg
	55 10 15	Met 1 Leu Arg	(: Arg Val Arg Asn 50	(i) s ii) s xi) s Leu Val His 35 Val	SEQUI (A) (B) (D) MOLE SEQUI Leu Ser 20 Pro	ENCE ) LET ) TO ) TO  CULE  ENCE  Thr 5  Gly  Lys	CHANGTH PE: POLO TYP  DES Arg Leu Lys	RACT: 41 amin GY: E: p  CRIP Val Ala Leu Thr 55	ERIS'8 am o ac line rote TION Leu Cys Thr 40 Leu	rics ino id ar in : SE Leu Gly 25 Pro	Q ID Val 10 Pro Leu	NO: Ser Gly Ala	Leu Arg Tyr Gly 60 Leu	Gly Lys 45 Arg	Tyr 30 Gln Tyr	Gly Phe	Arg

Ser Ser

	5	(2)	INFC	RMAT	CION	FOR	SEQ	ID N	10:15	:							
	10		(	(i) S	(A) (B)	INCE LEN TYP TOP	IGTH: E: a	475 minc	ami aci	.no a .d							
			(i	i) M	OLEC	CULE	TYPE	: pr	otei	.n							
	15		(2	ki) S	EQUE	ENCE	DESC	RIPT	: NOI	SEÇ	) ID	NO:1	.5:				
	20	Met 1	Leu	Leu	Leu	Ala 5	Arg	Cys	Leu	Leu	Leu 10	Val	Leu	Val	Ser	Ser 15	Leu
		Leu	Val	Cys	Ser 20	Gly	Leu	Ala	Cys	Gly 25	Pro	Gly	Arg	Gly	Phe 30	Gly	Lys
Man Man	25	Arg	Arg	His 35	Pro	Lys	Lys	Leu	Thr 40	Pro	Leu	Ala	Tyr	Lys 45	Gln	Phe	Ile
		Pro	Asn 50	Val	Ala	Glu	Lys	Thr 55	Leu	Gly	Ala	Ser	Gly 60	Arg	Tyr	Glu	Gly
	30	Lув 65	Ile	Ser	Arg	Asn	Ser 70	Glu	Arg	Phe	Lys	Glu 75	Leu	Thr	Pro	Asn	Tyr 80
	35	Asn	Pro	Asp	Ile	Ile 85	Phe	Lys	Asp	Glu	Glu 90	Asn	Thr	Gly	Ala	Asp 95	Arg
	33	Leu	Met	Thr	Gln 100	Arg	Cys	Lys	Asp	Lys 105	Leu	Asn	Ala	Leu	Ala 110	Ile	Ser
	40	Val	Met	Asn 115	Gln	Trp	Pro	Gly	Val 120	Lys	Leu	Arg	Val	Thr 125	Glu	Gly	Trp
		Asp	Glu 130	Asp	Gly	His	His	Ser 135	Glu	Glu	Ser	Leu	His 140	Tyr	Glu	Gly	Arg
	45	Ala 145	Val	Asp	Ile	Thr	Thr 150	Ser	Asp	Arg	Asp	Arg 155	Ser	Lys	Tyr	Gly	Met 160
	50	Leu	Ala	Arg	Leu	Ala 165	Val	Glu	Ala	Gly	Phe 170	Asp	Trp	Val	Tyr	Tyr 175	Glu
	50	Ser	Lys	Ala	His 180	Ile	His	Cys	Ser	Val 185	Lys	Ala	Glu	Asn	Ser 190	Val	Ala
	55	Ala	Lys	Ser 195	Gly	Gly	Сув	Phe	Pro 200	Gly	Ser	Ala	Thr	Val 205	His	Leu	Glu
		Gln	Gly 210		Thr	Lys	Leu	Val 215	Lys	Asp	Leu	Ser	Pro 220	Gly	Asp	Arg	Val
	60	Leu	Ala	Ala	Asp	Asp	Gln	Gly	Arg	Leu	Leu	Tyr	Ser	Asp	Phe	Leu	Thr

		225					230					235					240
	5	Phe	Leu	Asp	Arg	Asp 245	Asp	Gly	Ala	Lys	Lys 250	Val	Phe	Tyr	Val	Ile 255	Glu
	3	Thr	Arg	Glu	Pro 260	Arg	Glu	Arg	Leu	Leu 265	Leu	Thr	Ala	Ala	His 270	Leu	Leu
	10	Phe	Val	Ala 275	Pro	His	Asn	Asp	Ser 280	Ala	Thr	Gly	Glu	Pro 285	Glu	Ala	Ser
		Ser	Gly 290	Ser	Gly	Pro	Pro	Ser 295	Gly	Gly	Ala	Leu	Gly 300	Pro	Arg	Ala	Leu
	15	Phe 305	Ala	Ser	Arg	Val	Arg 310	Pro	Gly	Gln	Arg	Val 315	Tyr	Val	Val	Ala	Glu 320
	20	Arg	Asp	Gly	Asp	Arg 325	Arg	Leu	Leu	Pro	Ala 330	Ala	Val	His	Ser	Val 335	Thr
	20	Leu	Ser	Glu	Glu 340	Ala	Ala	Gly	Ala	Tyr 345	Ala	Pro	Leu	Thr	Ala 350	Gln	Gly
	25	Thr	Ile	Leu 355	Ile	Asn	Arg	Val	Leu 360	Ala	Ser	Сув	Tyr	Ala 365	Val	Ile	Glu
Will High Reg		Glu	His 370	Ser	Trp	Ala	His	Arg 375	Ala	Phe	Ala	Pro	Phe 380	Arg	Leu	Ala	His
	30	Ala 385		Leu	Ala	Ala	Leu 390	Ala	Pro	Ala	Arg	Thr 395	Asp	Arg	Gly	Gly	Asp 400
	35	Ser	Gly	Gly	Gly	Asp 405	Arg	Gly	Gly	Gly	Gly 410	Gly	Arg	Val	Ala	Leu 415	Thr
	35 40	Ala	Pro	Gly	Ala 420		Asp	Ala	Pro	Gly 425	Ala	Gly	Ala	Thr	Ala 430	Gly	Ile
	40	His	Trp	Tyr 435	Ser	Gln	Leu	Leu	Tyr 440		Ile	Gly	Thr	Trp 445		Leu	Asp
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	50			(i)	(A	ENCE  ) LE  ) TY  ) TC	NGTH	: 41 amin	.1 am	ino id	s: acid	ls					
	55		(	(ii)	MOLE	CULE	TYF	E: p	rote	in							
			(	(xi)	SEQU	ENCE	DES	CRIE	MOIT	: SE	EQ II	NO:	16:				
	60	Met	: Ser	Pro	Ala	Arç	Let	ı Arg	, Pro	Arg	J Let		Phe	e Cys	s Leu	ı Val	

		Thr	Leu	Val	Val 340	Glu	Asp	Val	Val	Ala 345	Ser	Cys	Phe	Ala	Ala 350	Val	Ala
	5	Asp	His	His 355	Leu	Ala	Gln	Leu	Ala 360	Phe	Trp	Pro	Leu	Arg 365	Leu	Phe	His
	10	Ser	Leu 370	Ala	Trp	Gly	Ser	Trp 375	Thr	Pro	Gly	Glu	Gly 380	Val	His	Trp	Tyr
•	10	Pro 385	Gln	Leu	Leu	Tyr	Arg 390	Leu	Gly	Arg	Leu	Leu 395	Leu	Glu	Glu	Gly	Ser 400
	15	Phe	His	Pro	Leu	Gly 405	Met	Ser	Gly	Ala	Gly 410	Ser					
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	20		ı	(i) §		LEI TY	NGTH PE: a	RACTI : 396 amino GY: 1	am:	ino a id		3					
	25		( =	ii) M	MOLE	CULE	TYP	E: p	rote	in							
			•	•	-			CRIP									
	30	Met 1	Ala	Leu	Leu	Thr 5	Asn	Leu	Leu	Pro	Leu 10	Cys	Cys	Leu	Ala	Leu 15	Leu
		Ala	Leu	Pro	Ala 20	Gln	Ser	Cys	Gly	Pro 25	Gly	Arg	Gly	Pro	Val 30	Gly	Arg
	35	Arg	Arg	Tyr 35		Arg	Lys	Gln	Leu 40	Val	Pro	Leu	Leu	Tyr 45	Lys	Gln	Phe
		Val	Pro 50	Gly	Val	Pro	Glu	Arg 55	Thr	Leu	Gly	Ala	Ser 60		Pro	Ala	Glu
	40	Gly 65	-	Val	Ala	Arg	Gly 70		Glu	Arg	Phe	Arg 75		Leu	Val	Pro	Asn 80
	45	Tyr	Asn	Pro	Asp	Ile 85		Phe	Lys	Asp	Glu 90		Asn	Ser	Gly	Ala 95	Asp
	<b>4</b> 5	Arg	Leu	Met	Thr 100		Arg	Cys	Lys	Glu 105		Val	Asn	n Ala	Leu 110		Ile
	50	Ala	. Val	Met 115		Met	Trp	) Pro	Gly 120		Arg	Leu	Arg	y Val 125		Glu	Gly
		Trp	Asp 130		ı Asp	Gly	7 His	His 135		Gln	Asp	Ser	Leu 140		туг	Glu	Gly
	55	Arg 145		Leu	ı Asp	Ile	Thr 150		Ser	Asp	Arg	Asp 155		g Asr	ı Lys	Tyr	Gly 160
	60	Leu	ı Leu	. Ala	a Arg	J Let 165		a Val	. Glu	ı Ala	Gly 170		e As <u>r</u>	o Trp	Val	175 175	Tyr

	ı	Glu	Ser	Arg	Asn 180	His	Val	His	Val	Ser 185	Val	ГÀЗ	Ala	Asp	Asn 190	Ser	Leu
5		Ala	Val	Arg 195	Ala	Gly	Gly	Cys	Phe 200	Pro	Gly	Asn	Ala	Thr 205	Val	Arg	Leu
		Trp	Ser 210	Gly	Glu	Arg	Lys	Gly 215	Leu	Arg	Glu	Leu	His 220	Arg	Gly	Asp	Trp
10		Val 225	Leu	Ala	Ala	Asp	Ala 230	Ser	Gly	Arg	Val	Val 235	Pro	Thr	Pro	Val	Leu 240
1.4		Leu	Phe	Leu	Asp	Arg 245	Asp	Leu	Gln	Arg	Arg 250	Ala	Ser	Phe	Val	Ala 255	Val
15	,	Glu	Thr	Glu	Trp 260	Pro	Pro	Arg	Lys	Leu 265	Leu	Leu	Thr	Pro	Trp 270	His	Leu
20	)	Val	Phe	Ala 275	Ala	Arg	Gly	Pro	Ala 280	Pro	Ala	Pro	Gly	Asp 285	Phe	Ala	Pro
Constitution of the consti		Val	Phe 290	Ala	Arg	Arg	Leu	Arg 295	Ala	Gly	Asp	Ser	Val 300	Leu	Ala	Pro	Gly
≒ 2: ∏⊔	5	Gly 305		Ala	Leu	Arg	Pro 310	Ala	Arg	Val	Ala	Arg 315	Val	Ala	Arg	Glu	Glu 320
<b>1 1 3 3</b>	0	Ala	Val	Gly	Val	Phe 325	Ala	Pro	Leu	Thr	Ala 330	His	Gly	Thr	Leu	Leu 335	Val
	U	Asn	Asp	Val	Leu 340	Ala	Ser	Cys	Tyr	Ala 345		Leu	Glu	Ser	His 350	Gln	Trp
⊨ ⊨ 3	5	Ala	His	Arg 355		Phe	Ala	Pro	Leu 360		Leu	Leu	His	Ala 365	Leu	Gly	Ala
1 3 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1		Leu	Leu 370		Gly	Gly	Ala	Val 375		Pro	Thr	Gly	Met 380	His	Trp	Tyr	Ser
<b>=</b> 4	0	Arg		Leu	Tyr	Arg	1 Leu 390		. Glu	Glu	Leu	1 Leu 395		•			
4	5	(2)	INF	ORMA	MOITA	I FOR	SEÇ	) ID	NO: 1	.8:							
				(i)	( <i>I</i>	A) LE	NGTI PE:	I: 41 amir	.6 an		: acio	ds					
5	0			(ii)		) TO											
											EQ II	D NO	:18:				
	55					g Lei						e Ala		ı Le	ı Cy:	s Phe	e Ile 5
(	50	Se	r Lei	ı Leı	ı Let 20		r Pro	о Су:	s Gl	y Let 2!		a Cy	s Gl	y Pr	o Gl		g Gly

	Tyr	Gly	Lys 35	Arg	Arg	His	Pro	Lys 40	Lys	Leu	Thr	Pro	Leu 45	Ala	Tyr	Lys
5	Gln	Phe 50	Ile	Pro	Asn	Val	Ala 55	Glu	Lys	Thr	Leu	Gly 60	Ala	Ser	Gly	Lys
10	Tyr 65	Glu	Gly	Lys	Ile	Thr 70	Arg	Asn	Ser	Glu	Arg 75	Phe	Lys	Glu	Leu	Ile 80
10	Pro	Asn	Tyr	Asn	Pro 85	Asp	Ile	Ile	Phe	Lys 90	Asp	Glu	Glu	Asn	Thr 95	Asn
15	Ala	Asp	Arg	Leu 100	Met	Thr	Lys	Arg	Cys 105	Lys	Asp	Lys	Leu	Asn 110	Ser	Leu
	Ala	Ile	Ser 115	Val	Met	Asn	His	Trp 120	Pro	Gly	Val	Lys	Leu 125	Arg	Val	Thr
20	Glu	Gly 130	Trp	Asp	Glu	Asp	Gly 135	His	His	Leu	Glu	Glu 140	Ser	Leu	His	Tyr
25	Glu 145	Gly	Arg	Ala	Val	Asp 150	Ile	Thr	Thr	Ser	Asp 155	Arg	Asp	Lys	Ser	Lys 160
25 30	Tyr	Gly	Met	Leu	Ser 165	Arg	Leu	Ala	Val	Glu 170	Ala	Gly	Phe	Asp	Trp 175	Val
30	Tyr	Tyr	Glu	Ser 180	Lys	Ala	His	Ile	His 185	Cys	Ser	Val	Lys	Ala 190	Glu	Asn
	Ser	Val	Ala 195	Ala	Lys	Ser	Gly	Gly 200	Cys	Phe	Pro	Gly	Ser 205	Gly	Thr	Val
35	Thr	Leu 210	Gly	Asp	Gly	Thr	Arg 215	Lys	Pro	Ile	Lys	Asp 220	Leu	Lys	Val	Gly
40	Asp 225	Arg	Val	Leu	Ala	Ala 230	Asp	Glu	Lys	Gly	Asn 235	Val	Leu	Ile	Ser	Asp 240
	Phe	Ile	Met	Phe	Ile 245	Asp	His	Asp	Pro	Thr 250	Thr	Arg	Arg	Gln	Phe 255	Ile
45	Val	Ile	Glu	Thr 260	Ser	Glu	Pro	Phe	Thr 265	Lys	Leu	Thr	Leu	Thr 270	Ala	Ala
	His	Leu	Val 275	Phe	Val	Gly	Asn	Ser 280	Ser	Ala	Ala	Ser	Gly 285	Ile	Thr	Ala
50	Thr	Phe 290		Ser	Asn	Val	Lys 295		Gly	Asp	Thr	Val 300	Leu	Val	Trp	Glu
55	Asp 305		Cys	Glu	Ser	Leu 310	_	Ser	Val	Thr	Val 315	_	Arg	Ile	Tyr	Thr 320
	Glu	Glu	His	Glu	Gly 325		Phe	Ala	Pro	Val 330		Ala	His	Gly	Thr 335	Ile
60	Ile	Val	Asp	Gln 340		Leu	Ala	Ser	Cys 345		Ala	Val	Ile	Glu 350		His

		Lys	Trp	Ala 355	His	Trp	Ala	Phe	Ala 360	Pro	Val	Arg	Leu	Cys 365	His	Lys	Leu		
	5	Met	Thr 370	Trp	Leu	Phe	Pro	Ala 375	Arg	Glu	Ser	Asn	Val 380	Asn	Phe	Gln	Glu		
	10	Asp 385	Gly	Ile	His	Trp	Tyr 390	Ser	Asn	Met	Leu	Phe 395	His	Ile	Gly	Ser	Trp 400		
	10	Leu	Leu	Asp	Arg	Asp 405	Ser	Phe	His	Pro	Leu 410	Gly	Ile	Leu	His	Leu 415	Ser		
	15	(2)	INFO	ORMAT	rion	FOR	SEQ	ID 1	NO:19	∍:									
	20		(i)	( <i>I</i> (I	A) LI 3) T C) S	CE CH ENGTH YPE: TRANI OPOLO	H: 14 nucl	116 ] leic ESS:	oase acio botl	pai:	cs								
			(ii)	MOI	LECU	LE T	PE:	cDN	A										
The state of the s	25		(ix)		A) N	E: AME/I OCATI			1413										
	30		(xi)	) SE	QUEN	CE DI	ESCR:	IPTI(	ON:	SEQ :	ID N	0:19	:						
	35					AGC Ser 5										_	_	4	48
Table 1	40					GGA Gly											_	:	96
	40					AGC Ser												1	44
	45			Met		CAC His												1	92
	50		-			GCC Ala		Leu										2	40
	55					TGC Cys 85						Leu						2	88
	60					CCG Pro					Gln					Leu		3	36

	GAG Glu	TAC Tyr	ACG Thr 115	AAC Asn	AGC Ser	GCC Ala	TCC Ser	GGA Gly 120	CCT Pro	CTG Leu	GAG Glu	GGT Gly	GTG Val 125	ATC Ile	CGT Arg	CGG Arg	384
5	_	TCG Ser 130	CCC Pro	AAA Lys	TTC Phe	AAG Lys	GAC Asp 135	CTC Leu	GTG Val	CCC Pro	AAC Asn	TAC Tyr 140	AAC Asn	AGG Arg	GAC Asp	ATC Ile	432
10	CTT Leu 145	TTC Phe	CGT Arg	GAC Asp	GAG Glu	GAA Glu 150	GGC Gly	ACC Thr	GGA Gly	GCG Ala	GAT Asp 155	GGC Gly	TTG Leu	ATG Met	AGC Ser	AAG Lys 160	480
15	CGC Arg	TGC Cys	AAG Lys	GAG Glu	AAG Lys 165	CTA Leu	AAC Asn	GTG Val	CTG Leu	GCC Ala 170	TAC Tyr	TCG Ser	GTG Val	ATG Met	AAC Asn 175	GAA Glu	528
20	TGG Trp	CCC Pro	GGC Gly	ATC Ile 180	CGG Arg	CTG Leu	CTG Leu	GTC Val	ACC Thr 185	GAG Glu	AGC Ser	TGG Trp	GAC Asp	GAG Glu 190	GAC Asp	TAC Tyr	576
	CAT His	CAC His	GGC Gly 195	CAG Gln	GAG Glu	TCG Ser	CTC Leu	CAC His 200	TAC Tyr	GAG Glu	GGC Gly	CGA Arg	GCG Ala 205	GTG Val	ACC Thr	ATT Ile	624
25	GCC Ala	ACC Thr 210	TCC Ser	GAT Asp	CGC Arg	GAC Asp	CAG Gln 215	TCC Ser	AAA Lys	TAC Tyr	GGC Gly	ATG Met 220	CTC Leu	GCT Ala	CGC Arg	CTG Leu	672
30	GCC Ala 225	GTC Val	GAG Glu	GCT Ala	GGA Gly	TTC Phe 230	GAT Asp	TGG Trp	GTC Val	TCC Ser	TAC Tyr 235	GTC Val	AGC Ser	AGG Arg	CGC Arg	CAC His 240	720
14 35 15 15 15 15 15 15 15 15 15 15 15 15 15	ATC Ile	TAC Tyr	TGC Cys	TCC Ser	GTC Val 245	AAG Lys	TCA Ser	GAT Asp	TCG Ser	TCG Ser 250	ATC Ile	AGT Ser	TCC Ser	CAC His	GTG Val 255	CAC His	768
<b>1 4</b> 0	GGC Gly	TGC Cys	TTC Phe	ACG Thr 260	CCG Pro	GAG Glu	AGC Ser	ACA Thr	GCG Ala 265	Leu	CTG Leu	GAG Glu	AGT Ser	GGA Gly 270	GTC Val	CGG Arg	816
40	AAG Lys	CCG Pro	CTC Leu 275	Gly	GAG Glu	CTC Leu	TCT Ser	ATC Ile 280	Gly	GAT Asp	CGT Arg	GTT Val	TTG Leu 285	AGC Ser	ATG Met	ACC Thr	864
45	GCC Ala	AAC Asn 290	Gly	CAG Gln	GCC Ala	GTC Val	TAC Tyr 295	Ser	GAA Glu	GTG Val	ATC Ile	CTC Leu 300	Phe	ATG Met	GAC Asp	CGC Arg	912
50	AAC Asn 305	Leu	GAG Glu	CAG Gln	ATG Met	CAA Gln 310	Asn	TTT Phe	GTG Val	Gln	CTG Leu 315	His	ACG Thr	GAC Asp	GGT Gly	GGA Gly 320	960
55	GCA Ala	GTG Val	CTC Leu	ACG Thr	GTG Val 325	Thr	CCG Pro	GCT Ala	CAC His	CTC Lev 330	ı Val	AGC Sei	GTT Val	TGG Trp	Gln 335	CCG Pro	1008
60	GAG Glu	AGC Ser	CAG Glr	AAG Lys 340	Leu	ACG Thr	TTT Phe	GTC Val	F TTT Phe 345	e Ala	G CAT	CGC Arg	C ATC	GAC Glu 350	ı Glu	AAG Lys	1056

	AAC Asn	CAG Gln	GTG Val 355	CTC Leu	GTA Val	CGG Arg	GAT Asp	GTG Val 360	GAG Glu	ACG Thr	GGC Gly	GAG Glu	CTG Leu 365	AGG Arg	CCC Pro	CAG Gln	1104
5	CGA Arg	GTG Val 370	GTC Val	AAG Lys	TTG Leu	GGC Gly	AGT Ser 375	GTG Val	CGC Arg	AGT Ser	AAG Lys	GGC Gly 380	GTG Val	GTC Val	GCG Ala	CCG Pro	1152
10	CTG Leu 385	ACC Thr	CGC Arg	GAG Glu	GGC Gly	ACC Thr 390	ATT Ile	GTG Val	GTC Val	AAC Asn	TCG Ser 395	GTG Val	GCC Ala	GCC Ala	AGT Ser	TGC Cys 400	1200
15	TAT Tyr	GCG Ala	GTG Val	ATC Ile	AAC Asn 405	AGT Ser	CAG Gln	TCG Ser	CTG Leu	GCC Ala 410	CAC His	TGG Trp	GGA Gly	CTG Leu	GCT Ala 415	CCC Pro	1248
20	ATG Met	CGC Arg	CTG Leu	CTG Leu 420	TCC Ser	ACG Thr	CTG Leu	GAG Glu	GCG Ala 425	TGG Trp	CTG Leu	CCC Pro	GCC Ala	AAG Lys 430	GAG Glu	CAG Gln	1296
20	TTG Leu	CAC His	AGT Ser 435	TCG Ser	CCG Pro	AAG Lys	GTG Val	GTG Val 440	AGC Ser	TCG Ser	GCG Ala	CAG Gln	CAG Gln 445	CAG Gln	AAT Asn	GGC Gly	1344
25 25	ATC Ile	CAT His 450	Trp	TAT Tyr	GCC Ala	AAT Asn	GCG Ala 455	CTC Leu	TAC Tyr	AAG Lys	GTC Val	AAG Lys 460	Asp	TAC Tyr	GTG Val	CTG Leu	1392
□ □ □ □ □			AGC Ser														1416
35	(2)	(	ii)	SEQU (A (E (C MOLE	ENCE ) LE ) TY ) TO	CHA NGTH PE: POLO	RACT : 47 amin GY:	ERIS 1 am o ac line	TICS ino id ar	s: acid		:20:					
45	Met 1		) Asn	His	s Ser		· Val	Pro	Trp	Ala 10		Ala	a Ala	a Ser	. Val	Thr	
50	Cys	Leu	ı Ser	: Let 20		/ Сув	Glr.	ı Met	25		n Phe	e Glr	n Phe	e Glr 30	n Phe	e Gln	
30	Leu	ı Glr	ı Ile 35		g Sei	: Glu	ı Lev	His		ı Arg	J Lys	s Pro	Ala 4!		g Arg	y Thr	
55	Glr	1 Th:		: Ar	g His	s Ile	e Ala		s Th	r Glı	ı Arç	g Cy:		ı Se:	r Arg	J Leu	
	Thi 65		r Lei	ı Va	l Ala	a Lei 70		ı Lei	ı Ile	e Va	l Lei 7		o Me	t Vai	l Phe	e Ser 80	
60	Pro	o Ala	a His	s Se	r Cy:	s Gly	y Pro	o Gl	y Ar	g Gl	y Le	u Gl	y Ar	g Hi	s Ar	g Ala	

					85					90					95	
5	Arg	Asn	Leu	Tyr 100	Pro	Leu	Val	Leu	Lys 105	Gln	Thr	Ile	Pro	Asn 110	Leu	Ser
J	Glu	Tyr	Thr 115	Asn	Ser	Ala	Ser	Gly 120	Pro	Leu	Glu	Gly	Val 125	Ile	Arg	Arg
10	Asp	Ser 130	Pro	Lys	Phe	Lys	Asp 135	Leu	Val	Pro	Asn	Tyr 140	Asn	Arg	Asp	Ile
	Leu 145	Phe	Arg	Asp	Glu	Glu 150	Gly	Thr	Gly	Ala	Asp 155	Gly	Leu	Met	Ser	Lys 160
15	Arg	Cys	Lys	Glu	Lys 165	Leu	Asn	۷al	Leu	Ala 170	Tyr	Ser	Val	Met	Asn 175	Glu
20	Trp	Pro	Gly	Ile 180	Arg	Leu	Leu	Val	Thr 185	Glu	Ser	Trp	Asp	Glu 190	Asp	Tyr
enso,	His	His	Gly 195	Gln	Glu	Ser	Leu	His 200	Tyr	Glu	Gly	Arg	Ala 205	Val	Thr	Ile
₽ ₩25	Ala	Thr 210	Ser	Asp	Arg	Asp	Gln 215	Ser	Lys	Tyr	Gly	Met 220	Leu	Ala	Arg	Leu
	Ala 225	Val	Glu	Ala	Gly	Phe 230	Asp	Trp	Val	Ser	Tyr 235	Val	Ser	Arg	Arg	His 240
30	Ile	Tyr	Cys	Ser	Val 245	Lys	Ser	Asp	Ser	Ser 250	Ile	Ser	Ser	His	Val 255	His
± ± 35	Gly	Cys	Phe	Thr 260	Pro	Glu	Ser	Thr	Ala 265	Leu	Leu	Glu	Ser	Gly 270	Val	Arg
35	Lys	Pro	Leu 275	_	Glu	Leu	Ser	Ile 280	Gly	Asp	Arg	Val	Leu 285	Ser	Met	Thr
40		290	_		Ala		295					300				
	Asn 305		Glu	Gln	Met	Gln 310	Asn	Phe	۷al	Gln	Leu 315	His	Thr	Asp	Gly	Gly 320
45	Ala	۷al	Leu	Thr	Val 325	Thr	Pro	Ala	His	Leu 330		Ser	Val	Trp	Gln 335	Pro
50	Glu	Ser	Gln	Lys 340	Leu	Thr	Phe	Val	Phe 345		His	Arg	Ile	Glu 350	Glu	Lys
	Asn	Gln	Val 355		Val	Arg	Asp	Val 360		Thr	Gly	Glu	Leu 365		Pro	Gln
55	Arg	Val 370		Lys	Leu	Gly	Ser 375		Arg	Ser	Lys	Gly 380		Val	Ala	Pro
	Leu 385		Arg	Glu	Gly	Thr 390		· Val	Val	Asn	Ser 395		Ala	Ala	Ser	Cys 400
60	Tvr	Ala	Val	Ile	Asn	Ser	Gln	Ser	Leu	Ala	His	Trn	Glv	Leu	Ala	Pro

					405					4	10					415	5		
_	Met	Arg	Leu	Leu 420	Ser	Thr	Leu	Glu	Ala 425		rp I	Leu	Pro	Ala	Lys 430	Glı	ı G	ln	
5	Leu	His	Ser 435	Ser	Pro	Lys	Val	Val 440	Sei	c S	er A	Ala	Gln	Gln 445	Gln	Ası	ı G	ly	
10	Ile	His 450	Trp	Tyr	Ala	Asn	Ala 455	Leu	Туз	r L	ys 1		Lys 460	Asp	Tyr	Va:	l L	eu	
	Pro 465	Gln	Ser	Trp	_	His 470	Asp												
15	(2)	INFO	RMAT	CION	FOR	SEQ	ID 1	NO:2	1:										
20		(i)	( <i>P</i>	A) LE B) TY	E CH NGTH PE:	: 22 ami	21 at 10 at	mino cid		ids									
i.		(ii)	MOI	ECUI	E TY	PE:	pep	tide											
25		(v)	FRA	AGMEN	IT TY	PE:	int	erna	1										
		(xi)	SEÇ	QUENC	CE DE	SCR	IPTI	: NC	SEQ	ID	) NO	:21:							
30		Cys 1	Gl <sub>y</sub>	7 Pro	Gly	Arg 5	g Gl	y Xa	a G	ly	Xaa	Arg 10	Arg	g Hi	s Pr	o L	ys	Lys 15	Leu
35		Thr	Pro	Let	a Ala 20	ту:	r Ly	s Gl	n P	he	Ile 25	Pro	Ası	n Va	l Al	a G 3		Lys	Thr
		Leu	ı Gly	7 Ala 35	a Ser	Gl;	y Ar	д Ту	r G 4		Gly	Lys	: Ile	e Xa	a Ar 45		sn	Ser	Glu
40		Arg	Phe 50	e Lys	s Glu	ı Le	u Th	r Pr 55		sn	Tyr	Asr	n Pro	o As 60		e I	le	Phe	Lys
		Asp 65	Glı	ı Glu	ı Ası	ı Th	r Gl 70	y Al	a A	sp	Arg	Leu	75	t Th	r Gl	n A	rg	Cys	80 Lys
45		Asp	ь Гу	s Lei	ı Ası	1 Xa 85	a Le	u Al	a I	le	Ser	Va] 90	. Me	t As	n Xa	аТ	rp	Pro 95	Gly
50		Va]	l Xaa	a Lei	u Arg	-	l Th	r Gl	.u G	1y	Trp 105		Gl:	u As	p Gl		is 10	His	Xaa
		Glı	ı Glı	u Se:	r Lei 5	ı Hi	в Ту	r Gl		lу 20	Arg	Ala	a Va	l As	p Il 12		hr	Thr	Ser
55		Asp	2 Ar		p Xaa	a Se	r Ly	s Ty 13	_	ly	Xaa	. Le	ı Xa	a Ar 14		u A	la	Val	Glu
		Ala 145		y Ph	e Asj	o Tr	р Va 15		r T	yr	Glu	Sei	с Ly 15	_	a Hi	s I	le	His	Cys 160
60		Sei	r Va	l Ly	s Ala	a Gl	u As	n Se	er V	al	Ala	Ala	а Ьу	s Se	r Gl	.у С	ly	Cys	Phe

						165					170					175	
5		Pro	Gly	Ser	Ala 180	Xaa	Val	Xaa	Leu	Xaa 185	Xaa	Gly	Gly	Xaa	Lys 190	Xaa	Val
5		Lys	Asp	Leu 195	Xaa	Pro	Gly	Asp	Xaa 200	Val	Leu	Ala	Ala	Asp 205	Xaa	Xaa	Gly
10		Xaa	Leu 210	Xaa	Xaa	Ser	Asp	Phe 215	Xaa	Xaa	Phe	Xaa	Asp 220	Arg			
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20		(ii)	MOLE	ECULI	E TYI	PE: 1	pept:	ide									
		(v)	FRAC	GMEN'	г түі	PE: :	inte	rnal									
3 25		(xi)	SEQU	JENC	E DES	SCRI	PTIO	M: S	EQ II	ои с	:22:						
25		Cys 1	Gly	Pro	Gly	Arg 5	Gly	Xaa	Xaa	Xaa	Arg 10	Arg	Xaa	Xaa	Xaa	Pro 15	Lys
		Xaa	Leu	Xaa	Pro 20	Leu	Xaa	Tyr	Lys	Gln 25	Phe	Xaa	Pro	Xaa	Xaa 30	Xaa	Glu
35		Xaa	Thr	Leu 35	Gly	Ala	Ser	Gly	Xaa 40	Xaa	Glu	Gly	Xaa	Xaa 45	Xaa	Arg	Xaa
A CONTROL OF THE CONT		Ser	Glu 50	Arg	Phe	Xaa	Xaa	Leu 55	Thr	Pro	Asn	Tyr	Asn 60	Pro	Asp	Ile	Ile
40		Phe 65	Lys	Asp	Glu	Glu	Asn 70	Xaa	Gly	Ala	Asp	Arg 75	Leu	Met	Thr	Xaa	Arg 80
45		Cys	Lys	Xaa	Xaa	Xaa 85	Asn	Xaa	Leu	Ala	Ile 90	Ser	Val	Met	Asn	Xaa 95	Trp
		Pro	Gly	Val	Xaa 100	Leu	Arg	Val	Thr	Glu 105		Xaa	Asp	Glu	Asp 110		His
50		His	Xaa	Хаа 115		Ser	Leu	His	Tyr 120		Gly	Arg	Ala	Xaa 125		Ile	Thr
		Thr	Ser 130		Arg	Asp	Xaa	Xaa 135		Tyr	Gly	Xaa	Leu 140		. Arg	Leu	Ala
55		Val 145		Ala	Gly	Phe	150		Val	Tyr	Tyr	Glu 155		· Xaa	. Xaa	His	Xaa 160
60		His	Xaa	. Ser	· Val	Lys 165		. Xaa	L								

- 98 -

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#### We claim:

- 1. A method for modulating the immune function of an animal comprising administering to the animal a therapeutic amount of a *hedgehog* or *ptc* therapeutic.
- 5 2. A method for suppressing the immune system of an animal comprising contacting the cells with an effective amount of a *hedgehog* protein, or agonist thereof.
  - 3. A method for enhancing the immune system of an animal comprising administering to a immunostimulatory amount of a *hedgehog* antagonist.
- 4. The method of any of claims 1, wherein the *hedgehog* therapeutic is a polypeptide which includes a hedgehog amino acid sequence which is identical or homologous to an amino acid sequence of any one of SEQ ID Nos. 10-18.
  - 5. The method of claim 4, wherein the hedgehog amino acid sequence is sufficient for specific binding of the polypeptide to a *patched* protein.
  - 6. The method of claim 4, wherein the hedgehog amino acid sequence is at least 80 percent identical to an amino acid sequence of any one of SEQ ID Nos. 10-18.
  - 7. The method of claim 4, wherein the hedgehog amino acid sequence is encodable by a nucleic acid which hybridizes under stringent conditions to any one of SEQ ID Nos. 1-9.
  - 8. The method of claim 4, wherein the hedgehog amino acid sequence is of a vertebrate hedgehog protein.
  - 9. The method of claim 4, wherein the polypeptide includes at least a 50 amino acid extracellular portion of a vertebrate hedgehog protein.
  - 10. The method of claim 4, wherein the polypeptide includes at least an extracellular portion of a vertebrate hedgehog protein corresponding to residues 24-194 of SEQ ID No:15.
- 11. The method of claim 4, wherein the hedgehog polypeptide is modified with one or more lipophilic moieties.
  - 12. The method of claim 11, wherein the hedgehog polypeptide is modified with one or more sterol moieties.
  - 13. The method of claim 12, wherein the sterol moiety is cholesterol.
- 14. The method of claim 11, wherein the hedgehog polypeptide is modified with one or more fatty acid moieties.

- 15. The method of claim 14, wherein each fatty acid moiety is independently selected from the group consisting of myristoyl, palmitoyl, stearoyl, and arachidoyl.
- 16. The method of claim 11, wherein the hedgehog polypeptide is modified with one or more aromatic hydrocarbons.
- 5 17. The method of claim 16, wherein each aromatic hydrocarbon is ondependently selected from the group consisting of benzene, perylene, phenanthrene, anthracene, naphthalene, pyrene, chrysene, and naphthacene.
  - 18. The method of claim 11, wherein the hedgehog polypeptide is modified one or more times with a C7 C30 alkyl or cycloalkyl.
- 10 19. The method of claim 1, wherein the ptc therapeutic is a small organic molecule.
  - 20. The method of claim 19, wherein the binding of the ptc therapeutic to *patched* results in upor down-regulation of patched and/or gli expression.
  - 21. The method of claim 1, wherein the *ptc* therapeutic binds to *patched* and mimics *hedgehog*-mediated *patched* signal transduction.
  - 22. The method of claim 19, wherein the ptc therapeutic is an inhibitor of protein kinase A.
  - 23. The method of claim 22, wherein the PKA inhibitor is a 5-isoquinolinesulfonamide
  - 24. The method of claim 22, wherein the PKA inhibitor is represented in the general formula:

wherein,

R<sub>1</sub> and R<sub>2</sub> each can independently represent hydrogen, and as valence and stability permit a lower alkyl, a lower alkynyl, a carbonyl (such as a carboxyl, an ester, a formate, or a ketone), a thiocarbonyl (such as a thioester, a thioacetate, or a thioformate), an amino, an acylamino, an amido, a cyano, a nitro, an azido, a sulfate, a sulfonate, a sulfonamido,  $-(CH_2)_m-R_8$ ,  $-(CH_2)_m-OH$ ,  $-(CH_2)_m-OH$ ,  $-(CH_2)_m-OH$ ,  $-(CH_2)_m-OH$ 

 $(CH_2)_m$ - $R_8$ , - $(CH_2)_m$ -SH, - $(CH_2)_m$ -S-lower alkyl, - $(CH_2)_m$ -S-lower alkenyl, - $(CH_2)_m$ - $R_8$ , or

R<sub>1</sub> and R<sub>2</sub> taken together with N form a heterocycle (substituted or unsubstituted);

 $R_3$  is absent or represents one or more substitutions to the isoquinoline ring such as a lower alkyl, a lower alkynyl, a carbonyl (such as a carboxyl, an ester, a formate, or a ketone), a thiocarbonyl (such as a thioester, a thioacetate, or a thioformate), an amino, an acylamino, an amido, a cyano, a nitro, an azido, a sulfate, a sulfonate, a sulfonamido, - $(CH_2)_m$ - $R_8$ , - $(CH_2)_m$ -O-lower alkyl, - $(CH_2)_m$ -O-lower alkenyl, - $(CH_2)_m$ -O-( $CH_2)_m$ -R<sub>8</sub>, - $(CH_2)_m$ -S-lower alkyl, - $(CH_2)_m$ -S-lower alkenyl, - $(CH_2)_m$ -S-( $CH_2)_m$ -R<sub>8</sub>;

10 R<sub>8</sub> represents a substituted or unsubstituted aryl, aralkyl, cycloalkyl, cycloalkenyl, or heterocycle; and

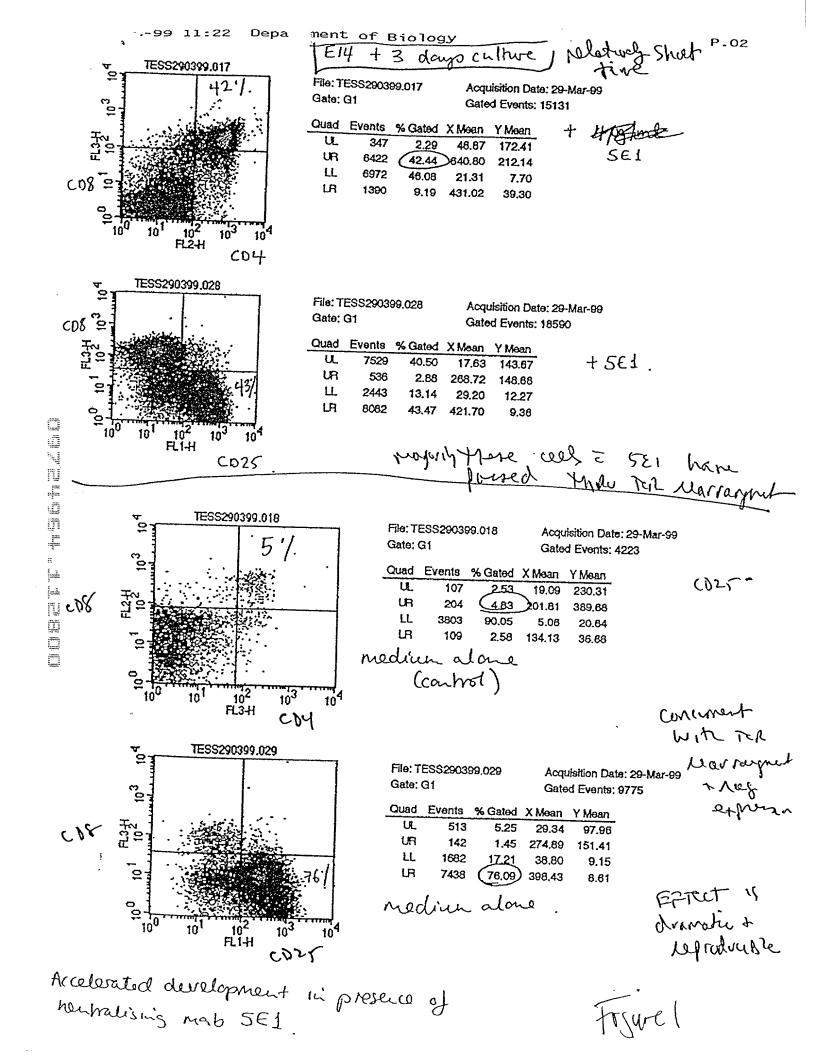
n and m are independently for each occurrence zero or an integer in the range of 1 to 6.

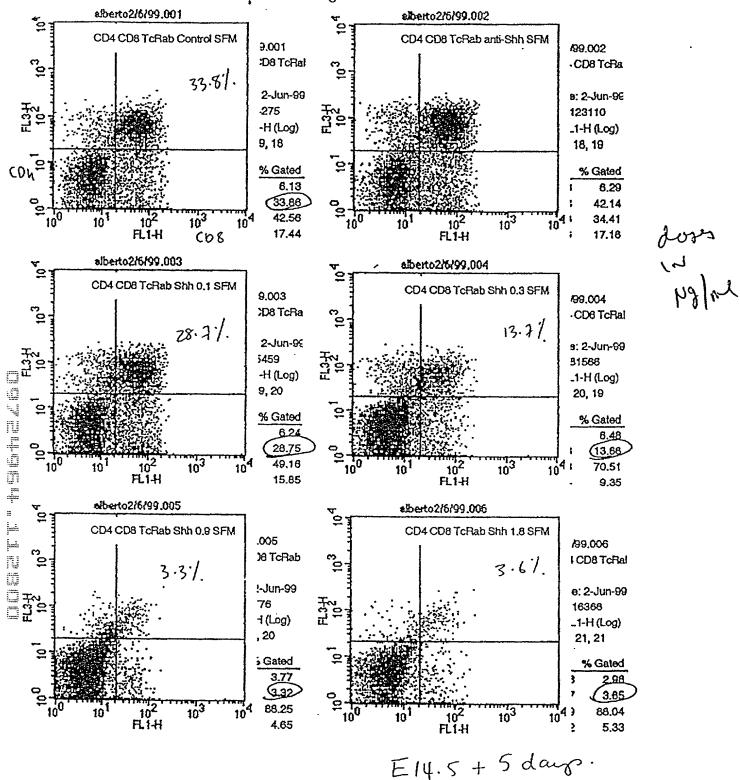
- 25. The method of claim 22, wherein the PKA inhibitor is cyclic AMP analog.
- 26. The method of claim 22, wherein the PKA inhibitor is selected from the group consisting of N-[2-((p-bromocinnamyl)amino)ethyl]-5-isoquinolinesulfonamide, 1-(5-isoquinolinesulfonyl)-2-methylpiperazine, KT5720, 8-bromo-cAMP, dibutyryl-cAMP and PKA Heat Stable Inhibitor isoform α.
- 27. A therapeutic preparation of a small molecule antagonist of *patched*, which *patched* antagonist is provided in a pharmaceutically acceptable carrier and in an amount sufficient to modulate the immune system of an adult human patient.
- 28. A method for modulating T lymphocytes maturation, comprising administering to a patient a gene activation construct which recombines with a genomic *hedgehog* gene of the patient to provide a heterologous transcriptional regulatory sequence operatively linked to a coding sequence of the *hedgehog* gene.
- 25 29. A method of claim 2, wherein suppressing the immune function of an animal comprises inhibiting T lymphocyte maturation.
  - 30. A method of claim 3, wherein enhancing the immune function of an animal comprises stimulating T lymphocyte maturation.

#### Abstract

The present application is directed to the discovery that hedgehog gene products, and signal transduction pathways involving hedgehog, are involved in maturation of T lymphocytes.

Certain aspects of the invention are directed to preparations of hedgehog polypeptides, agonists, antagonists, or other molecules which regulate *patched* or *smoothened* signalling, and their uses as immunomodulatory agents.





FTOC +5 Serum Free Medium

Figure 2

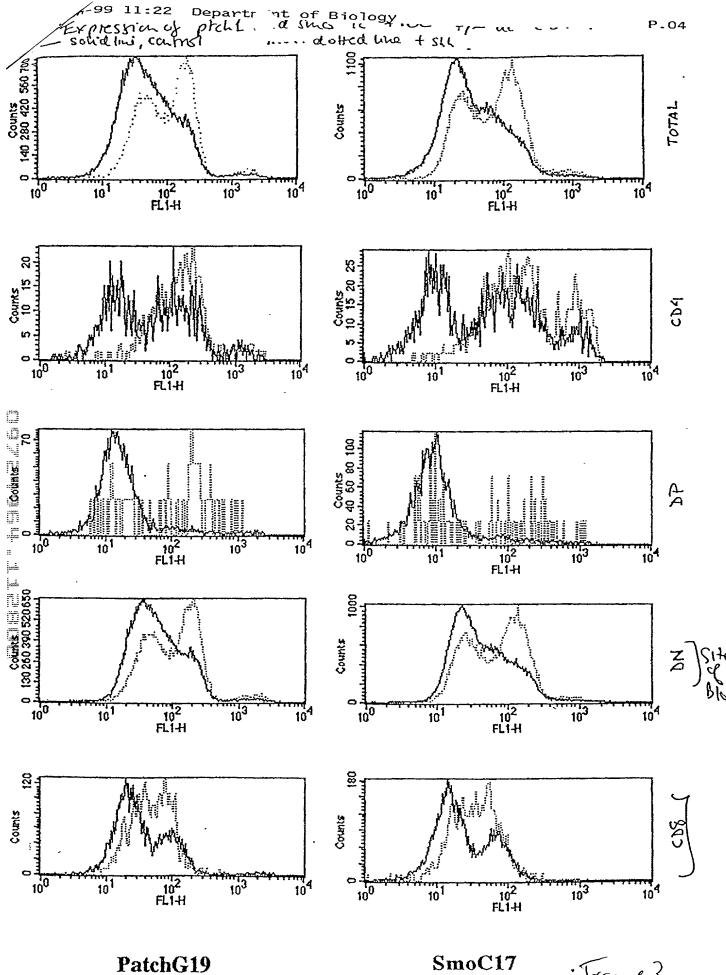


Figure 3

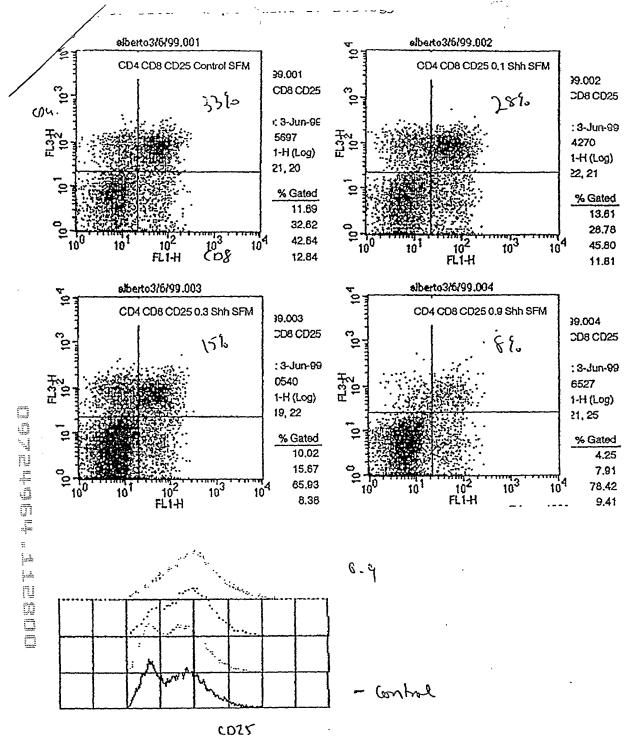


Figure 4

FTOC +6 Serum Free Medium + Shh

### DECLARATION FOR PATENT APPLICATION

Docket Number ONV-080.01

As a below named inventor, I hereby declare that:

My residence, pos	t office address and citi	izenship are as stated below n	ext to my name.			
		inventor (if only one name is for which a patent is sought or	listed below) or an original, first and a the invention entitled:	joint inventor (if plural names	s are listed	below) of
		Methods and Composition	ons for Regulating Lymphocyte	Activity		
the specification o	f which (check one)					
	is attached heretom is attached heretom is attached on income is attached on income.		Application Number or PCT Internal	tional Application Number		_, and was
I hereby state that referred to above.	I have reviewed and un	nderstand the contents of the	above identified specification, includ	ing the claims, as amended by	any amer	ıdment
I acknowledge the	duty to disclose inform	nation, which is material to pa	ntentability as defined in Title 37, Co	ode of Federal Regulation, § 1.	.56.	
	so identified below any		ode, § 119(a)-(d) of any foreign app t or inventor's certificate having a fil			
Prior Foreign Appl	ication(s)				Priority (	Claimed
(Number)		(Country)	(Day/Month/Year Filed)	<del></del>	☐ Yes	□ No
(Number)		(Country)	(Day/Month/Year Filed)		☐ Yes	□ No
(Number)	1 6 1 74 25	• • • • • • • • • • • • • • • • • • • •	, ,	-1:+:(a) 1:-+# h-al		
To be us	benefit under 11tie 35,		of any United States Provisional ap	pheation(s) listed below.		
(Application (Application	Number)	November 30, 1999 (Filing Date)	-			
(Application	Number)	(Filing Date)	-			
which became ava	nilable between the filin		material to patentability as defined in and the national or PCT internation		on.	
- 1	rumovi,	(* <b>g</b>		(+, p, p	<i>S</i> )	
(Application	Number)	(Filing Da	te)	(Status: patented, pend	ling, aband	oned)
46,663; Jason Gis K. Holmes, Reg N Mazzarese, Reg N 32,376; Wolfgang Sharon Webb, Re and Trademark O	h, Reg. No. 42,581; Da No 46,778; Edward J. I No. 42,852; Jim Olesen g E. Stutius, Reg. No. 4 g. No. 46,967; and Joh ffice connected therew	ana Gordon, Reg. No. 44,719; Kelly, Reg. No. 38,936; David A, Reg. No. 47,172; Kevin A. 0,256; Kingsley L. Taft, Reg. n L. Welch, Reg. No. 28,129 ith.	ng No. P-47,326; Kirk A. Damman, I Robert A. Greenberg, Reg. No. 44,1 A. Lane, Reg. No. 39,261; W. Hug Oliver, Reg. No. 42,049; Chinh H. P No. 43,946; Anita Varma, Reg. No. as attorneys/agents to prosecute this	133; David P. Halstead, Reg. No Liepmann, Reg. No. 20,407; ham, Reg. No. 39,329; Philip 43,221; Matthew P. Vincent,	No. 44,735 Robert A C. Swain, Reg. No.	; Jennifer  Reg. No. 36,709;
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		Patent Group Foley, Hoag & Eliot LLP One Post Office Square Boston, Ma. 02109-2170				
and further that th	nese statements were m on 1001 of Title 18 of t	ade with the knowledge that v	e are true and that all statements mad willful false statements and the like s at such willful false statements may j	o made are punishable by fine	or impris	onment, or
		n name, family name): <u>Tessa C</u>		D		
	on, United Kingdom			Date:Citizenship:		

Additional inventors are being named separately numbered sheets attached hereto.